



Pharmacological Promises of Genus *Artemisia* (Asteraceae): a Review

Adil Hussain^{1*}, Muhammad Q. Hayat², Sumaira Sahreen³, Qurrat ul Ain⁴,
and Syed A. I. Bokhari¹

¹Department of Bioinformatics and Biotechnology, International Islamic University,
Islamabad, 44000 Pakistan

²Department of Plant Biotechnology, Atta-ur-Rahman School of Applied Biosciences,
National University of Sciences and Technology, Islamabad 44000, Pakistan

³Botanical Sciences Division, Pakistan Museum of Natural History, Islamabad 44000, Pakistan

⁴Department of Biological Sciences, Karakoram International University,
Gilgit 15100, Pakistan

Abstract: A huge amount of scientific literature is available about plant extracts and their compounds having great pharmacological importance. Although, plant-based medicines have been used since antiquity, but knowledge about their effectiveness on human health is still unclear. *Artemisia* genus belongs to the plant family *Asteraceae*. This genus comprises of about 500 species, which are well known for their medicinal properties. The aim of this review was to provide an insight about recent published scientific literature concerning pharmacological aspects of genus *Artemisia*. The compilation of literature has been done by using references from important databases such as Science Direct, Medicinal and Aromatic Plants Abstracts, PubMed, Chemical Abstracts, Kings American Dispensatory, SciFinder, Research gate, Google Scholar and Phytochemical and Ethnobotanical Databases. In this review, special emphases have been given to the reported chemical compounds and biological activities from different *Artemisia* species. This review emphasis on the plant species from genus *Artemisia* possessing significant phytochemicals holding a broad range of biological actions like antimicrobial, antimalarial, anti-cancerous, antioxidant and anthelmintic activity. Certain crucial drugs have been unveiled from different species of *Artemisia*. One important constituent focused by researchers is artemisinin, which retains potential anti-malarial assets and is attained from *Artemisia annua*. Other groups of phytochemicals like flavonoids, steroids, glycosides, terpenoids, caffeoylquinic acids, acetylenes, coumarins and sterols are also found in this crucial genus. This genus also holds great possibilities for comprehensive scrutiny for other biological activities. The effects of constituents from various species of this genus on other deadly diseases may give better consequences.

Keywords: *Asteraceae*, genus *Artemisia*, phytochemicals, biological activities, bioactive compounds

1. INTRODUCTION

Asteraceae is one of the largest families of angiosperms, which contain 1,600 to 1,700 genera and nearly 24,000 species which are distributed in mostly all parts of the world, but not found in Antarctica [1, 2]. *Artemisia* belongs to the family *Asteraceae* which is pharmacologically one of the crucial polymorphic genera. Plants of this genus are mostly found in the temperate sectors of northern hemisphere, but limited numbers of

species are also found in the southern hemisphere of the world [2-3]. Around five hundred species of herbs and shrubs fall in this genus [4, 5]. This is basically a cosmopolitan [6] and is considered as the diverse and largest genus of the *Asteraceae* family [2, 4, 7]. Different other conducts although have accredited to the genus from 350 to 550 species [3, 5, 8, 9, 10, 11]. Studies by Oberprieler et al. [2] further stretched out the contemporary modifications of subtribe Anthemideae corroborating 522 species to the genus. Since

ancient times plants sources are utilized for food and medicinal purposes and it has been estimated that around 70-95% population of the globe trusts on these plant based remedies for primary care [12].

It has been found that plants have the ability to generate a lot of secondary metabolites which occurs naturally, and may be important in pharmacologically. These essential metabolites may include essential oils, saponins, flavonoids, cyanogenic glycosides, phenols, tannins, phenolic glycosides, unsaturated lactones, and glucosinolates [13, 14, 15] and are predominantly utilized in contradiction of multiple diseases like cancer, malaria, hepatitis, inflammation, and fungal, bacterial, and viral infections. *Artemisia* is expanding medicinal genus among the variety of world cultures [16, 17, 18]. Ethnobotany, economic botany, medicinal importance and phytotherapy of this genus have been reviewed [19] and an extensive work on *Artemisia* unfolded different traditional and medicinal uses of its species [17].

In a lot of investigations, plants and their products have been screened for health purposes, because voluminous number of people has been indulged openly or ramblingly in the traditional usage of different products from plant origin. Among the numerous herbs used in modern medicine, many species of genus *Artemisia* are also included. So in this review, the primary objective is to deliver a brief understanding on different species of the genus *Artemisia* with extraordinary attention on their phytochemistry and pharmacological potentials.

2. PHYTOCHEMICALS AND ANTIOXIDANT ACTIVITY OF *Artemisia* SPECIES

A plant encompasses a lot of extracts and phytochemicals and their utilization could be a milestone in developing therapeutic strategies for the treatment of many diseases. This milestone could be easily achieved if their antimicrobial and antioxidant worth is well understood [20]. *In vitro* studies have shown that, polyphenols with antioxidant potential are plant based compounds

which are pharmacologically active against neurological ailments [21]. These complexes may also play their part as anti-cancerous, antimutagenic and cardio-protective agents because they have free radical hunting capacity [22]. Studies by Ferreira and Stade [23] revealed that polyphenols are chemo preventive representatives because they pull down the level of cholesterol and have preventive cell damage property. Due to these astounding findings, researchers are looking for the replacement of synthetic antioxidants used in foods/medicinal purposes with new naturally occurring harmless antioxidants [21].

The phytochemical and antioxidant activity of different *Artemisia* species have been checked and confirmed by numerous researchers suggesting that the species from genus *Artemisia* are rich in different phytochemical constituents with better antioxidant activity [24–31]. One of the important plants paved with highest attention pharmacologically is *Artemisia annua*, which has been extensively studied and hundreds of secondary metabolites have been discovered and acknowledged so far [32, 33]. Phytochemical analysis of this plant authenticates the presence of sesquiterpenoid along with sesquiterpene lactones and between these constituents artemisinin is highly focused. This compound has endoperoxide sesquiterpene lactone that act as a bioactive drug constituent [34]. In a study, Iqbal et al. [29] confirmed that the ethanolic extract of *A. annua* leaves possess highest amount of phenolics and flavonoids.

Besides *A. annua*, another important plant is *Artemisia amygdalina* having prodigious therapeutic and economic prominence. Its chloroform, ethyl acetate, ethanolic, methanolic, aqueous and crude extracts encloses flavonoid and alkaloids while tri-terpenoids exists only in the ethanolic extract. Tannins are present only in the ethyl acetate extract while saponins are present in ethanolic and aqueous extracts which is absent in methanolic, chloroformic and ethyl acetate extract respectively [28]. Moreover the petroleum ether extracts of *Artemisia amygdalina* contain artemisinin, while other compounds like terpenes,

alkaloids, phenolics, tannins, cardiac glycosides, and steroids in tissue cultured and wild *Artemisia amygdalina* were also observed [24].

Also, with their antimicrobial activity, the methanolic extracts from *Artemisia scoparia* and *Artemisia spicigera* have a vigorous free radical rummaging potential [35]. *Artemisia nilagirica* (Clarke) also contain essential compounds in its extracts like sesquiterpene lactones, exiguaflavone (A and B), benzo furan and mackianin [36]. This plant is anti-leishmanial, antimalarial anthelmintic, antiseptic, astringent, aromatic, anti-inflammatory, appetizer, diuretic, antiasthma, and also used for the treatment of leprosy and skin diseases [37]. Additionally, its oil also has an antioxidant activity [38]. Extracts of *Artemisia nilagirica* also encloses certain other compounds i.e., steroids, terpenoids, flavonoids, saponins, tannins, proteins and essential oil. Due to the existence of these compounds this plant holds a remarkable antibacterial activity [31]. In one study it has been corroborated that the hexane, ethyl acetate, methanol and water extracts from leaves and flower of this plant contain different phytochemicals. Leaves contain saponins, alkaloids, tannins, flavonoid, coumarins, phenols with steroids and flower contain alkaloids, coumarins, saponins, terpenoids, tannins, flavonoids, and phenols which confirms its high therapeutic potential [26].

Another important medicinal plant *Artemisia parviflora* is employed for injuries, cuts and for skin infections [39]. High blood pressure, diabetes, and anthelmintic infections are also treated with this plant [40]. These advantages of *Artemisia parviflora* might be due to the amalgamation of phytochemicals present in it [41]. This could be easily confirmed by a study where various extracts of aerial parts of *Artemisia parviflora* showed the presence of flavonoids, triterpenoids, sterols, tannins, alkaloids, and coumarins that makes it a best nominee for *in vitro* antioxidant activity and phytochemicals extraction [42]. Also, Ahameethunisa and Hopper [43] reported that the methanolic extracts of *A. parviflora* possess greater amount of phenolic compounds as compared to other extracts.

Oils from *Artemisia absinthium* (also called Wormwood) can be used as a cardiac stimulating agent that have the tendency to mend circulation of blood and also employed as a stomach remedy [44]. The whole plant is also used against diseases like tuberculosis, diabetes, and antihypertensive [45]. Studies substantiated that the leaves of this plant contain certain type of compounds which are antimalarial [46], antimicrobial activity [47], antidiabetic activity [48] and antifertility [49]. On the other hand, some traditional uses of *Artemisia absinthium* have also been reported by many researchers [30-50]. Its extracts have shielding influence on hepatic impairment and used as a remedy for gastric pain [50]. Furthermore, the antioxidant activity and occurrence of phenolic compounds and flavonoids in *Artemisia absinthium* have also been reported [30]. Flavonoids are essential because they impede the sensitivity of pain and shows anti-inflammatory properties [51]. These inhibit the enzymes responsible for the synthesis of prostaglandins, that's why they are anti-inflammatory. Other studies for the revelation of actions of the pain killing activity, hepatoprotective and anti-inflammatory potential of *Artemisia annua* and *Artemisia absinthium* were also performed [27]. In one study, the important compound santonin was detected in different *Artemisia* species and its quantitation was done with the help of HPLC-UV. The identification and quantification of santonin was done in the leaves of *A. gmelinni*, *A. terra-albae*, *A. scoparia*, *A. sublesingiana*, *A. foetida*, *A. schrenkiana*, *A. frigida*, *A. absinthium* from Kazakhstan and also in the extracts of leaves of *A. cina* [52].

The methanolic extract of *Artemisia vulgaris*, contain phenolics, flavonoids and sesquiterpenoid type compounds [53] and possess antimicrobial, antitumour, antispasmodic, antiseptic, antimalarial, hepatoprotective and antirheumatic qualities [54].

Aerial parts of *Artemisia vulgaris* contain polysaccharides which are employed to treat numerous diseases and carbohydrates extracted from this plant exhibit several beneficial properties. However the main polysaccharide in

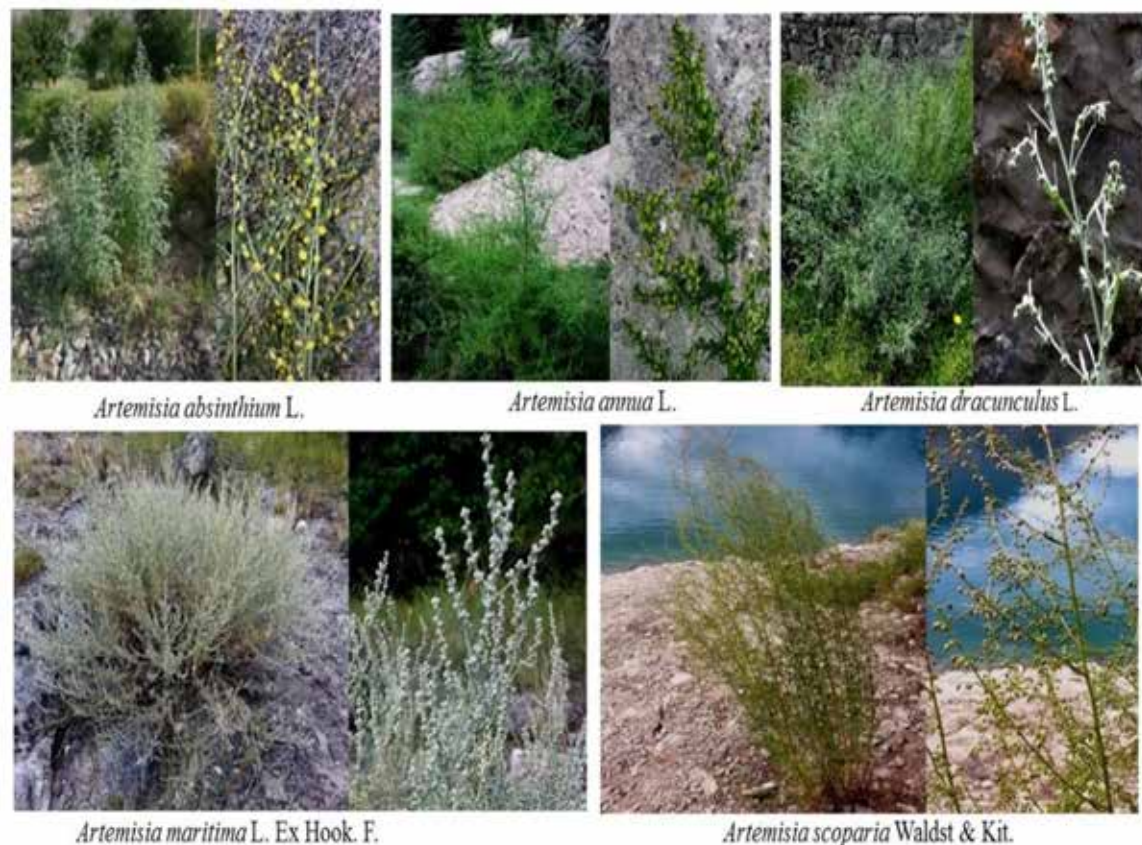


Fig. 1. Plant and synflorescences of *Artemisia* species (Photographs by Adil Hussain).

the infusion is inulin-type fructan [55]. Another revelation showed that the oils from *Artemisia feddei* contain camphor, borneol, cineole, alpha terpineol, chamazulene, alpha thujone, alpha phellandrene, alpha terpinyl acetate, beta caryophyllene, and teroinen-4-ol with antibacterial potential [56]. Similarly, oil from different plant parts (stem, leaf and flower) of *Artemisia chamaemelifolia* Vill. contains compounds like Menthyl acetate, (Z)-nerolidol, 1,8-cineole, yomogi alcohol, and artimesyl acetate while the aerial parts of *Artemisia turcomanica* have compounds like spathulenol, camphor, santolina alcohol and trans beta terpineol [57] and oil of *Artemisia spicigera* have many chemical ingredients like camphor, 1,8-cineole, camphene, p-cymene with alpha and beta thujone showing antibacterial activity [58].

In another investigation, the essential oil of *Artemisia aucheri* was assessed in-vitro and identified compounds includes Linalool, Camphor, Borneol, p-Cymene, Davanone, α -Thujone, β -

Thujone, 1,8-Cineole and α -Pinene having pharmacological importance [59]. An Iranian flora *Artemisia khorassanica* have been assessed and antimalaric compounds like Chrysanthenone, palmitic acid and cis-thujone were confirmed [60]. Studies on *Artemisia indica* Willd revealed that this plant is helpful to lessen chronic fever, hepatobiliary and dyspepsia like illnesses [61]. The stem and leaves are anthelmintic, antiseptic, antispasmodic, expectorant and stomachic [62]. Methanol, ethanol and hydro-methanol extracts from the aerial parts of this plant are rich in flavonoids, sterols carbohydrates, tri-terpenoids, reducing sugars, glycosides and phenolics. The ethanolic extracts do not contain Saponins and tannins, while methanol and hydro-methanol extracts contained Saponins and tannins. Also, with these compounds, methanolic extracts of *A. indica* contain amino acids and alkaloids [25].

In one investigation, phenolic compounds have been detected in *Artemisia judaica* L. where the crude ethanol extract of this plant was tested

on the black bean aphid *Aphis fabae*. The insecticidal activity of the tested crude extract was due to the phenolic compounds present in it, which makes this plant known for the bio-insecticidal activity [63]. Different representative species of the genus *Artemisia* reported globally are illustrated in figure 1.

3. ANTIMALARIAL ACTIVITY OF *Artemisia SPECIES*

Malaria is a severe and highly contagious infection faced by human being [127] and a universal health problem leading to death about 1 million per annum [128]. Malaria in terms of sickness and death is the world's most dangerous parasitic disease [129] and *Plasmodium sp.* protozoa are responsible for malaria, where the *Plasmodium falciparum* is prominent in causing infection. The

transmission can be initiated through transfusion or inoculation of infected blood from person to person. It could also be transferred through the placenta of an infected mother to her unborn child [130]. Now a day, five species of *Plasmodium* primarily, *Plasmodium falciparum*, *Plasmodium ovale*, *Plasmodium malariae*, *Plasmodium knowlesi* and *Plasmodium vivax* are active in causing human malaria. The life cycle of malaria begins when blood is searched as meal by a female *Anopheline* mosquito (*Plasmodium* infected) and inserts sporozoites into the dermal cells [131, 132].

The parasite is resistant to conventional antimalarial drug that's why; there is a dire need to develop multiple approaches that can control distribution of this disease [34, 129]. Recently multi-drug resistant strains of the *Plasmodium* have developed and there are also evidences of the

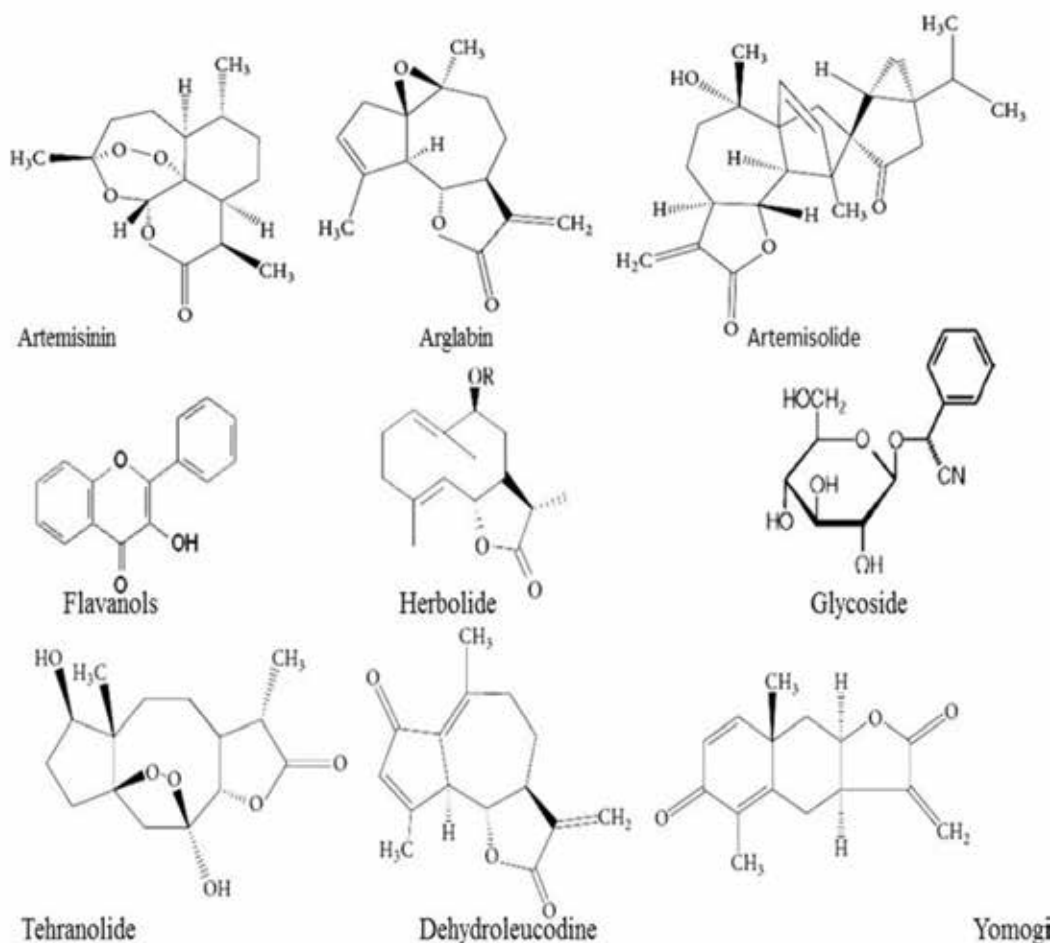


Fig. 2. Structure of some bioactive compounds derived from different *Artemisia* species (Source: Wikipedia commons).

occurrence of parasites resistant to artemisinin [133]. Several species of *Artemisia* are employed in China as antimalarial since prehistoric times especially *Artemisia annua*, *Artemisia apiacea* Hance and *Artemisia lancea* Vaniot [16, 134]. In 1970s, Chinese scientists found the effective antimalarial medicine, i.e. artemisinin from *Artemisia annua* L. which is a Chinese traditional medicinal plant [71]. Artemisinin is also obtained from different species of *Artemisia* and other microbial sources through genetic engineering techniques. Malaria and other diseases are treated with Artemisinin, which is actually a current drug of choice against *plasmodium falciparum*.

Artemisinin is essentially a sesquiterpenoid, and it is obtained from the glandular trichomes of *Artemisia annua* [135]. It contains endoperoxide bridge with in the 1, 2, 3-trioxane system (as shown in figure 2) that holds a significant rank due to its antimalarial properties. *Artemisia annua*, is wide spread in China and it is utilized since 2000 years to treat malaria [134]. Besides that, the presence of artemisinin has also been documented in *Artemisia lancea* and *Artemisia apiacea* [134], *Artemisia cina* [136] *Artemisia vulgaris*, *Artemisia japonica* [137] and in the upper portions of *Artemisia sieberi* [138]. Moreover, its presence has also been confirmed in *Artemisia absinthium* [139], *Artemisia dubia* and *Artemisia indica* [140].

Currently, the supreme operational means to get rid of the spreading rate of malaria is Artemisinin combination therapy (ACT) [141] and ACT is recommended by the World Health Organization (WHO) as a principal treatment for malaria initiated by *Plasmodium falciparum* [142]. Artemisinin also showed its detrimental effects against some other parasites together with *Schistosoma* [143], *Leishmania* [144] and *Toxoplasma* [145]. ACT is comparatively harmless medication because it has no clear antagonistic reactions or sterling side effects [146].

Studies confirmed that the hydro alcoholic and aqueous extracts of *Artemisia annua* L. are very operative on malaria and it gives antimalarial activity by potentiating artemisinin activity on *Plasmodium* [75]. Another study corroborated that the crude methanolic and ethanolic extracts of

Artemisia nilagirica (Clarke) have reasonably high potency against *plasmodium falciparum* [147]. An *In vitro* examination of crude extracts of *Artemisia abrotanum* L. showed that two compounds (isofraxadin and a novel sesquiterpene 1(S*)-hydroxy- α -bisaboloxide A acetate) are effective against *plasmodium falciparum* [148].

In a rodent antimalarial model, *Artemisia vulgaris* leaf extract is orally active, nontoxic against *Plasmodium yoelii* and as well as antiparasitic and antidisease weed. This plant has the tendency to be a low cost antimalarial source [149]. In other study, the light petroleum and ethanol extracts of *Artemisia maritima*, *Artemisia nilagarica* and *Artemisia japonica*, inhibit schizont maturation and are found to be atimalarial [150]. Another investigation revealed fifteen extracts of 3 *Artemisia* species from Iran namely; *Artemisia ciniformis*, *Artemisia biennis* and *Artemisia turanica* have better antimalarial activity. Dichloromethane extracts of *Artemisia ciniformis* have highest activity followed by *Artemisia biennis* and *Artemisia turanica* [81]. The dichloroethane extract of *Artemisia armeniaca* Lam. and *Artemisia aucheri* Boiss. also have better antimalarial activity [79].

In the quest of novel antimalarial compounds from plants, chloroform extracts fractions of *Artemisia maciverae* against *plasmodium berghei* have been assessed where triterpenes and alkaloids were found with antimalarial activity [151]. Investigations showed that the diverse solvent extracts and volatile oil of *Artemisia indica* have Artemisinin and polymethoxy flavanoid having antiprotozoal potential against different protozoal parasites, showing the antiparasitic worth of *Artemisia indica*. This plant extracts also have a potential malaria prophylactic effect, due to the inhibition of two plasmodial fatty acid biosynthesis enzymes [152].

An *In vitro* anti leishmanicidal analysis of ethanolic extracts of *Artemisia ciniformis*, *Artemisia santolina* and *Artemisia kulbadica* showed some compounds which have better anti leishmanicidal activity and found best for leishmaniasis management [153]. In another study, the anti-malarial effect have been found *In vivo* in

Iranian flora *Artemisia khorassanica* [60] and in *Artemisia turanica* Krasch crude extracts against *Plasmodium berghei* [116]. A dose dependent antiplasmodial activity of the methanolic extract from upper part of *Artemisia abyssinica* against chloroquine sensitive *Plasmodium berghei* has promising antiplasmodial activity [69]. Moreover, other *Artemisia* species like *Artemisia scoparia* Waldst. & Kit. And *Artemisia spicigera* C. Koch also possess antimalarial and free radical scavenging action and the dichloromethane extracts of these species have moderate level of antimalarial action [35]. These studies collectively endorse the pharmacological importance of *Artemisia* plants because of their high potency to combat with malaria.

4. ANTIMICROBIAL ACTIVITY OF *Artemisia* SPECIES

Since past decades, a lot of studies have been performed to reveal the anticancer, anti-inflammatory and antimicrobial properties of different constituents of plants [154–159]. The exploration of novel antimicrobial compounds with high effectiveness for deadly diseases is today's continuous and dire need [160]. Researchers are trying to develop effective drugs against microbial diseases by dragging their attention towards traditional medicine [161]. There are a lot of scientific revelations on the antimicrobial activity of plants [162] and numerous antimicrobial components have been identified from plant origin which are aromatic or might be some saturated carbon-based compounds. These aromatic compounds are attained by means of ethanolic or methanolic extraction [163]. Saponins and sterols are important compounds which can be extracted easily when methanol and ethanol are used as extracting solvents [164]. Other compounds like polyphenols [165] alkaloids [166] and terpenoids [167] can also be extracted using methanol and ethanol as extracting solvent. On the other hand, dichloromethane is also used for terpenoids extraction [162].

Fore mostly, the crude alcohol extraction method is employed in initial plant screening for antimicrobial activities and secondly, several other organic extraction methods are implemented

[121]. Numerous investigations validate the methanolic and ethanolic extracts of *Artemisia* species as better antimicrobial candidates [64, 66, 76, 97, 110, 120, 121, 168, 169, 170]. In a study antimicrobial efficacy of methanolic extracts of upper section of *Artemisia diffusa*, *Artemisia oliveriana*, *Artemisia scoparia* and *Artemisia turanica* against *S. aureus*, *B. subtilis*, *E. coli*, *C. albicans* and *P. aeruginosa* has been documented [169]. Important compounds like flavones could be obtained from *Artemisia giraldui* that have extraordinary antibiotic action contrary to several microorganisms including *P. aeruginosa*, *S. aureus*, *S. lutea*, *E. coli*, *Proteus sp.*, *T. viride* and *A. flavus* [91].

Essential oils of *Artemisia aucheri* contain compounds such as decane, p-cymene borneol, 1,8-cineole, linalool, lavandulol, triene, bornyl acetate, p-mentha-8-ol, chrysanthenyl acetate and caryophyllene oxide. These all essential compounds are recovered from the upper portions of *Artemisia aucheri* and the oils from seeds of this plant have better antimicrobial activity against *E. coli*, *S. aureus* and *Listeria monocytogenes* [171].

Similarly, in the essential oil of *Artemisia spicigera* compounds like, camphor-a-theojone, B-theojone, 1,8-cineole and p-cymene are active against various types of bacteria, i.e., *Bacillus cereus*, *Serratia marcescens*, *E. coli*, *Enterobacter aerogenes*, *Citrobacter amalanoficus*, *Bacillus megaterium*, *St. saprophyticus* and *Bacillus megatarium* [58]. Oils obtained from the aerial portion of *Artemisia incana* L. also contain a lot of compounds where camphor and borneol are abundant, showing inhibitory efficacy against twenty six bacteria, fifteen fungi and three yeast species [172]. Oils from *Artemisia feddei* also contain important compounds, which are highly active against obligate anaerobic bacteria [56]. *Artemisia chamaemelifolia*, *Artemisia turcomanica* and *Artemisia sipicigera* also possess antibacterial activity [58]. Invitro assessment of essential oil of *Artemisia aucheri* Boiss for antimicrobial effect authenticates better results against *B. cereus*, *P. vulgaris*, *P. aeruginosa*, *S. cereviciae*, *C. utilis*, *P. digitatum* and *A. niger* [59].

Table 1. Biological activities of substances, parts and fractions derived from *Artemisia* species.

Plant Species	Bioactive part or fraction	Bioactive substance	Reference
<i>Artemisia abrotanum</i> L.	Ethanolic extract	Antifungal Antibacterial	[64]
<i>Artemisia absinthium</i> L.	Aqueous MeOH extract	Antitumor	[65]
	Essential oil, methanolic and ethanolic extract	Antioxidant	[66-30]
	Hot alcoholic extract	Antimalarial	[67]
	Ethanolic extract	Cytoprotective	[30]
	Essential Oil Aqueous extract	Antifungal Anthelmintic	[68] [214]
<i>Artemisia abyssinica</i>	Methanolic extract	Antiplasmodial,	[69]
	Water and alcoholic extract	Antioxidant	[70]
<i>Artemisia annua</i> L.	Petroleum ether and ethyl acetate extract	Antimalarial Anti-algal	[71-72] [73]
	Methanol, water extract	Antibacterial	[74]
	Aqueous and hydro alcoholic extract	Antimalarial	[75]
	Essential oil	Antibacterial	[76]
	Ethanol extract	Antiulcerogenic	[77]
	Methanol and water extracts	Anticancer	[74]
<i>Artemisia arborescens</i>	Essential oils and methanolic extract	Antioxidant	[66]
<i>Artemisia argyi</i>	Methanolic extract	Antitumor	[78]
<i>Artemisia armeniaca</i> Lam	Dichloromethane extract	Antimalarial	[79]
<i>Artemisia aucheri</i> Boiss	Essential oil	Antimicrobial	[59]
	Dichloromethane extract	Antimalarial	[79]
<i>Artemisia barrelieri</i>	Eudesmanolides	Anti-inflammatory	[80]
<i>Artemisia biennis</i>	Ethyl acetate extract	Antimalarial	[81]
<i>Artemisia capillaris</i>	Methanolic extract	Antitumor	[82]
	Polyacetylene extract	Antiviral	[83]
	Crude extract	Antihepatitis	[84]
	Methanolic extract	Anticancer	[82-85]
	Methanolic extract	Antiobesity	[82-86]
<i>Artemisia cina</i>	Essential Oil	Antibacterial	[87]
		Antioxidant DNA protecting	
<i>Artemisia ciniformis</i>	Dichloromethane extract	Antimalarial	[81]
<i>Artemisia douglasiano</i>	Methanolic extract	Hepatoprotective	[88]
	Ethanol extract	Antiulcerogenic	[89]
<i>Artemisia feddei</i>	Essential Oil	Antibacterial	[56]
<i>Artemisia fukudo</i>	Ethanol, n-hexane, dichloromethane, ethylacetate, and butanol	Cytotoxic	[90]
<i>Artemisia girouldi</i> Pamp.	Flavones	Antimicrobial	[91]
<i>Artemisia iudoviciana</i>	Methanolic extract	Antimicrobial	[92]

Contd.....

Table 1 (Contd.....)

<i>Artemisia indica</i>	Chloroform, ethyl acetate and n-hexane extract	Antimicrobial	[93]
	Methanol extract	Anti-inflammatory	[94]
	Ethyl acetate extract	Antitumor	[97]
<i>Artemisia judaico</i>	Flavone, cirsimaritin	Antispasmodic	[95]
	Ethanol extract	Insecticidal	[63]
<i>Artemisia montana</i>	Ethanol extract	Cytoprotective	[30]
	Ethanol extract	Antioxidant	
<i>Artemisia myriantha</i>	Arglabin	Antitumor	[96]
<i>Artemisia Nilagirica</i>	Ethanol extract	Antibacterial	[31]
	Petroleum ether, n-hexane, dEther, Ethanol and water	Phytochemical analysis	[98]
	Essential oil	Antioxidant	[38]
<i>Artemisia pacifica</i>		Antimicrobial	[99]
<i>Artemisia princeps</i>	Smoke and water soluble extract	Anticancerous	[100]
<i>Artemisia parviflora</i>	Ethanol extract	Antioxidant	[42]
	Methanolic extract	Anthelmintic	[213]
<i>Artemisia rehan</i>	Ethanol extract	Antimalarial	[101]
<i>Artemisia scoparia</i>	Methanolic extract	Cytotoxic	[102]
	Dichloromethane extract	Antioxidant	[103]
	Methanol extract	Insecticidal	
	Essential oil and methanolic extract	Antioxidant Antibacterial Antimalarial	[66]
<i>Artemisia sieversiana</i>	Flavonoids	Antitumor	[65]
	Methanolic extract	Anthelmintic	[213]
<i>Artemisia stolonifera</i>	Ethanol extract	Cytostatic Antiobesity Anticancer Anti-inflammatory	[104]
<i>Artemisia tridentata</i> Nutt.	Ethanol extract	Plant growth regulator	[105]
<i>Artemisia xanthochroa</i>	Flavonoids	Antitumor	[65]
<i>Artemisia herba-alba</i> Asso.	Ethyl acetate, Butanol, Water and Chloroform extract	Anti-inflammatory Antioxidant Activity	[106]
	Essential oils	Antibacterial	[107]
	Aqueous extract	Hypoglycemic	[108]
	Essential oil	Antiradical	[107]
	Essential oil and crude extract	Hypoglycaemic	[109]
	Methanol extract	Anti-Listerial	[110]
<i>Artemisia Japonica</i>	Methanol extract	Anticancer	[82]
		Antiobesity	
		Anti-inflammatory	
<i>Artemisia Montana</i>	Methanol extract	Anticancer	[82]
		Antiobesity	
		Anti-inflammatory	
<i>Artemisia judaica</i>	Water and alcoholic extract	Anti-Diabetic	[111]

Contd.....

Table 1 (Contd.....)

<i>Artemisia Sylvatica</i>	Methanol extract	Anticancer Antiobesity Anti-inflammatory	[82]
<i>Artemisia Keiskeana</i>	Methanol extract	Anticancer Antiobesity Anti-inflammatory	[82]
<i>Artemisia Selengensis</i>	Methanol extract	Anticancer Antiobesity Anti-inflammatory	[82]
<i>Artemisia khorassanica</i>	Ethanol extract	Anti-plasmodial	[60]
<i>Artemisia kulbadica</i>	Methanol extract	Anticancer	[112]
<i>Artemisia sieberi</i>	Crude extracts	Antimalarial	[113]
	Essential oils	Antimalarial	[114]
	Ethanol extract	Antidiabetic	[115]
<i>Artemisia turanica</i>	Ethyl acetate extract	Anticancer	[112]
	Crude extract	Antimalarial	[81]
	Ethanol extract	Antimalarial	[116]
<i>Artemisia santolina</i>	Ethanol extract	Anticancer Activity	[112]
<i>Artemisia diffusa</i>	Essential oil	Anticancer Activity	[112]
<i>Artemisia campestris L.</i>	Ethanol, hexane and water extract	Antiradical	[107]
	Aqueous extract	Antioxidant, Antitumor	[117]
	Essential oils, Ethyl acetate extract	Antidiabetic	[118]
	Methanolic extract	Antibacterial	[119-107]
	Essential oils and methanolic extract	Antibacterial	[120-121]
	Essential oil	Antioxidant	[120, 66, 121]
	Dichloromethane extract, aqueous extract Water and alcoholic extract	Antimutagenic Anti-venomous	[122] [123-124]
<i>Artemisia afra</i>	Ethanol extract	Antioxidant	[70]
	Essential Oil	Antimalarial	[101]
<i>Artemisia dracunculus L.</i>	Essential Oil	Antioxidant	[68]
<i>Artemisia santonicum</i>	Essential oil and methanolic extract	Antifungal, Antibacterial	[68]
	Essential Oil	Antibacterial	[66]
<i>Artemisia spicigera</i>	Essential Oil	Antifungal, Antibacterial	[68]
	Dichloromethane extracts	Antimalarial Insecticidal	[103]
	Methanolic extract	Antioxidant	[103]
<i>Artemisia thuscula</i>	Ethanol extract	Diuretic	[125]
<i>Artemisia pallens</i>	Ethanol extract	Antibacterial Antifungal	[64]
<i>Artemisia vulgaris L.</i>	Essential oil	Antibacterial Antioxidant DNA protecting	[87]
	Methanolic extract	Anti-inflammatory	[126]

Against certain type of microorganisms, the methanolic extracts of *Artemisia campestris* L. are considered to be vigorous [120]. This might be due to the presence of bioactive metabolites of countless chemical types, like phenolic compounds. Shoko et al. [170] confirmed that phenolic compounds are very dynamic substances against microorganisms particularly bacteria. These compounds are quite active in contradiction of few Gram-positive species while the same extracts are weak against some Gram-negative species. *Artemisia campestris* is not merely an antimicrobial plant but also contains effective phenolic antioxidants [121]. The antimicrobial compounds modes of action in bacteria comprises membrane damage, membrane potential, changes in pH inside the cell, and the synthesis of ATP [173, 174].

Another study revealed the effective anti-viral properties of *Artemisia Parviflora* [41]. The antibacterial effect of crucial oil and crude extracts of *Artemisia herba-alba* Asoo. against *Listeria monocytogenes* have properties, that can hinder the progression of psychrophils resistant organisms [110]. One more study showed that the aqueous and solvent extracts of *Artemisia indica* were highly active against Gram-positive organism where *S. aureus*, was maximally inhibited [168]. These inhibitions might be due to the presence of essential compounds like phenols, steroids, triterpenoids, valavinoids, carotenoids, tetratriterpenoids azadirachtin and ketones [175]. Even though, extracts of few *Artemisia* species like *Artemisia aspera* and *Artemisia parviflora*, were not effective or having negligible inhibition on human and phytopathogenic bacteria [168].

Ethanolic extracts of other species of genus *Artemisia* like *Artemisia abrotanum* and *Artemisia pallens* are active against *Pseudomonas cepacia* and *Bacillus stearothermophilus*. These plants extracts not only possess antibacterial activity but also have maximum antifungal activity against *Trichosporon beigeli* and *Saccharomyces cerevisiae*. This suggests that the ethanolic extracts of these two novel plants have both antibacterial and antifungal potential [64]. *Artemisia nilagirica* is another important plant

containing numerous compounds including saponins, tannins, steroids, flavonoids, terpenoids, proteins and essential oil with better antibacterial action [97].

Studies of Erel et al. [66] substantiated that the methanolic extracts and essential oils of *Artemisia santonicum* and *Artemisia scoparia* holds fine antimicrobial activity where *Staphylococcus aureus* was the supreme sensitive bacteria to oils. Also these two plants are active against *Candida albicans* respectively. Some bacterial species, viz., *Salmonella enteritidis*, *Escherichia coli* O157, *Salmonella typhi*, *Listeria monocytogenes* and *Yersinia enterocolitica* were tested against the essential oil and compounds of *Artemisia annua* showing their high sensitivity [76]. In another study, Javid et al. [93] showed the chloroform, butanol and ethyl acetate extracts of *Artemisia indica* with better inhibitory activities towards *Salmonella typhi*. On the other hand, chloroform and n-Hexane extracts of this plant fully hinder the progression of fungal species like *Aspergillus flavus* and *Fusarium solani*.

Another study indicated that the methanolic extracts of *Artemisia ludoviciana* are more active against *Vibrio cholera* because these extracts encompasses compounds which are able to disturb the cell membranes of *Vibrio cholerae* cells with pH reduction, cell membrane hyperpolarization, and cellular ATP reduction [92]. Besides the antibacterial and antifungal activities, compounds from the extracts of *Artemisia annua* have anti-algal activity against *Microcystis aeruginosa*. This might be due to the presence of artemisinin which escalates the level of reactive oxygen species (ROS) in algae cells [73].

5. ANTICANCEROUS ACTIVITY OF *Artemisia* SPECIES

Medicinal plants possess a lot of natural products with better properties for cancer treatment [176]. Plants have numerous essential products like lignin and flavonoids of polyphenols. These products are evaluated *in vitro* and *in vivo* to find potential biological activities like antitumor activity [177]. Beforehand, a lot of studies have

been conducted to unfold the *in vitro* cytotoxic action of various plant extracts for their anticancer action on different types of human cancer cell lines [178, 179].

Like previously reported in other plants, several studies confirmed *Artemisia* species as better cytotoxic and anti-cancerous candidates [30, 90, 102, 126, 180, 197]. The poisonousness of *Artemisia* species on cancer cells has also shown *in vitro* [181, 182] and *in vivo* [183] respectively. These activities might be due to the presence of one or more essential compounds present in the plant. Among those compounds, Artemisinin, is very active ingredient of many *Artemisia* species mainly *Artemisia annua*, having better cellular toxicity against human lymphoid leukaemia cells [180]

Also the artemisinin and its allied compounds have the capacity to thwart cellular growth of human colorectal and breast cancer [180, 184]. Other compounds like terpenoids, sesquiterpen lactones and flavonoids are correspondingly important antitumor constituents acquired from *Artemisia* species [185]. Another offshoot of artemisinin, called Artesunate, possess both *in vitro* and *in vivo* anticancer properties [186].

A lot of beneficial compounds have also been well-known in *Artemisia absinthium* and *Artemisia vulgaris*, which have low molecular weight. These compounds are flavonoids, sesquiterpene, lactones, lignans and monoterpenes [187, 188, 189]. These are considered to be the main vigorous anticancerous compounds of these plants [190, 191, 192]. Another study corroborate the infusions from aerial parts of *Artemisia vulgaris* and *Artemisia absinthium* contain polysaccharides which are used in traditional plant made medicine [55]. Studies showed that the crucial consequence of the vigorous constituents of *Artemisia* species is apoptosis; it is a programmed cell death which is initiated via the cell cycle arrest [85, 90]. Instigation of caspases, mitochondrial membrane depolarization potential or the down governing expression of Bcl-2 gene might also induce apoptosis of cells [100]. Kim et al. [90] validates the utilization of *Artemisia fukudo* as a defensive measure against cancer. The

most active compound artemisinin induces apoptosis and it does not induce necrosis against human lymphoid leukaemia (Molt-4) cells [180].

Hitosugi et al. [193] reported, in the myelogenous leukaemia cell line of human (HL-60), *Artemisia capillaries* smoke and aqueous extracts are responsible for cellular decease, but these extracts are not effective in breast cancer (MCF-7) and other sort of tumour cells. On the other hand, macro molecular constituents of *Artemisia capillaris* are liable to encourage apoptosis in hepatoma cell lines in human [85]. The water soluble extracts of *Artemisia argyi* are not very much active against human tumour cell lines and also in breast cancer cell lines, but profoundly active in murine tumour cells [194]. In a study, the induction of apoptosis caused by the smoke and water extracts of *Artemisia princeps* in human breast cancer MCF-7 cells diminishes cells through the mitochondrial alleyway that seems to be a milestone for breast cancer treatment [100].

Artemisia argyi and *Artemisia Asiatic* also contain essential compound called flavones, which have the potency to impede certain types of cancer by promoting apoptosis including human lung cancer, prostate cancer, myeloid leukaemia, gastric cancer and melanoma [78, 195]. Nevertheless, other researchers found flavones to be unproductive in contradiction of human breast cancer cells [196]. Similarly, n-hexane extracts of *Artemisia turanica* Krash. possess better cytotoxic, antiproliferative and anticancer effects against two leukemic cancer cell lines predominantly HL-60 and K562 [197]. In another study dichloromethane, methanol, ethyl acetate, and n-hexane extracts from upper parts of different *Artemisia* species (*Artemisia ciniformis*, *Artemisia diffusa* Karasch, and *Artemisia vulgaris*) have potent antiproliferative properties which could be a promising chemotherapeutic agent in cancer treatment [198].

Studies confirmed that the ethanolic extracts of *Artemisia montana* and *Artemisia absinthium* are rich in essential compounds like, flavonoids and phenolic acids. These compounds have better antioxidant activity and also have cytoprotective influence towards oxidative damage in fibroblast-

like cells. This validates *Artemisia montana* and *Artemisia absinthium* both as better nominees for the treatment of skin disorders [30].

Extracts of *Artemisia scoparia* in human muscle cancer cells have devastating effect against 88-93% cancer cells that endorse anticancer activity of this plant extract [102]. Moreover, the apex parts of two novel species of *Artemisia* i.e., *Artemisia vulgaris* and *Artemisia absinthium* have anthelmintic, antipyretic, cytostatic, stomachic, antibacterial, and antitumor actions [68, 199, 200] while the *In vitro* assessment of methanol extracts of other species like *Artemisia Japonica*, *Artemisia stolonifera*, *Artemisia montana*, *Artemisia selengensis*, *Artemisia capillaris*, *Artemisia sylvatica*, *Artemisia scoparia* and *Artemisia keiskeana* possess better anti-inflammatory, anticancer, and antiobesity activity [82]. Studies of Emami et al. [112] corroborated *Artemisia sieberi*, *Artemisia kulbadica*, *Artemisia santolina*, *Artemisia turanica*, and *Artemisia diffusa* with cytotoxic activity in contradiction of human Caucasian hepatocyte and larynx carcinoma (HepG-2 and Hep-2) cell lines.

6. ANTHELMINTIC ACTIVITY OF *Artemisia* SPECIES

Helminthic problems are exceedingly widespread, predominantly in the 3rd world countries [201] and documented as the cause of much chronic ailments. Numerous studies have found *Artemisia* species with potent anthelmintic activity [62, 202, 203, 204, 205]. *Artemisia cina* is one of the best candidates with anthelmintic activity which contains santonin, a sesquiterpenic lactone that might be the reason of this activity [206, 207]. Other species like *Artemisia santonica* L, *Artemisia maritima*, *Artemisia herba-alba*, *Artemisia absinthium*, *Artemisia vulgaris*, *Artemisia afra* and *Artemisia ludoviciana* are also most prominent species with the same activity [208].

In one study, Extracts from *Artemisia vestita* and *Artemisia maritima* are found active against *Haemonchus contortus* in infected sheep's and indicated significant activity against larvae and

adult worms [209]. Moreover, in ruminants, the water, aqueous, sodium bicarbonate, dichloromethane, and ethanol extracts obtained from leaves of *Artemisia annua* have better anthelmintic action [202]. Perennial plant *Artemisia indicia* also possess this activity. In a study chloroform, methanol and aqueous extracts of this plant confirmed anthelmintic property against adult earthworm *Pheretima posthuma* [62]. *Artemisia absinthium* extracts are also a promising way to treat GI nematodes of sheep [210]. An important member of *Artemisia* is *Artemisia herba alba*, that can be employed for controlling heterakid infection because it induces anthelmintic consequence by dropping worm burden and egg shedding in the diseased birds [203] and also the methanolic extracts from leaves of *Artemisia herba-alba* possess nematicidal activity [211]. The anthelmintic effects on *Haemonchus contortus* from methanol and crude aqueous and of *Artemisia brevifolia* have been proved and it is confirmed that the whole plant holds strong anthelmintic activity against nematodes [212]. On the other hand, the essential oil of *Artemisia pallens* have tendency of strong anthelmintic action against *Taenia solium*, *Pheritima posthuma* and *Ascaris lumbricoides* [204]. Chloroform extracts of stem and root of *Artemisia siversiana*, also hold potency to eradicate *H. nana* from infected mice [205].

The anthelmintic activity of extracts from *Artemisia parviflora* and *Artemisia sieversiana* was evaluated *in vitro* and *in vivo* on *Haemonchus contortus*, which is a parasitic nematode of small ruminants. Methanolic extract of these plants tested against three different developmental stages using different assays were found to be better anthelmintic candidates [213].

An *in vitro* study was conducted to find an alternative to anthelmintic praziquantel by checking the activity of the crude aqueous extract of *Artemisia absinthium* against *H. nana*. The extract from *Artemisia absinthium* was found to increase ultrastructural alterations, worm paralysis and ultimately death in a dose-dependent manner. Also a significant decrease in the EPG and worm burden has been noticed in mice treated with *A.*

absinthium [214]. Such studies clearly indicate *Artemisia* species with their ability to control helminthic disease to a broad spectrum.

7. CONCLUSIONS

Artemisia is a noteworthy genus with a large variety of biological activities. Plants of this genus possess an extensive range of pharmacological activities which could be used in numerous medical applications. Various active compounds achieved from these plants need to be characterized and well documented; also, the exploration of other novel species with disclosure of new chemical constituents is necessary. Their clinical effectiveness must be tested to obtain better results against lethal diseases. Finally, the toxic effect of these plants should also be clarified and the genetically modified varieties of *Artemisia* need to be cultivated.

8. REFERENCES

- Funk, V., A. Susanna, T.F. Stuessy, & R. Bayer (Ed.) *Systematics, evolution, and biogeography of the compositae*. IAPT. Vienna (2009).
- Oberprieler, C., S. Himmelreich, M. Kaallersjo, J. Valle's, & R. Vogt. Anthemideae. In: *Systematics, evolution, and biogeography of the Compositae*, Funk V, Susanna A, Stuessy TF. Bayer R (Ed.). IAPT, Vienna, p. 631–666 (2009).
- Bremer, K. & C.J. Humphries. Generic Monograph of the *Asteraceae- Anthemideae*. *Bulletin of Natural History Museum, London (Bot.)* 23: 71-177 (1993).
- Martin, J., M. Torrel, A.A. Korobkov, & J. Valles. Palynological features as systematic marker in *Artemisia* L. and related genera (*Asteraceae, Anthemideae*)—II: implications for subtribe Artemisiinae delimitation. *Plant Biology* 5(1): 85-93 (2003).
- Valle's, J. & E.D. McArthur. *Artemisia* systematics and phylogeny: Cytogenetic and molecular insights. In: *Shrub land Ecosystem Genetics and Biodiversity*. Proceedings, Rocky Mountain Research Station, E.D. McArthur & D.J. Fairbanks (Ed.), p. 67–74 (2001).
- Jarvis, C.E., F.R. Barrie, D.M. Allan, & J.L. Reveal. (Ed.) *A list of Linnaean Generic Names and Their Types Regnum Vegetabile*. Koeltz Scientific Books, for the International Association of Plant Taxonomy. *Konigstein*, 127 pp. (1993).
- Watson, L.E., P.L. Bates, T.M. Evans, M.M. Unwin, & J.R. Estes. Molecular phylogeny of Subtribe *Artemisiinae (Asteraceae)*, including *Artemisia* and its allied and segregate genera. *BMC Evolutionary Biology* 2: 17 (2002).
- Mabberley, D.J. *Mabberley's Plant-Book. A Portable Dictionary of Plants, Their Classifications and Uses*, 3rd ed. Cambridge University Press, Cambridge (completely revised) (2008).
- Valle's, J. & T. Garnatje. *Artemisia* and its allies: Genome organization and evolution and their biosystematic, taxonomic and phylogenetic implications in the Artemisiinae and related subtribes (*Asteraceae, Anthemideae*). In: *Plant Genome: Biodiversity and Evolution. Vol. 1B: Phanerogams*, A. Sharma (Ed.), p. 255–285 (2005).
- Ling, Y.R. On the systematics of genus *Artemisia* L. and the relationship with its allies. *Bulletin of Botanical Laboratory of North-East Forestry Institute* 2: 1-60 (1982).
- McArthur, E.D. & C.L. Pope. Karotypes of four *Artemisia* species: *A. carruthii*, *A. filifolia*, *A. frigida*, and *A. Spinescens*. *Great Basin Naturalist* 39(4): 419–426 (1979).
- Robonson, M.M. & X. Zhang. *The World Medicine Situation 2011, Traditional Medicine: Global Situation, Issues and Challenges*, 3rd ed. WHO, Geneva (2011).
- Al-Zubairi, A.S., A.B. Abdul, S.I. Abdel wahab, C.Y. Peng, S. Mohan, & M.M. Elhassan. *Eleucine indica* possesses antioxidant, antibacterial and cytotoxic properties. *Evidence Based Complement Alternate Medicine* 2011:965370 (2011). doi: 10.1093/ecam/nep091
- Ekwenye, U.N. & N.N. Elegalam. Antibacterial activity of ginger (*Zingiber officinale* Roscoe and Garlic (*Allium sativum* L.) extracts on *Escherichia coli* and *Salmonella typhi*. *International Journal of Molecular Medicine and Advance Sciences* 1(4): 411-417 (2005).
- Dahanukar, S.A., R.A. Kulkarni, & N.N. Rege. Pharmacology of medicinal plants and natural products. *Indian Journal of Pharmacology* 32: S81–S118 (2000).
- Willkox, M. *Artemisia* species: From traditional medicines to modern antimalarials and back again. *Journal of Alternative & Complementry Medicine* 15(2): 101–109 (2009).
- Wright, C.W. (Ed.) *Artemisia*. Taylor & Francis, London. (series Medicinal and Aromatic Plants—Industrial Profiles) (2002).
- Tan, R.X., W.F. Zheng, & H.Q. Tang. Biologically active substances from the genus *Artemisia*. *Planta Medica* 64(4): 295–302 (1998).

19. Parada, M., E. Carrio, M.A. Bonet, & J. Valles. Ethnobotany of the Alt Emporda` region (Catalonia, Iberian Peninsula). Plants used in human traditional medicine. *Journal of Ethnopharmacology* 124(3): 609–618 (2009).
20. Lewis, W.H. & M.P. Elvin-Lewis. Medicinal plants as a source of new therapeutics. *Annals of Missouri Botanical Garden* 82(1): 16-24 (1997).
21. Parr, A.J. & G.P. Bolwell. Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying the phenols content or profile. *Journal of Science of Food & Agriculture*, 80(7): 985-1012 (2000).
22. Alghazeer, R., H. Gao, & N.K. Howell. Cytotoxicity of oxidised lipids in cultured colonal human intestinal cancer cells (caco-2 cells). *Toxicology Letters* 180(3): 202-211 (2008).
23. Ferreira, D. & D. Stade. Oligomeric proanthocyanidins: naturally occurring O-heterocycles. *Natural Product Research* 19(5): 517-4 (2002).
24. Lone, S.H., K.A. Bhat, & M.A. Khuroo. Chemical and Pharmacological Perspective of *Artemisia amygdalina*. *Springers. Briefs in Pharmacology & Toxicology* (2015), DOI 10.1007/978-3-319-25217-9_2.
25. Ruwali, P., K.T. Ambwani, P. Gautam, & A. Thapliyal. Qualitative and quantitative phytochemical analysis of *Artemisia indica* Will. *Journal of Chemistry & Pharmaceutical Research* 7(4): 942-949 (2015).
26. Parameswari, P. & R. Devika. Phytochemical screening and evaluation of *Artemisia nilagirica* (clarke) pamp. BY GC-MS. *International Journal of Pharm Science Research* 8(1): 222-225 (2017).
27. Ashok, P.K. & K. Upadhyaya. Preliminary Phytochemical Screening and Physico-Chemical Parameters of *Artemisia absinthium* and *Artemisia annua*. *Journal of Pharmacognosy and Phytochemistry* 1(6): 229-235 (2013).
28. Sivagnanam, S.K., R.K. Rao, U.M. Mudiganti Dar, P.G. Jeelani. Preliminary phytochemical analysis of *Artemesia amygdalina*, *Neriumodorum* and *Strychno spotatorium*. *Journal of Pharmaceutical Research* 5(7): 3734-3739 (2012).
29. Iqbal, S., U. Younas, K.W. Chan, M. Zia-Ul-Haq, & M. Ismail. Chemical Composition of *Artemisia annua* L. Leaves and Antioxidant Potential of Extracts as a Function of Extraction Solvents. *Molecules* 17(5): 6020-6032 (2012).
30. Craciunescu, O., D. Constantin, A. Gaspar, L. Toma, & E. Utoiu, L. Moldovan. Evaluation of antioxidant and cytoprotective activities of *Arnica montana* L. and *Artemisia absinthium* L. ethanolic extracts. *Chemistry Central Journal* 6: 97 (2012).
31. Devmurari, V.P. & N.P. Jivani. Phytochemical screening and antibacterial activity of ethanolic extract of *Artemisia Nilagirica*. *Annals of Biological Research* 1(1): 10-14 (2011).
32. Willcox, M., G. Bodeker, G. Bourdy, V. Dhingra, J. Falquet, J.F.S. Ferreira, B. Graz, H.M. Hir, E. Hsu, P.M. de Magalhães, D. Provendier, & W.C. Wright. *Artemisia annua* as a traditional herbal antimalarial. In: *Traditional Medicinal Plants in Malaria*, M.L. Willcox, G. Bodeker, P. Rasoanaivo (Ed.). CRC Press, BocaRaton, FL, p. 43–59 (2004).
33. Marco, J.A. & O. Barbera. Natural products from the genus *Artemisia* L. In: Atta-ur Rahman (Ed.). *Studies in Natural Product Chemistry* 7: 201-264 (1990).
34. Li, Y., H. Huang, & Y.L. Wu. Qinghaosu (Artemisinin) – a fantastic antimalarial drug from a traditional Chinese medicine. In: *Medicinal Chemistry of Bioactive Natural Products*, Liang, Fang (Ed.). Wiley, p. 183–256 (2006).
35. Afshar, F.H., A. Delazar, O. Janneh, H. Nazemiyeh, A. Pasdaran, L. Nahar, & S.D. Sarker. Evaluation of antimalarial, free-radical scavenging and insecticidal activities of *Artemisia scoparia* and *A. spicigera*, *Asteraceae*. *Brazilian Journal of Pharmacognosy* 21(6): 986-990 (2010).
36. Krtikar, K.P. & B.D. Basu. In *Indian Medicinal Plant*, 2nd ed. Periodical Expert, New Delhi, 887 (1975)
37. Shafi, P.M., M.K. Nambier, M.K., Geetha, R.A. Clery, Y.R. Sarma, & S.S. Veena. Composition and Antifungal Activity of the Oil of *Artemisia nilagirica* (Clarke) Pamp. *Journal of Essential Oil Research* 16(4): 377-379 (2004).
38. Tripathi, Y.C., V. Bisht, & N. Anjum. Compositional Analysis and In-Vitro Antioxidant Activity of Essential Oil of *Artemisia nilagirica* leaves. *World Journal of Pharmaceutical Research* 4(9): 1663-1679 (2015).
39. Kimothi, G.P. & B.C.L. Shah. Some medicinal plants of gopeshwar - tungnath region of Uttar Pradesh. *Ancient Science Life* 8(3&4): 283-292 (1989).
40. Ahmad, S., A. Ali, H. Beg, A.A. Dasti, & Z.K. Shinwari. Ethnobotanical studies on some medicinal plants of Booni Valley, District Chitral Pakistan. *Pakistan Journal of Weed Science Research* 12(3): 183-190 (2006).
41. Anonymous. The Wealth of India. Vol I: A, Publications and Information Directorate, CSIR, New Delhi. 442-443 (1985).

42. Ahuja, J., J. Suresh, A. Deep, Madhuri, & R. Pratyusha. Phytochemical screening of aerial parts of *Artemisia parviflora* Roxb: A medicinal plant. *Der Pharmaca Lettre* 3(6): 116-124 (2011).
43. Ahameethunisa, A.R. & W. Hopper. In: vitro antimicrobial activity on clinical microbial strains and antioxidant properties of *Artemisia parviflora*. *Annals of Clinical Microbiology and Antimicrobials* 11(30): 1-7 (2012).
44. Alzoreky, N.S. & K. Nakahara. Antibacterial activity of extracts from some edible plants commonly consumed in Asia. *International Journal of Food Microbiology* 80(3): 223-230 (2003).
45. Guarrera, P.M. Traditional antihelmintic, antiparasitic and repellent uses of plants in Central Italy. *Journal of Ethnopharmacology* 68(1-3): 183-192 (1999).
46. Mitich, L.W. Absinth wormwood—a problem weed? *Proceedings of NC Weed Cont. Conference* 30: 41-42 (1975).
47. Jansen, F.H. The herbal tea approach for artemisinin as a therapy for malaria. *Transactions of Royal Society of Tropical Med Hygiene*, 100(3): 285-286 (2006).
48. Hernandez, H., J. Mendiola, D. Torres, N. Garrido, & N Perez. Effect of aqueous extract of *Artemisia* on the invitri culture of *Plasmodium falciparum*. *Fitoterapia* 41(6): 540-541 (1990).
49. Rao, V.S.N., A.M.S. Menezes, & M.G.T. Gadelha. Antifertility screening of some indigenous plants of Brazil. *Fitoterapia* 59: 17-20 (1988).
50. Park, E., J. Nan, J. Kim, H. Kang, J. Choi, S. Lee, B. Lee, S. Kim, J. Lee, Y. Kim, & D. Sohn. The ethanol-soluble part of a hot-water extract from *Artemisia iwayomogi* inhibits liver fibrosis induced by carbon tetrachloride in rats. *Journal of Pharmacy and Pharmacology* 52(7): 875-81 (2000).
51. Maefi, F.R., M. Carini, G. Aldin, i E. Bombardelli, P. Marazzoni, et al. Free radicals scavenging action and anti-enzyme activities of procyanidins from vitisvinifera. *Arzneimittel-Forschung Drug Research* 44(5): 592-601 (1994).
52. Sakipova, Z., N.S.H. Wong, T. Bekezhanova, Sadykova, A. Shukirbekova, & F. Boylan. Quantification of santonin in eight species of *Artemisia* from Kazakhstan by means of HPLC-UV: Method development and validation. *PLoS ONE* 12(3): e0173714 (2017).
53. Haniya, A.K. & P.R. Padma. Phytochemical investigation of methanol extracts of *Artemisia vulgaris* L. Leaves. *International Journal of Pharmac Biological Science* 5(2): 184-195 (2014).
54. Terra, D.A., L. de Fátima Amorim, M.T.J. de Almeida Catanho, A. de Souza da Fonseca, S.D. Santos-Filho, J. Brandão-Neto, A. da Cunha Medeiros, & M. Bernardo-Filho. Effect of an extract of *Artemisia vulgaris* L. (Mugwort) on the in vitro labelling of red blood cells and plasma proteins with technetium-99m. *Brazilian Archives of Biology and Technology* 50: 123-128 (2007).
55. Corrêa-Ferreira, M.L., G.R. Noletto, & C.L.O. Petkowicz. *Artemisia absinthium* and *Artemisia vulgaris*: A comparative study of infusion polysaccharides. *Carbohydrate Polymer* 102: 738-745 (2014).
56. Cha, J.D., E.E. Jung, B.S. Kil, & K.Y. Lee. Chemical composition and antibacterial activity of essential oil from *Artemisia feddei*. *Journal of Microbiology and Biotechnology* 17(12): 2061-65 (2007).
57. Masoudi, S., A. Rustaiyan, & M. Vahedi. Volatile oil constituents of different parts of *Artemisia chamaemelifolia* and the composition and antibacterial activity of the aerial parts of *A. turcomanica* from Iran. *Natural Product Communication* 7(11): 1519-22 (2012).
58. Chehregani, A., M. Atri, S. Yousefi, Z. Albooyeh, & F. Mohsenzadah. Essential oil variation in the populations of *Artemisia spicegera* from northwest of Iran: Chemical composition and antibacterial activity. *Pharmaceutical Biology* 51(2): 246-52 (2013).
59. Aali, H., Mohamadi, A. Sani, & S.H. Mohseni. Chemical composition and antimicrobial effect of the essential oil of *Artemisia aucheri* boiss endemic in khorasan Iran. *Trends in Life Science* 3(2): 1-9 (2014).
60. Nahrevanian, H., B. Esmaeili, M. Kazemi, H. Nazem, & M. Amini. In vivo antimalarial effects of IranianFlora *Artemisia khorassanica* against plasmodium bergheiaand pharmaco chemistry of its natural components. *Iranian Journal of Parasitology* 5(1): 6-19 (2010).
61. Rather, M.A., B.A. Dar, W.A. Shah, A. Prabhakar, K. Bindu, J.A. Banday, & M.A. Qurishi. Comprehensive GC-FID GC-MS and FT-IR spectroscopic analysis of the volatile aroma constituents of *Artemisia indica* and *Artemisia vestita* essential oils. *Arab Journal of Chemistry* 5:1-6 (2014).
62. Sarnim, G., S.T. Sanjay, A. Roshan, A.B. Vedamurthy, & H. Joy Hoskeri. *Artemisia indica* extracts as anthelmintic agent against *Pheretima posthuma*. *International Journal of Pharmacy and Pharmaceutical Science* 5(3): 259-262 (2013).
63. Acheuk, F., W. Lakhdari, K. Abdellaoui, M. Belaid, R. Allouane, F. Halouane F. Phytochemical study and bioinsecticidal effect of

- the crude ethonolic extract of the algerian plant *Artemisia judaica* L. (*Asteraceae*) against the black bean aphid, *Aphis fabae* Scop. *Agriculture and Forestry* 63 (1): 95-104 (2017)
64. Suresh, J., A. VasaviReddy, D. Rajan, M. Ihsanullah, & M.N. Khan. Antimicrobial activity of *Artemisia abrotanum* and *Artemisia pallens*. *International Journal of Pharmacognosy and Phytochemistry Research* 3(2): 18-2 (2010).
 65. Chemesova, L.L., L.M. Belenovskaya, & A.N. Stukov. Anti-tumour activity of flavonoids from some *Artemisia* species. *Rastitel'nye Ressearch* 23: 100-103 (1987).
 66. Erel, S.B., G. Reznicek S.G. Şenol, N.U.K. Yavaşoğlu, S. Konyalıoğlu, & A.U. Zeybek. Antimicrobial and antioxidant properties of *Artemisia* L. species from western Anatolia. *Turkish Journal of Biology* 36(2012): 75-84 (2012).
 67. Irshad, S., & A. M. Mirza. Antimalarial Activity of Three Pakistani Medicinal Plants. *Pakistan Journal of Pharmaceutical Science* 24(4): 589-591 (2011).
 68. Kordali, S., R. Kotan, A. Mavi, A. Cakir, & A. Ala. Yildirim. Determination of the chemical composition and antioxidant activity of the essential oil of *Artemisia dracunculus* and of the antifungal and antibacterial activities of Turkish *Artemisia absinthium*, *A. dracunculus*, *Artemisia santonicum*, and *Artemisia spicigera* essential oils. *Journal of Agriculture and Food Chemistry* 53(24): 9452-9458 (2005).
 69. Adugna, M., T. Feyera, W. Taddese, & P. Admasu. In vivo antimalarial activity of crude extract of aerial part of *Artemisia abyssinica* against *Plasmodium berghei* Mice. *Global Journal of Pharmacology* 8(3): 460-468 (2014).
 70. Burits, M., K. Asres, & F. Bucar. The antioxidant activity of the essential oils of *Artemisia afra*, *Artemisia abyssinica* and *Juniperu sprocera*. *Phytotherapy Research* 15(2): 103-108 (2001).
 71. Qinghaosu Antimalarial Coordinating Research Group. *Chininese Medicine Journal* 92(12): 811-816 (1979)
 72. Liu, C.Z., S.J. Murch, & M. El-Demerdash. Regeneration of Egyptian medicinal plant *Artemisia judaica* L. *Plant Cell Reproduction* 21(6): 525-530 (2003).
 73. Ni, L., K. Acharya, X. Hao, & S. Li. Isolation and identification of an anti-algal compound from *Artemisia annua* and mechanisms of inhibitory effect on algae. *Chemosphere* 8: 1051-1057 (2012).
 74. Nageeb, A., A. Al-Tawashi, A.H.M. Emwas, Z.A.H. Al-Talla, & N. Al-Rifai. Comparison of *Artemisia annua* bioactivities between traditional medicine and chemical extracts. *Current Bioactive Compounds* 9(4): 324-332 (2013).
 75. Diawara, H.Z., H. Ganfon, F. Gbaguidi, A. Yemoa, J. Bero, O. Jansen, B. Evrard, M. Moudachirou, M. Frederich, & J.Q. Leclercq. The antimalarial action of aqueous and hydro alcoholic extracts of *Artemisia annua* L. Cultivated in Benin: In vitro and in vivo studies. *Journal of Chemistry and Pharm Research* 7(8): 817-823 (2015).
 76. Donato, R., F. Santomauro, A.R. Bilia, G. Flamini, & C. Sacco. Antibacterial activity of Tuscan *Artemisia annua* essential oil and its major components against some foodborne pathogens. *LWT-Food Science and Technology* 64(2): 1251-1254 (2015).
 77. Foglio, M.A., P.C. Dias, M.A. Antonio, A. Possenti, R.A.F. Rodrigues, E.F. Silva, V.L.G. Rehder, & J.E. Carvalho. Antiulcerogenic activity of some sesquiterpene lactones isolated from *Artemisia annua*. *Planta Medica* 68(6): 515-518 (2002).
 78. Seo, J.M., H.M. Kang, K.H. Son, J.H. Kim, C.W. Lee, H.M. Kim, S.I. Chang, & B.M. Kwon. Antitumor activity of flavones isolated from *Artemisia argyi*. *Planta Medica* 69(3): 218-222 (2003).
 79. Mojarrab, M., A. Shiravand, A. Delazar, & F.H. Afshar. Evaluation of in vitro antimalarial activity of different extracts of *Artemisia aucheri* Boiss. and *A. armeniaca* Lam. and fractions of the most potent extracts. *The Scientific World Journal* Article ID 825370: p. 6 (2014).
 80. Zafra-Polo, M.C. and M.A. Blazquez. Antiinflammatory activity of Sesquiterpene lactones from *Artemisia barrelieri* in rats. *Phytotherapy Research* 5: 91-93 (1991).
 81. Mojarrab M, R. Naderi, & F.H. Afshar. Screening of different extracts from *Artemisia* species for their potential antimalarial activity. *Iran Journal of Pharmaceutical Research* 14(2): 603-608 (2015).
 82. Choi, E., H. Park, J. Lee, & G. Kim. Anticancer, antiobesity, and anti-inflammatory activity of *Artemisia* species in vitro. *Journal of Traditional Chinese Medicine* 33(1): 92-97 (2013).
 83. Matsumoto, A., H. Katsuya, T. Matsumoto, & H. Tokuda. *Antitumor polyacetylene extraction from plants*. Japan Kokai, Tokyo Koho, Japanese Patent: JP 03287532 A2 19911218 Heisei 4: (1991).
 84. Kiso, Y., S. Ogasawara, K. Hirota, N. Watanabe, Y. Oshima, C. Konno, & H. Hikino. Antihepatotoxic principles of *Artemisia capillaris* buds. *Planta Medica* 50: 81-85 (1984).
 85. Hu, Y.Q., R.X. Tan, M.Y. Chu, & J. Zhou. Apoptosis in human hepatoma cell line SMMC-7721 induced by water-soluble macromolecular

- components of *Artemisia capillaries* Thunberg. *Japanese journal of Cancer Research* 91(1): 113-7 (2000).
86. Yoon, M. & M.Y. Kim. The anti-angiogenic herbal composition Ob-X from *Morus alba*, *Melissa officinalis*, and *Artemisia capillaris* regulates obesity in genetically obese ob/ob mice. *Pharmaceutical Biology* 49(8): 614-619 (2011).
 87. Prakash, B., P. Singh, A. Kedia, & N.K. Dubey. Assessment of some essential oils as food preservatives based on antifungal, antiaflatoxin, antioxidant activities and in vivo efficacy in food system. *Food Research International* 49(1): 201-208 (2012).
 88. Guerreiro, E., E.E. Garcia, M.J. Pestchanker, R.D. Enri, A.M. Rodriguez, A. Maria, & G.H. Wendel. Cytoprotective activity of minor constituents of *Artemisia douglasiana*. *Natural Product Letters* 6(4): 269-280 (1995).
 89. Giordano, O.S., E. Guerreiro, M.J. Pestchanker, J. Guzman, D. Pastor, & T. Guardia. The gastric cytoprotective effect of several sesquiterpene lactones. *Journal of Natural Product* 53(4): 803-809 (1990).
 90. Kim, K.N., J.A. Lee, W.J. Yoon, J.Y. Kim, G.P. Song, & S.Y. Park. The cytotoxicity of *Artemisia fukudo* extracts against HL-60 cells. *Journal of Korean Society of Food Science & Nutrition* 36: 819-824 (2007)
 91. Zheng, W.F., R.X. Tan, L. Yang, & Z.L. Liu. Two flavones from *Artemisia giraldii* and their antimicrobial activity. *Planta Medica* 62(2): 160-2 (1996).
 92. Sa'nchez, E., S. Garcia, & N. Heredia. Extracts of edible and medicinal plants damage membranes of *Vibrio cholera*. *Applied & Environmental Microbiology* 76(20): 6888-6894 (2010).
 93. Javid, T., M. Adnan, A. Tariq, A. Akhtar, R.U. Naser, & M. AbdElsalam. Antimicrobial activity of three medicinal plants (*Artemisia indica*, *Medicago falcate* and *Tecomastans*). *African Journal of Traditional Complementary and Alternative Medicine* 12(3): 91-96 (2015).
 94. Sagar, M.K., P.K. Ashok, H. Chopra, & K. Upadhyaya. Phytochemical and pharmacological potential of *Artemisia indica* in experimental animal models. *Pharmacologyonline* 2: 1-4 (2010).
 95. Abdalla, S.S. & M.H. Abu-Zarga. Effects of cirsimaritin, a flavone isolated from *Artemisia judaica*, on isolated guinea-pig ileum. *Planta Medica* 53(04): 322-324 (1987).
 96. Appendino, G., P. Gariboldi, & F. Menichini. The stereochemistry of arglabin. a. Cytotoxic guaianolide from *Artemisia myriantha*. *Fitoterapia* 62(3): 275-276 (1991).
 97. Zeng, Y.T., J.M. Jiang, H.Y. Lao, J.W. Guo, N.Y. Lun, & M. Yang. Antitumor and apoptotic activities of the chemical constituents from the ethyle acetate extract of *Artemisia indica*. *Molecular Medicine Reports* 11(3): 2234-2240 (2015).
 98. Rani, N.P., C. Moorthi, R. Senthamarai, & K. Kathiresan. A study to explore the pharmacognostic and phytochemical screening of *Artemisia nilagirica* leaves found in Nilgiris district of Tamil nadu. *International Journal of Pharmacy and Pharmaceutical Science* 4(4): 441-447 (2012).
 99. Wahyuono, S., J.J. Hoffmann, & S.R. McLaughlin. Dehydro faltarindiol, potential antimicrobial agent from *Artemisia pacifica*. *Fitoterapia* 63: 368 (1992).
 100. Sarath, V.J., C.S. So, D.W. Young, & S. Gollapudi. *Artemisia princeps* varoriental is induces apoptosis in human breast cancer MCF-7 cells. *Anticancer Research* 27(6B): 3891-3898 (2007).
 101. Kassa, M., R. Mshana, A. Regassa, & G. Assefa. *In vitro* test of five Ethiopian medicinal plants for antimalarial activity against *plasmodium falciparum*. SINET. *Ethiopian Journal of Science* 21(1): 81-89 (1998).
 102. Maqsood, M., R. Qureshi, M. Ikram, S. Ali, M. Rafi, J.A. Khan, & M.S. Ahmed. Preliminary screening of methanolic plant extracts against human rhabdomyo sarcoma cell line from salt range Pakistan. *Pakistan Journal of Botany* 47(1): 353-357 (2015).
 103. Afshar, F.H., A. Delazar, O. Janneh, H. Nazemiyeh, A. Pasdaran, L. Nahar, & S.D. Sarker. Evaluation of antimalarial, free-radical scavenging and insecticidal activities of *Artemisia scoparia* and *A. spicigera*, Asteraceae. *Brazilian Journal of Pharmacognosy* 21(6): 986-990 (2011).
 104. Lee, K.R., S.W. Hong, J.H. Kwak, S. Pyo, & O.P. Jee. Phenolic constituents from the aerial parts of *Artemisia stolonifera*. *Archives of Pharmacal Research* 19(3): 231-234 (1996).
 105. Farmer, E.E. & C.A. Ryan. Interplant communication: Airborne methyl jasmonate induces synthesis of proteinase inhibitors in plant leaves. *Proceedings of National Academy of Science. USA* 87(19): 7713-7716 (1990).
 106. Eddine, L.S., O.M. Redha, & S. Ladjel. Influence of solvent extraction on phenolic content, antioxidant and anti-inflammatory activities of aerial parts extract from Algerian *Artemisia Herba-alb*. *Journal of Pharmaceutical Research* 10(1): 58-64 (2016).
 107. Akrou, A., H. El Jani, S. Amouri, & N. Neffati. Screening of antiradical and antibacterial activities of essential oils of *Artemisia*

- campestris* L., *Artemisia herba Alba* asso, & *Thymus capitatus* Hoff. ET link. Growing wild in the southern of Tunisia. *Recent Research in Science and Technology* 2(1): 29–39 (2010).
108. Twaij, H.A.A. & A.A. Al-Badr. Hypoglycemic activity of *Artemisia herba-alba*. *Journal of Ethnopharmacology* 24(2-3): 123-126 (1988).
 109. Al-Shamaony, L.A., S.M. Al-Khazraji, & H.A.A. Twaij. Hypoglycaemic effect of *Artemisia herba alba* .II. Effect of a valuable extract on some blood parameters in diabetic animals. *Journal of Ethnopharmacology* 43(3): 167-171 (1994).
 110. Makhloufi, A., A. Bouyahyaoui, N. Seddiki, L. Benlarbi, L. Mebarki, & A. Boulanouar. Phytochemical Screening and Anti-Listerial Activity of Essential Oil and Crude Extracts from Some Medicinal Plants Growing Wild in Bechar (South West of Algeria). *International Journal of Phytotherapy* 4(2): 95-100 (2014).
 111. Nofal, S.M., S.S. Mahmoud, A. Ramadan, G.A. Soliman, & R. Fawzy. Anti-diabetic effect of *Artemisia Judaica* extracts. *Research journal of Medical Sciences* 4(1): 42-48 (2009).
 112. Emami, S.A., N.V. Mashhadian, R. Vosough, & B. Mohammad Oghazian. The anticancer activity of five species of *Artemisia* on Hep2 and HepG2 cell lines. *Pharmacologyonline* 3: 327-339 (2009).
 113. Nahrevanian, H., B.S.K. Milan, M. Kazemi, R.H. Hosseini, S.S. Mashhadi, & S. Nahrevanian. Antimalarial effects of Iranian flora *Artemisia sieberi* on *Plasmodium berghei* in vivo in mice and photochemistry analysis of its herbal extracts. *Malaria Research and Treatment* 727032: 8 (2012).
 114. Akkawi, M., Q. Aburemeleh, S. Jaber, M. Qutob, & P. Lutgen. The effect of *Artemisia sieberi* extracts on the Formation of β -Hematin. *British Journal of Pharmacology and Toxicology* 5(1): 49-54 (2014)
 115. Irshaid, F., K. Mansi, & T. Aburjai. Antidiabetic effect of essential oil from *Artemisia sieberi* growing in Jordan in normal and an alloxan induced diabetic rats. *Pakistan Journal of Biological Science* 13(9): 423-430 (2010).
 116. Taherkhani, M., A. Rustaiyan, H. Nahrevanian, S. Naeimi, & T. Taherkhani. Comparison of antimalarial activity of *Artemisia turanica* extract with current drugs in vivo. *Journal of Vector Borne Disease* 50(1): 51–56 (2013).
 117. Akrouf, A., H. Mighri, M. Krid, F. Thabet, H. Turki, H. El-Jani, & M. Neffati. chemical composition and antioxidant activity of aqueous extracts of some wild medicinal plants in southern Tunisia. *International Journal of Life Science and Medical Science* 2(1): 1-4 (2012).
 118. Sefi, M., H. Fetouri, M. Makni, & N. Zeghal. Mitigating effects of antioxidant properties of *Artemisia campestris* leaf extract on hyperlipidemia, advanced glycation end products and oxidative stress in alloxan-induced diabetic rats. *Food Chemistry and Toxicology* 48(7): 1986-1993 (2010).
 119. Djidel, S. & S. Khennouf. Radical scavenging, reducing power, lipid peroxidation inhibition and chelating properties of extracts from *Artemisia campestris* L. aerial part. *Annual Research & Reviews in Biology* 4(10): 1691-1702 (2014).
 120. El Abed, N., F. Guesmi, M. Mejri, M.N. Marzouki, J. Ben Had, & S. Ahmed. Phytochemical screening and assessment of antioxidant, antibacterial and cytotoxicity activities of five Tunisian medicinal Plants. *International Journal of Pharma Research & Biological Science* 3(4): 770-789 (2014).
 121. Naili, M.B., R.O. Alghazeer, N.A. Saleh, & A.Y. Al-Najjar. Evaluation of antibacterial and antioxidant activities of *Artemisia campestris* (Astraceae) and *Ziziphus lotus* (Rhamnaceae). *Arab Journal of Chemistry* 3(2): 79–84 (2010).
 122. Aicha, N., S. Ines, B.S. Mohamed, B. Ines, K. Soumaya, G. Kamel, N. Mohamed, & C. Imed. Chemical composition, mutagenic and antimutagenic activities of essential oils from (Tunisian) *Artemisia campestris* and *Artemisia herba-alba*. *Journal of Essential Oil Research* 20(5): 471-477 (2008).
 123. Hamed, B.N., H.T. Serria, M. Lobna, Z. Khaled. Aqueous leaves extract of *Artemisia campestris* inhibition of the scorpion venom induced hypertension. *Journal of Medicinal Plant Research* 8(13): 538-542 (2014).
 124. Sefi, M., H. Fetoui, N. Lachkar, A. Tahraoui, B. Lyoussi, T. Boudawara, & N. Zeghal. *Centaurium erythraea* (Gentianaceae) leaf extract alleviates streptozotocin-induced oxidative stress and β -cell damage in rat pancreas. *Journal of Ethnopharmacology* 135(2): 243-250 (2011).
 125. Benjumea, D., S. Abdala, F. Hernandez-Luis, P. Perez-Paz, & D. Martin-Herrera. Diuretic activity of *Artemisia thuscula*, an endemic canary species. *Journal of Ethnopharmacology* 100(1-2): 205–209 (2005).
 126. Afsar, S.K., K.R. Kumar, J.V. Gopal, & P. Raveesha. Assessment of anti-inflammatory activity of *Artemisia vulgaris* leaves by cotton pellet granuloma method in Wistar albino rats. *Journal of Pharmacy Research* 7(6): 463-467 (2013).
 127. World Economic Forum. *Guidelines for Employer-Based Malaria Control Programmes*. Global Health Initiatives and Harvard School of Public Health, Massachusetts, USA (2006).

128. Enserink, M. Epidemiology: Lower malaria numbers reflect better estimates and a glimmer of hope. *Science* 321(5896): 1620 (2008).
129. Lin, J., J.J. Juliano, & C. Wongsrichanalai. Drug-resistant malaria: the era of ACT. *Current Infectious Disease Reports* 12(3): 165–173 (2010).
130. Bonnet, S., L. Gouagna, R. Paul, I. Safeukui, & J. Meunier. Estimation of malaria transmission from humans to mosquitoes in two neighbouring villages in South Cameroon: Evaluation and comparison of several indices. *Trans of Royal Society of Tropical Medicine & Hygiene* 97(1): 53-59 (2003).
131. Dhangadamajhi, G., K. Kumar, & H. Ranjit. The survival strategies of malaria parasite in the red blood cell and host cell polymorphisms. *Malaria Research & Treatment* Article ID-973094 (2010).
132. Batista, R., A. de Jesus Silva Junior, & A.B. de Oliveira. Plant-derived antimalarial agents: new leads and efficient phytomedicines. Part II. Non-alkaloidal natural products. *Molecules* 14(8): 3037-3072 (2009).
133. Dondorp, A.M., F. Nosten, Y. Poravuth, D. Das, A.P. Phyto, J. Tarning, K.M. Lwin, F. Ariey, W. Hanpithakpong, S.J. Lee, P. Ringwald, K. Silamut, M. Imwong, K. Chotivanich, P. Lim, T. Herdman, S.S. An, S. Yeung, P. Singhasivanon, N.P. Day, N. Lindegardh, & D. Socheat. Artemisinin resistance in *Plasmodium falciparum* malaria. *New England Journal of Medicine* 361: 455–467 (2009).
134. Hsu, E. The history of qinghao in the Chinese materia medica. *Trans of Royal Society of Tropical Medicine and Hygiene* 100: 505-508 (2006).
135. Covello, P.S. Making artemisinin. *Phytochemistry* 69(17): 2881-2885 (2008).
136. Aryanti, B.M., T.M. Ermayanti, & I. Mariska. Production of antileukemic agent in untransformed and transformed root cultures of *Artemisia cina*. *Annales Bogorenses* 8(1): 11-16 (2001).
137. Rashmi, R.T., S.M. Francis, & S. Murali. Determination of Artemisinin in Selected *Artemisia* L. species by HPLC. *Indo American Journal of Pharmaceutical Research* 4(5): 2637-2644 (2014).
138. Arab, H.A., S. Rahbari, A. Rassouli, M.H. Moslemi, & F.D.A. Khosravirad. Determination of artemisinin in *Artemisia sieberi* and anticoccidial effects of the plant extract in broiler chickens. *Tropical Animal Health & Production* 38(6): 497-503 (2006).
139. Zia, M., M. Abdul, & M.F. Chaudhary. Effect of growth regulators and amino acids on artemisinin production in the callus of *Artemisia absinthium*. *Pakistan Journal of Botany* 39: 799-805 (2007).
140. Mannan, A., N. Shaheen, W. Arshad, R.A. Qureshi, M. Zia, & B. Mirza. Hairy roots induction and artemisinin analysis in *Artemisia dubia* and *Artemisia indica*. *African Journal of Biotechnology* 7(18): 3288-3292 (2008)
141. Mutabingwa, T.K. Artemisinin-based combination therapies (ACTS): Best hope for malaria treatment but inaccessible to the needy. *Acta Tropica* 95(3): 305-315 (2005).
142. Li, J. and B. Zhou. Biological actions of artemisinin: Insights from medicinal chemistry studies. *Molecules* 15(3): 1378-1397 (2010).
143. Utzinger, J., S.H. Xiao, M. Tanner, & J. Keiser. Artemisinins for schistosomiasis and beyond. *Current Opinion in Investigating Drugs* 8(2): 105-116 (2007).
144. Sen, R., S. Bandyopadhyay, A. Dutta, G. Mandal, S. Ganguly, P. Saha, & M. Chatterjee. Artemisinin triggers induction of cell-cycle arrest and apoptosis in *Leishmania donovani* promastigotes. *Journal of Medical Microbiology* 56(Pt9) 1213-1218 (2007).
145. Dunay, I.R., W.C. Chan, R.K. Haynes, & L.D. Sibley. Artemisone and artemiside control acute and reactivated toxoplasmosis in a murine model. *Antimicrobial Agents in Chemotherapy* 53(10): 4450-4456 (2009).
146. Dellicour, S., S. Hall, D. Chandramohan, & B.D. Greenwood. The safety of artemisinins during pregnancy: a pressing question. *Malaria Journal* 6: 15 (2007).
147. Petrus, A.J.A. & R.T. Seetharaman. Antioxidant flavone c-biosides from the aerial parts of *Alternanthera pungens*. *Indian Journal of Pharmaceutical Science* 67(2): 187 (2005).
148. Cubuku, B., H.D. Bray, C.D. Warhurst, H.A. Mericli, N. Ozhatay, & G. Sariyar. Invitro antimalarial activity of crude extracts and compounds from *Artemisia abrotanum* L. *Phytotherapy Research* 4(5): 203-204 (1990).
149. Kodippilli, K., D.W. Ratnasooriya, S. Premakumara, & V.P. Udagam. An investigation of the antimalarial activity of *Artemisia vulgaris* leaf extract in rodent malaria model. *International Journal of Green Pharmacy* 5(4): 276-281 (2011).
150. Valecha, N., S. Biswas, V. Badoni, K.S. Bhandari, & O.P. Sati. Antimalarial activity of *Artemisia japonica*, *Artemisia maritima* and *Artemisia nilegarica*. *Indian Journal of Pharmacology* 26(2): 144-146 (1994).
151. Ene, A.C., S.E. Atawodi, & H.O. Khwanashie. Bioassay-guided fractionation and invivo antiplasmodial effect of fractions of chloroform extracts of *Artemisia maciverae* Linn. *Acta Tropica* 112(2): 288-294 (2009).

152. Tasdemir, D., M. Tierney, R. Sen, M.C. Bergonzi, B. Demirci, A.R. Bilia, K.H. Can Baser, R. Brun, & M. Chatterjee. Antiprotozoal effect of *Artemisia indica* extracts and essential oil. *Planta Medica* 81(12/13): 1029-1037 (2015).
153. Emami, S.A., S.Z.T. Rabe, & M. Mahmoudi. Inhibitory effect of eleven *Artemisia* species from Iran against *Leishmania major* parasites. *Iran Journal of Basic Medical Science* 15(2): 807-11 (2012).
154. El-Naili, M.A., N.A. Saleh, H.I. Ahmed, B.K. Rammash, & A.A. El-Buni. *Journal of Egypt Academic Society of Environmental Development* 9: 1 (2008).
155. Jebri, A.O. A Screening Study on the Antimicrobial and Hepatoprotective Effects of Some Medicinal Plants in Libya. MSc. Thesis, Faculty of Science, Al-Fateh University, Tripoli Libya (2008).
156. Benedek, B., B. Kopp, & M.F. Melzig. *Achillea millefolium* L. is antiinflammatory activity mediated by protease inhibition? *Journal of Ethnopharmacology* 113(2): 312-317 (2007).
157. Esra, K., A. Tuson, & E. Yesilada. *Journal of Ethnopharmacology* 113: 332-337 (2007).
158. Harrison, A.P. & E.M. Bartels. A modern appraisal of ancient Etruscan herbal practices. *American Journal of Pharmacology and Toxicology* 1(2): 26 (2006).
159. Alonso-Paz, E., M.P. Cerdeiras, J. Fernandez, F. Ferreira, P. Moyna, M. Soubes, A. Vazquez, S. Veros, & L. Zunno. Screening of Uruguayan medicinal plants for antimicrobial activity. *Journal of Ethnopharmacology* 45(1): 67-70 (1995).
160. Rojas, R., B. Bustamante, & J. Bauer. Antimicrobial activity of selected Peruvian medicinal plants. *Journal of Ethnopharmacology* 88(2-3): 199-204 (2003).
161. Benkeblia, N. Antimicrobial activity of essential oil extracts of various onions (*Allium cepa*) and Garlic (*Allium sativum*). *LWT. Food Science & Technology* 37(2): 263-268 (2004).
162. Cowan, M.M. Plant products as antimicrobial agents. *Clinical Microbiology Review* 12(4): 564-582 (1999).
163. Nostro, A.M.P., A. Germano, V. D'Angelo, A. Marino, & M.A. Cannatelli. Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity. *Letters in Applied Microbiology* 30(5): 379-84 (2000).
164. Hui, M.E., H.H. Cheng, & A.K. Radhakrishnan. Anti-Proliferative and mutagenic activities of aqueous and methanol extracts of leaves from *Pereskia bleo* (Kunth) DC (Cactacea). *Journal of Ethnopharmacology* 113(3): 448-456 (2007).
165. Ferrero, A., A. Menitti, C. Bras, & N. Zanetti. Acute and subacute toxicity evaluation of ethanolic extract from fruits of *Schinus molle* in rats. *Journal of Ethnopharmacology* 113(3): 441-7 (2007).
166. Ivanovska, N., S. Philipov, R. Istatkova, & P. Georgieva. Antimicrobial and immunological activity of ethanol extracts and fractions from *Isopyrum thalictroides*. *Journal of Ethnopharmacology* 54(2-3): 143-151 (1996).
167. Taylor, R.S., F. Edel, N.P. Manandhar, & G.H. Towers. Antimicrobial activities of southern Nepalese medicinal plants. *Journal of Ethnopharmacology* 50(2): 97-102 (1996).
168. Sukanya, S.L., J. Sudisha, P. Hariprasad, S.R. Niranjana, H.S. Prakash, & S.K. Fathima. Antimicrobial activity of leaf extracts of Indian medicinal plants against clinical and phytopathogenic bacteria. *African Journal Biotechnology* 8(23): 6677-6682 (2009).
169. Ramezani, M., B.S. Fazli-Bazzaz, F. Saghafi-Khadem, & A. Dabaghian. Antimicrobial activity of four *Artemisia* species of Iran. *Fitoterapia* 75(2): 201-203 (2004).
170. Shoko, T., T. Soichi, M.M. Megumi, K. EriJun, & Michiko W. Isolation and identification of an antibacterial compound from grape and its application to foods. *Nippon Nogeikagaku Kaishi* 73: 125-128 (1999).
171. Asghari, G., M. Jalali, & E. Sadoughi. Antimicrobial activity and chemical composition of essential oil from the seed of *Artemisia aucheri* Boiss. *Jundishapur Journal of Natural Pharmaceutical Product* 7(1): 11-5 (2012).
172. Cetin, B., H. Ozer, A. Cakir, E. Mete, M. Tosun, E. Ozturk, T. Polat, & A. Kandemir. Chemical composition of hydro distilled essential oils of *Artemisia incana* (L.) DRUCE and antimicrobial activity against foodborne microorganisms. *Chemistry & Biodiversity* 6(12): 2302-2310 (2009).
173. Lambert, R.J.W., P.N. Skandamis, P. Coote, & G.L.E. Nychas. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol. *Journal of Applied Microbiology* 91(3): 453-462 (2001).
174. Ultee, A., E.P.W. Kets, & E.J. Smid. Mechanisms of action of carvacrol on the food-borne pathogen *Bacillus cereus*. *Applied Environmental Microbiology* 65(10): 4606-4610 (1999).
175. Kraus, W. Source of unique natural products for integrated pest management, medicine, industry and other purposes. In: *The Neem Tree*, Schmutterer, H. (Ed.). p. 35-88 (1995).
176. Shinwari, Z.K. Medicinal plants research in Pakistan. *Journal of Medicinal Plant Research* 4(3): 161-176 (2010).

177. Koyama, J., I. Morita, I. Kobayashi, N. Hirai, K. Simamura, E. Nobukawa, & T. Kadota. Antiallergic activity of aqueous extracts and constituents of *Taxus yunnanensis*. *Biological & Pharmaceutical Bulletin* 29(11): 2310-2312 (2006)
178. Shamim, S., N. Farzananeh, M. Mahmoud, D. Elaheh, C.Y. Hoong, & N.R. Abdullah. Cytotoxic activity of some medicinal plants from Iran. *Ethno Medicine* 3(1): 81-82 (2009).
179. Mesia, G.K., G.L. Tona, T.H. Nangaa, R.K. Cimanga, S. Apers, P. Cosc, L. Maesc, L. Pieter, & A.J. Vlietinck. Antiprotozoal and cytotoxic screening of 45 plant extracts from Democratic Republic of Congo. *Journal of Ethnopharmacology* 115(3): 409-415 (2008).
180. Singh, N.P., & H.C. Lai. Artemisinin induces apoptosis in human cancer cells. *Anticancer Research* 24(4): 2277-2280 (2007).
181. Willoughby Sr, J.A., S.N. Sundar, M. Cheung, A.S. Tin, J. Modiano, & G.L. Firestone. Artemisinin blocks prostate cancer growth and cell cycle progression by disrupting Sp1 interactions with the cyclin-dependent kinase-4 promoter and inhibiting CDK4 gene expression. *The Journal of Biological Chemistry* 284(4): 2203-13 (2009).
182. Mercer, A.E., J.L. Maggs, X.M. Sun, G.M. Cohen, J. Chadwick, P.M. O'Neill, & B.K. Park. Evidence for the involvement of carbon-centered radicals in the induction of apoptotic cell death by artemisinin compounds. *The Journal of Biological Chemistry*. 282: 9372-9382 (2007).
183. Lai, H. & N.P. Singh. Oral artemisinin prevents and delays the development of 7, 12 dimethylbenz[a]anthracene (DMBA)-induced breast cancer in the rat. *Cancer Letters* 231(1): 43-8 (2006).
184. Efferth, T., H. Dunstan, A. Sauerbrey, H. Miyachi, C.R. Chitambar. The anti-malarial artesunate is also active against cancer. *International Journal of Oncology* 18(4): 767-773 (2001).
185. Wang, Q., L. Wu, A. Li, Y. Zhao, & N.P. Wang. Experimental studies of antitumor effect of artesunate on liver cancer. *Zhongguo Zhong Yao Za Zhi* 26(10): 707-720 (2001).
186. Li, P.C., E. Lam, W.P. Roos, M.Z. Zdzienicka, B. Kaina, & T. Efferth. Artesunate derived from traditional Chinese medicine induces DNA damage and repair. *Cancer Research* 68(11): 4347-4351 (2008).
187. Aberham, A., S.S. Cicek, P. Schneider, & H. Stuppner. Analysis of sesquiterpene lactones, lignans, and flavonoids in wormwood (*Artemisia absinthium* L.) using high-performance liquid chromatography (HPLC)-mass spectrometry, reversed phase HPLC, and HPLC-solid phase extraction-nuclear magnetic resonance. *Journal of Agriculture & Food Chemistry* 58(20): 10817-10823 (2010).
188. Govindaraj, S., B.D.R. Kumari, P.L. Cioni, & G. Flamini. Mass propagation and essential oil analysis of *Artemisia vulgaris*. *Journal of Biosciences & Bioengineering* 105(3): 176-183 (2008).
189. Lopes-Lutz, D., D.S. Alviano, C.S. Alviano, & P.P. Kolodziejczyk. Screening of chemical composition, antimicrobial and antioxidant activities of *Artemisia* essential oils. *Phytochemistry* 69(8): 1732-1738 (2008)
190. Khan, A.U. & A.H. Gilani. Antispasmodic and bronchodilator activities of *Artemisia vulgaris* are mediated through dual blockade of muscarinic receptors and calcium influx. *Journal of Ethnopharmacology* 126(3): 480-486 (2009).
191. Lee, H.G., H. Kim, W.K. Oh, K.A. Yu, Y.K. Choe, J.S. Ahn, S.H. Kim, C.A. Dinarello, K. Kim, & D.Y. Yoon. Tetramethoxyhydroxy flavone p7F down regulates inflammatory mediators via the inhibition of nuclear factor kappa B. *Annals of the New York Academy of Science* 1030: 555-568 (2004).
192. Gilani, A.U.H. and K.H. Janbaz. Preventive and curative effects of *Artemisia absinthium* on acetaminophen and CCl4-induced hepatotoxicity. *General Pharmacology* 26(2): 309-315 (1995).
193. Hitosugi, N., R. Ohno, I. Hatsukari, S. Nakamura, S. Mizukami, H. Nagasaka, I. Matsumoto, K. Satoh, T. Negoro, K. Hashimoto, & H. Sakagami. Induction of cell death by pro-oxidant action, of Moxa smoke. *Anticancer Research* 22(1A): 159-163 (2002).
194. Shoemaker, M., B. Hamilton, S.H. Dairkee, I. Cohen, M.J. Campbell. In vitro anticancer activity of twelve Chinese medicinal herbs. *Phytotherapy Research* 19(7): 649-651 (2005).
195. Kim, M.J., D.H. Kim, N.H. Na, T.Y. Oh, C.Y. Shin, & Y.J. Surh. Eupatilin, a pharmacologically active flavone derived from *Artemisia* plants, induces apoptosis in human gastric cancer (AGS) cells. *Journal of Environmental Pathology Toxicology & Oncology* 24(4): 261-269 (2005).
196. Adams, M., T. Efferth, & R. Bauer. Activity-guided isolation of scopoletin and isoscapoletin, the inhibitory active principles towards CCRF-CEM leukaemia cells and multi-drug resistant CEM/ADR5000 cells, from *Artemisia argyi*. *Planta Medica* 72(9): 862-864 (2006).
197. Najaran, T.Z., M. Sareban, A. Gholami, A. Emami, & M. Mojarab. Cytotoxic and apoptic effects of different extracts of *Artemisia turanica* krasch On K562 and HL-60 cell lines. *The scientific World Journal* 628073: 6p (2013).

198. Rabe, T.Z.S., M. Mahmoudi, A. Ahi, & A.S. Emami. Antiproliferative effects of extracts from Iranian *Artemisia* species on cancer cell lines. *Pharmaceutical Biology* 49(9): 962-9 (2011).
199. Lorenzi, H. & F.J.A. Matos. Plantas medicinais no Brasil: nativas e exóticas (2thed.). São Paulo: Instituto. *Plantarum* 11(25): 118–121 (2008).
200. Blagojevic, P., N. Radulovic, R. Palic, & G. Stojanovic. Chemical composition of the essential oils of Serbian wild-growing *Artemisia absinthium* and *Artemisia vulgaris*. *Journal of Agriculture and Food Chemistry* 54(13): 4780–4789 (2006).
201. Dhar, D.N., R.L. Sharma, G.C. Bansal. Gastrointestinal nematodes in sheep in Kashmir. *Veterinary Parasitology* 11(2-3): 271-7 (1982).
202. Cala, A.C., J.F.S. Ferreira, A.C.S. Chagas, et al. Anthelmintic activity of *Artemisia annua* L. extracts in vitro and the effect of an aqueous extract and artemisinin in sheep naturally infected with gastrointestinal nematodes. *Parasitology Research* 113(6): 2345 (2014).
203. Seddiek, S.A., M.M. Ali, H.F. Khater, & M.M. El-Shorbagy. Anthelmintic activity of the white wormwood, *Artemisia herba-alba* against *Heterakis gallinarum* infecting Turkey poults. *Journal of Medicinal Plant Research* 5(16): 3946-3957 (2011).
204. Nakhare, S. & S.C. Garg. Anthelmintic activity of the essential oil of *Artemisia pallens* Wall. *Ancient Science of Life* 10(3): 185-186 (1991).
205. Singhal, K.C. Anthelmintic activity of *Punica granatum* and *Artemisia siversiana* against experimental Infections in mice. *Indian Journal of Pharmacology* 15(2): 119-122 (1983).
206. Bezanger-Beauquesne, L., M. Pinkas, M. Torck, & F. Trotin. Les plantes medicinales des regions temperées. Maloine. Paris (1990).
207. Akhtar, M.S., M.I. Chattha, & A.H. Chaudhry. Comparative efficacy of santonin and piperazine against *Neoscaris vitulorum* in buffalo calves. *Journal of Veterinary Pharmacology & Therapy* 5(1): 71-76 (1982)
208. Proksch, P. *Artemisia herba-alba*. In: *Artemisia*, C.W. Wright (Ed.). Taylor & Francis, London (series Medicinal and Aromatic Plants—Industrial Profiles), p. 99–105 (2002).
209. Irum, S., H. Ahmed, M. Mukhtar, M. Mushtaq, K. Donskow-Lysoniewska, M. Qayyum, & S. Simsek. Anthelmintic activity of *Artemisia vestita* Wall ex DC. And *Artemisia maritima* L. against *Haemonchus contortus* from sheep. *Veterinary Parasitology* 212(3-4): 451–455 (2015).
210. Tariq, K.A., M.Z. Chishti, F. Ahmad, & A.S. Shawl. Anthelmintic activity of extracts of *Artemisia absinthium* against ovine nematodes. *Veterinary Parasitology* 160(1-2): 83–88 (2009).
211. Shen, X.L., M. Nielsen, M.R. Witt, O. Sterner, O. Bergendorff, & M. Khayyal. Inhibition of [methyl-3H] diazepam binding to rat brain membranes in vitro by dinatin and skrofulein. *Acta Pharmacologica Sinica* 15(5): 385-8 (1994).
212. Iqbal, Z., M. Lateef, M. Ashraf, & A. Jabbar. Anthelmintic activity of *Artemisia brevifolia* in sheep. *Journal of Ethnopharmacology* 93(2-3): 265–268 (2004).
213. Irum, S., H. Ahmed, & B. Mirza, K. Donskow-Lysoniewska, A. Muhammad, M. Qayyum, S. Simsek. In vitro and in vivo anthelmintic activity of extracts from *Artemisia parviflora* and *A. sieversiana*. *Helminthologia* 54: (3) 218–224 (2017).
214. Beshay, E.V.N. Therapeutic efficacy of *Artemisia absinthium* against *Hymenolepis nana*: in vitro and in vivo studies in comparison with the anthelmintic praziquantel. *Journal of Helminthology* 2017 Jun 13:1-11 (2017), doi: 10.1017/S0022149X17000529.