



Assessment of Soil Chemical Properties for Monitoring and Maintenance of Soil Fertility in Probolinggo, Indonesia

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Abstract: Soil is paramount to sustaining living in biomass production, water quality control, climatic mitigation, and biodiversity endurance. Closely associated with sustainable agriculture, it degrades soil in the long run, robbing the soil of its production capacity and food-generating ability. In Probolinggo, a regency in Indonesia, intensifying the use of chemical fertilizers and pesticides yet a declining trend in yield production was discovered. This research analyzed the acid, nitrogen, organic carbon, and nutrients focusing on phosphor, potassium, iron, and manganese contents. Organic carbon/nitrogen ratio, soil organic compound rate, and cation exchange capacity were also discussed in order to illustrate the correlations among chemical substances and their roles in soil and plant maintenance. While such a study has yet to be performed in Probolinggo, the results should show the degree of land deterioration and future attempts at damage control and correction open to facilitate. Employing a simple random method, soil and plant samples were collected from 18 villages in six districts and their chemical contents were compared to the standard set in Government Regulations No 150/2000. The results showed low N-total, P-Bray, P-Olsen, K, C-Organic, and C/N ratio availabilities at 0.18, 13.88, 14.41, 0.37, 1.36, and 7.38 respectively, contrasted to high rates on pH (5.94), Fe (153.46 mg kg⁻¹) and Mn (37.96 mg kg⁻¹). Biomass production is conclusively imperative to fix the land composition and meet the plant nutrient requirements through an organic approach; fertilizers from digester biogas are therefore recommended. This action requires field agricultural advisors to raise awareness of sustainable agriculture.

Keywords: Environmentally Friendly, Organic Approach, Soil Deterioration, Soil Fertility Evaluation, Sustainable Development Goals, Sustainable Farming

1. INTRODUCTION

A natural body as the result of complex biogeochemical and physical processes, soil is imperative in sustaining the living [1, 2]. Not only limited to plant substrate, but its role also expands as far as an exponential part of biodiversity by regulating the water cycle and nutrients [3, 4]. Vastly

contributing to the ecosystem, soil is crucial to sustainable development goals (SDGs) concerning biomass production, water quality control, climatic mitigation, and biodiversity endurance [5, 2]. Quality soil holds the potential to solve hunger and poverty issues while retaining robust health [6, 7] key to welfare, making it the most valuable asset of a nation [4].

While soil health is associated with sustainable agriculture [8, 9], it degrades soil in the long run due to supplying food for the ever-increasing human population [10, 11], robbing the soil of its production capacity and food-generating ability [11, 12]. Agricultural practices deprive soil properties [2, 13] have been reported in Uruguay after an agricultural production period between 2012 and 2014 [14], in Iran after forestland conversion to tea plantation [15], and in Italy where food self-sufficiency plummeted to less than 80 % [12].

Out of various agricultural practices and land exploitations, monocropping is one of the most detrimental methods toward soil quality [16–18]. Another cause is low irrigation water quality [19, 20]. Pesticides are also liable for affecting microorganisms [21, 22], particularly in enzymes entailing carbon, nitrogen, sulphur, and phosphorus cycles [23, 24]. While fertilizers may be helpful, excessive use of chemical fertilizers reduce soil biodiversity and function [25, 26] by triggering soil acidification and soil crusting that suppress the growth of organic matters [2, 27, 28].

Running soil fertility evaluation (SFE) should be feasible to assess the degree of soil deterioration and its result should help in determining the appropriate biomass treatment for each condition – some countries like Nigeria [29], the Philippines [30], and Indonesia [31, 32] have based their fertilizer application guides on it. Analyzing the chemical properties of soil – e.g. acid, organic carbon, nutrients – is one of the attainable approaches.

Soil deterioration is a serious issue in Indonesia; since the start of modern agricultural practices in the 1970s, biomass-producing areas have been exploited all year long. In addition to monocropping, excessive pesticides on rice production in Java has been reported [33] – a certain site applied up to 24.6 kg of pesticides $\text{ha}^{-1} \text{yr}^{-1}$ [34], and farmers in another locale violated the recommended amount of chemical pesticides [35]. The preliminary survey in Probolinggo Regency, a regency in the province of East Java, Indonesia, discovered intensifying use of chemical fertilizers and pesticides yet a declining trend in yield production.

Soil quality studies had been performed in a number of areas in Indonesia, among them are of apple plantations [36], cassava fields [37, 38], paddy fields [39, 40] and post-paddy fields [41], oil palm plantations [2, 42–46], rubber [47], sugarcane [48–50], even volcanic soils [51] and drylands [52–54]. A few researchers had also compared different fields at once in Bengkulu [55] and West Java [56]. However, not one of the such studies had been run in Probolinggo Regency. The aforementioned researches mostly put N, P, and K contents as their parameters – only some of them discussed about the C-organic content, pH rate, and micronutrient availability. Further, studies on Mn, Fe, SOC, and CEC contents in Indonesia are scarce. The case in the surveyed area calls for a more thorough soil quality study for biomass production to detect any occurrence of soil deterioration. In addition to the prevalently analyzed factors, the infrequently examined ones are put into consideration. Further, in the soil, the chemicals N, P, and K contained in a number of local commodities are also scrutinized.

2. MATERIALS AND METHODS

2.1 Location

Conducted in Probolinggo Regency, East Java, Indonesia, in 2020, a number of six districts – with three rural villages in each to make 18 in total – were randomly appointed to represent the regency: Krucil District (Krucil Village, Breml Village, and Betek Village), Tiris District (Ranu Agung Village, Tiris Village, and Andungsari Village), Gading District (Mojolegi Village, Wangkal Village, and Kaliacar Village), Pajarakan District (Karangpranti Village, Pajarakan Kulon Village, and Sukomulyo Village), Krejengan District (Kedung Caluk Village, Sumber Kalimoho Village, Seboro Village), and Kraksaan District (Semampir Village, Sidopekso Village, and Kregen Village) (Figure 1). The GPS details for the six districts are listed in Table 2.

Each district has its type of soil. While Pajarakan is of regosol and alluvial, Kraksaan soil is dominated by alluvial with a small part of andosol. Krejengan has equal portions of regosol and alluvial. Regosol takes most of Gading, leaving a little for andosol. Tiris is mostly regosol with a latosol touch, and Krucil is mostly regosol with bits

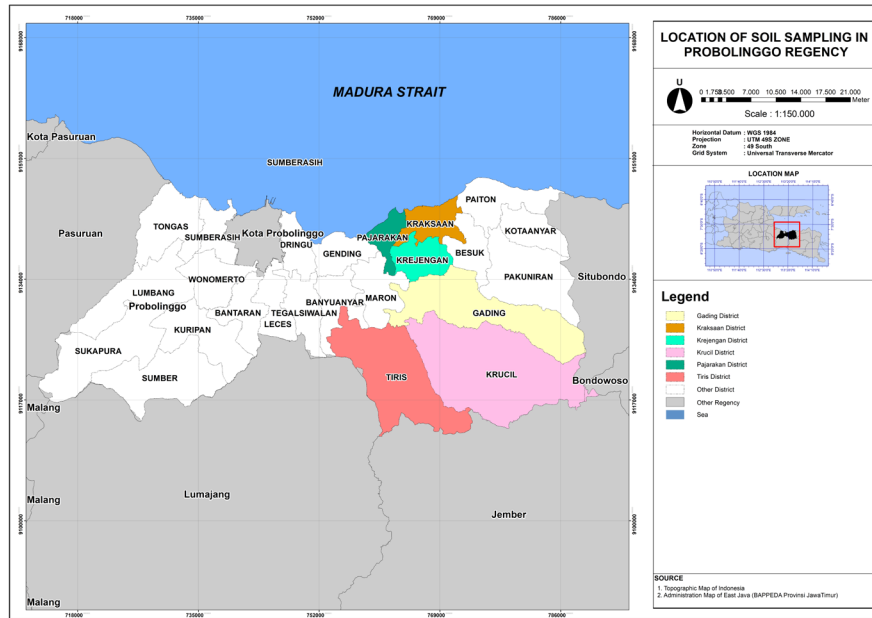


Fig. 1. Location soil sampling (Krucil District, Tiris District, Gading District, Pajarakan District, Krejengan District, and Kraksan District)

of latosol dan andosol.

Average rainfall from 2015 to 2020 at six soil sampling locations as follows: Pajarakan (119 mm), Kraksaan (144 mm), Krejengan (155 mm), Gading (108 mm), Tiris (133 mm), and Krucil (78 mm). According to Schmidt-Ferguson, the climate types and Q are Pajarakan (140.00 % - E), Kraksaan (120.00 % - E), Krejengan (120.00 % - E), Gading (83.33 % -D), Tiris (133.64 % - E), and Krucil (100.00 % - D). As a complement, cropping system data at six research locations are Pajarakan (1x paddy, corn), Kraksaan (1x paddy, corn), Krejengan (1x paddy -100 %, 2x -30 % paddy, corn, soybean), Gading (paddy 1x), Tiris (paddy 1x), and Krucil (paddy 2x).

Referring to Government Regulation No. 150/2000 that the solum critical benchmark is ≤ 20 cm, only six locations are ≥ 50 cm. As no soil crust was encountered, all districts were considered secure at this point. A series of the preliminary survey was conducted to identify soil characteristics in each village. Then, soil and plant samples were gathered for chemical analyses.

2.2 Materials

Soil and plant sample selection were of simple random sampling, taken from three different sites

in each village at any time during the research period. Once a plant was chosen, the area around it was leveled and cleaned from grass and rubbles, then dug as deep as 5 cm to 20 cm to take the soil sample. A lead tube was probed into the ground with the help of a wooden block until three-quarters part of it was buried. Another tube was pressed inside the first one at 1 cm deep to compact the dirt and separated it. The tube was then dug out of the ground along with some extra dirt to protect it from direct contact with the shovel. Excessive dirt on the upper end of the tube was carefully leveled and then covered with a plastic lid. The lower end received the same handling. The top lid was then labeled with information on gaining depth, date, and location.

2.3 Methods

Both soil and plant were tested for their acid (pH), organic carbon (C-organic), and nutrient (N, P, and K) contents. Acidity was measured with a pH meter. Organic carbon content was assessed on Walkey and Black method. As for nutrient content, N was calculated by employing the Kjeldahl method and P by Olsen and Bray, while K, Fe, and Mn with Atomic Absorption Spectrophotometry (AAS).

All results were then compared to the parameters set by the Indonesian government as declared in the

Government Regulations No 150/2000 on Land Deterioration Management for Biomass Production [57] – except for Fe which went with the standard set by the Soil Research Center of the Ministry of Agriculture [58] – to determine the land condition as listed in Table 1.

Table 1. Soil fertility scales based on chemical aspects

	Scale		
	Low	Medium	High
N-total	0.2	0.2 to 0.5	0.5
P-Bray2 (ug g ⁻¹)	20	20 to 40	40
K (me 100 g ⁻¹)	0.3	0.3 to 0.6	0.6
Mn (me100 g ⁻¹)	0.2	0.2 to 0.6	0.6
C-organic (%)	4	4 to10	10
Fe (mg kg ⁻¹)	3	5	9

Source: [57, 58]

Organic carbon/nitrogen ratio (C/N ratio), soil organic compound (SOC) rate, and cation exchange capacity (CEC) were also discussed to illustrate the correlations among chemical substances and their roles in soil and plant maintenance.

3. RESULTS AND DISCUSSIONS

3.1 Chemical Properties in Soil

The chemical properties in soil represent its ability to provide balanced nutrients for plants. The test results are recorded in Table 2 below.

3.2 Acidity and CEC

Affecting a vast number of biochemical processes in soil, pH is the master variable [59]. With an average of 5.94 in the depth of 0 cm to 25 cm, the general acidity level is within the critical benchmark rates between 4.5 and 8.5. The soil of eight villages came out quite acidic at 5.91, while four villages were neutral to fairly neutral at 7.02. In four villages, the soil conditions were passable despite being acidic at 5.11 since their N, P, K, Mn, and Fe contents were enough to nourish plants.

The fertilizer administration in the study area is deduced to be the reason for low soil pH in some sampling sites. Ammonium fertilizers run through the nitrification process to form nitrate. In such process, the released H⁺ ions turn the soil more acidic [59, 60]. Phosphorus and potassium can decrease soil pH after years of use [61]. Years of

land use for agriculture typically increases H⁺ ions which, consequently, accelerate soil acidification [62]. Plants grown in acidic land are prone to stresses, such as H and Mn poisoning and nutrient deficiency [59].

Acid also affects microbial diversity in soil [63, 64] and its presence indicates nutrient availability [66]. Neutral pH expands CEC, optimizing P and other nutrient contents in soil [6, 22, 66]. Out of all samples, the lowest rate was 10.49 me gr⁻¹ while the highest was 55.06 me gr⁻¹ – the ones with high CECs were taken after fertilizer application.

3.3 Organic Carbon Content, SOC, and C/N Ratio

While the C-organic should be at least 4 %, the average rate in samples was 1.35 % with the lowest quantity of 0.62 and the highest of 2.67 – these figures characterize badly-damaged soil. This finding corroborates an earlier report that more than 77 % of paddy fields in Java had low organic carbon [26, 67]. That there is inadequate vegetation due to monoculture in the research sites and that farmers barely recover soil biomass are deduced to be the reasons [2, 49, 50].

C-organic content reflects the amount of soil organic compound (SOC), another fertility factor [2, 68] and the most vital component resulting from interactions among producer, decomposer, and mineralogy [60, 70], attributable to its ability to improve soil chemical, physical, and biological characteristics [10, 28].

SOC is the key indicator of soil quality and agricultural sustainability [71]. SOC content relies heavily on nitrogen binding in soil [2, 67]. The SOC test results came out between 1.07 % and 4.62 % for all 18 samples, and seven samples reached the level of as low as 1.65 %. SOC is associated with vegetation type, hydrology, and organic matter.

Referring to the low SOC rate, it is proven that cultivation and land management can cause it, leading to land damage in the long run [71]. Disproportionate use of chemical pesticides and fertilizers suppresses the growth of nutrient-producing biota and, consequently, diminishes

Table 2. Soil chemical properties of 18 villages in six districts in Probolinggo Regency

District	Village	pH H ₂ O	pH KCl.	C- organic (%)	N-total (%)	C/N (%)	SOC (%)	P-Bray (%)	P-Olsen (%)	Available K (%)	CEC me gr ⁻¹	Fe mg kg ⁻¹	Mn mg kg ⁻¹
Pajajaran	Karang	6.1	5.2	1.22	0.16	8	2.11	22.49	-	0.17	55.06	238.10	36.70
	Pranti							-					
Kraksaan	Pajajaran	6.9	6.2	1.76	0.20	9	3.04	-	5.79	0.29	51.27	92.50	51.27
	Kulon												
Tiris	Sukomulyo	6.6	5.7	1.14	0.15	8	1.97	16.93	7.96	0.17	53.75	265.7	53.75
	Semampir	6.7	6.0	1.39	0.17	8	2.41	-	15.12	0.48	50.18	142.90	28.72
Gading	Sido Pekso	7.9	7.0	1.09	0.14	8	1.89	-	28.77	0.33	43.40	47.60	37.77
	Kregenan	6.4	5.7	1.13	0.14	8	1.95	19.59	-	0.47	48.12	190.50	47.95
Krejean	Kedung	6.1	5.2	0.75	0.11	7	1.29	37.69	-	0.60	36.58	263.70	17.58
	Caluk	6.5	5.7	0.62	0.12	5	1.07	-	-	0.49	39.61	190.50	11.03
Gading	Sumberkati												
	moho												
Tiris	Sebaroh	5.6	4.9	1.31	0.22	6	2.26	16.22	-	0.31	38.44	142.90	18.64
	Mojolegi	5.6	4.8	1.57	0.19	8	2.72	13.56	-	0.06	10.49	92.50	31.83
Krucil	Kaliacar	5.2	4.8	1.39	0.23	6	2.41	8.70	-	0.58	20.99	190.50	42.38
	Wangkal	6.0	5.2	1.47	0.19	8	2.54	3.84	-	0.46	34.37	47.60	17.47
Betek	Ranu	5.0	4.4	0.94	0.15	6	1.63	9.25	-	0.40	24.19	238.10	58.25
	Agung	5.5	4.5	1.41	0.21	7	2.43	8.78	-	0.08	15.12	142.90	22.16
Bremi	Andung	4.9	4.4	1.02	0.14	7	1.76	2.95	-	0.60	32.51	95.20	50.93
	Sari												
Average	Tiris												
	Betek	5.2	4.5	1.26	0.15	8	2.18	2.16	-	0.57	26.91	47.60	60.56
Compare Standards of Table 1	Krucil	5.6	5.0	2.26	0.24	9	3.91	10.97	-	0.28	26.38	190.50	43.39
	Bremi	5.2	4.9	2.67	0.38	7	4.62	21.18	-	0.37	27.10	142.90	52.85
		5.94	5.23	1.36	0.18	7.38	2.34	13.88	14.41	0.37	35.25	153.46	37.96
				Very low	Low			Very low		Medium		Very high	Very high

SOC [72]. Supporting the finding, applying both chemical and organic fertilizers for years in China is effective in enhancing SOC [63]. Sufficient fertilizer – combined with cattle manure – should encourage SOC content [71] and tackle the issue.

A well-balanced C-organic and nitrogen contents support microorganisms in decomposition [48, 72, 73] and the C/N ratio indicates the speed of the process [2, 74]. From the average ratio of 7.38 %, it is conclusive that the soil is not damaged. However, further treatments should be undergone to prevent it from declining – compost application is needed when the SOC drops to 11 %, and land relaxation is required if it reaches 15 %.

3.4 Nutrient Content

From Table 2, it is derived that N-total values are 0.11 % to 0.38 %, averaging 0.18 %, which is below the standard of 0.2 %. It is similar to the N-total in post-paddy fields in Labuhanbatu, Indonesia [41]. Samples from Sebaroh, Kaliacar, Andung Sari, Krucil, and Bremit Villages are medium, and none are high. The non-existent high N-total value corroborates another soil analysis on several agricultural lands in West Java [56] and adds urgency to the call for management to enhance nitrogen content in agricultural fields throughout Java recommended by a previous study [26, 75]. An essential nutrient for plants [24, 76], nitrogen determines plant growth [32, 77] and therefore significantly influential towards harvest. Nitrogen is assimilated by the plant into amino acid, the main component of protein [78] that is responsible for forming chloroplast, mitochondria, and other structures where biochemical processes in plants occur. Nitrogen is also the key element of nucleic acid, serving as genetic matter, that controls plant growth and sustainability [79]. Nitrogen deprivation leads to stunting in plants [80], so an optimal supply of nitrogen should maximize production [81].

A few features are viewed to be the causes of low N-total content in this study. The fact that tropical lands naturally contain less nitrogen has been proven by contrasting Java with Japan [75] since higher temperature accelerates organic matter decomposition [2, 28]. Improper use of pesticides in agricultural practice not only eradicates nitrogen-generating bacteria [21, 83] but also restrains the work of enzymes in the nitrogen cycle [23, 24].

As for phosphate, the contents of (2.16 to 37.69) ug g^{-1} – averaged at 6.30 ug g^{-1} – are far lower than even the low P-Bray standard of 20 ug g^{-1} . The low P contents in samples are linked to high acidity [32]. Samples from Karang Pranti, Sumber Katimoho, and Bremit are regarded as a medium, and none is high. This outcome validates the report on low P contents in West Java [56], although the same study also informed about the several sites containing high P. It contradicts the high P contents found in post-paddy fields in Labuhanbatu [41] apparently due to much P fertilizers added in every sowing period. Research held in Bogowonto Catchment, Central Java, noted higher P content in agricultural lands than in non-agricultural ones due to human intervention through fertilizers [84].

Another component in nucleic acid forming [79], along with nitrogen, phosphorus is valuable for plants [76] in ensuring sustainability for its roles in cell multiplication, respiration, and photosynthesis. Its importance spreads to energy storage and distribution via adenosine diphosphate (ADP) and adenosine triphosphate (ATP) formation [80, 85]. Energy gained from photosynthesis and carbohydrate metabolism is stored in the form of phosphate compound and later used in growth and reproduction [86], which makes this substance crucial in seed formation and root expansion [87].

The potassium (K) contents in samples span between 0.06 and 0.60, averaged at 0.17, which is below the benchmark of 0.3. Out of all tested villages, nine are medium and one is low and six are very low. A leachable material, K is easy to get washed away. It is often bound to clay and other organic substances, and it mostly gets absorbed when attached to fine soil particles. As such, K is prone to erode to rain or wind exposure [88]. Soil typically erodes more in harder rain [89] and harder wind [90]. The climate of the study area is indicated to be a cause of low K content in some sites. Another consideration is the shallow irrigation canals. Since the deeper a canal is, the more water there will be and the more K^+ ions will be absorbed [91], adding depth to those canals should help to answer the problem. Furthermore, the low potassium content is observed to be the effect of high CEC, which fortifies soil in keeping the substance within and decelerating its release. Some areas with high K contents are due to agricultural manipulation. However, continuous and excessive

use of fertilizers can wane potassium in soil [92].

A macro monovalent compound in soil, K is necessary for plant physiological processes [76] despite being excluded from basic organic compound formation since most enzymes require it as an activator [93], including protein synthesis in producing ATP [80]. K also supports the energy translocating process to various parts of the plant [93], regulates the opening and closing of stomata [80], and even strengthens the plant to resist diseases and stress [94, 95].

Iron contents are recorded at between (47.6 and 265.7) mg kg⁻¹, averaging at 153.45 mg kg⁻¹, which is high above the standard. Three villages are high and 19 are very high. Similarly, manganese contents extend over (11.03 to 60.5) mg kg⁻¹, averaged at 37.95 mg kg⁻¹ – five villages are high and 13 are very high. In addition to redox potential and pH content [96], the intensive use of inorganic fertilizers for decades has left residual buildups of Fe and Mn in soil. Mn helps to transport Fe throughout the plant [97] in binding phosphate for growth [98], and only a small amount of < 1 000 mg kg⁻¹ is required to activate protein-forming enzymes. While too little Fe and Mn lead to micronutrient deficiency [99, 98], too much of them is malicious for plants [100, 101].

The farmer's behavior in using pesticides is also influential in its production [23, 25]. In regards to controlling pests, diseases, and weeds, pesticides (as well as herbicides) have been reported to add nutrients to the soil, including Fe and Mn [102, 103]. However, their use must be controlled so as not to cause damage to the soil and the environment [21–28, 33, 35]. Several researchers, among others [28, 104], suggest using biomass ash to increase micronutrients, i.e. potassium, calcium, and magnesium. The referring to local wisdom is also recommended by some researchers [24, 83, 105–107], while the application of biological fertilizers is suggested by other researchers [22, 54, 66].

3.5 Chemical Properties in Plants

Ensuring nutrient adequacy in the soil is key to satisfying crops. The contents of N, P, and K towards cassava, coffee, corn, paddy, tobacco, shallot, chili,

and melon as local commodities were studied and then administered in Table 3.

The research reveals the N-total values of plants are ranged between 0.81 % and 3.86 %, while P is 0.03 % and 0.64 % and K is 0.4 % and 4.01 %. Conclusively, the dominant weight of the plant biomass is nitrogen, followed by phosphate and calcium respectively.

Comparing the results of the soil test (Table 2) with the plant test (Table 3), it is certain that the N and K contents in the soil are lower than the ones in the plant while the P content is higher. That plant requirement on P is lower than on N and K should be the reason. Furthermore, P is not easily soluble in water, causing it to progress slowly, since most of it is inorganic. Paddy, cassava, corn, tobacco, melon, and chili absorb higher percentages of N and K, draining them from the soil.

This study has revealed the low content of several important substances in soil. Further action should be carried out in Probolinggo Regency by decomposing all organic waste and kitchen waste into organic fertilizer, which result is then returned to agricultural lands. The decomposition of organic matter should be done anaerobically in a communal or household scale digester [108–110]. A biogas digester as such doubles the benefit for society as well as the environment by providing clean, renewable energy [111, 112] and producing two types of organic fertilizers, i.e., liquid and solid. Ideally, this biogas digester should be installed with inlet pipes from excrete disposal to septic tanks in each household [99–101]. Since there is a possibility of decomposition fluctuation due to various feedstocks, several researchers recommended a two-stage digester to overcome the problem [113–115].

Moreover, the farmers in Probolinggo should be educated to prevent excessive land management. Tilling aims to loosen the soil in preparation for seeding [116] so that the plants' roots will be able to penetrate the soil and respire more easily. However, too much handling wears soil organic matter away faster [117]. Land handling limitation in organic agriculture is the potential to improve the total stock of soil organic matter [118].

Table 3. N, P, and K contents in cassava, coffee, corn, paddy, tobacco, shallot, chili, and melon samples in Probolinggo Regency

Location	Commodity	N – Total (%)	P (%)	K (%)
<i>Pajarakan District</i>				
Karang Pranti	Corn	1.62	0.29	3.85
Pajarakan Kulon	Paddy	2.18	0.22	1.89
Sukomulyo	Melon	3.86	0.64	3.53
<i>Kraksaan District</i>				
Semampir	Paddy	0.81	0.11	2.72
Sido Pekso	Tobacco	2.47	0.12	3.19
Kregenan	Tobacco	1.92	0.16	3.20
<i>Krejengan District</i>				
Kedung Caluk	Tobacco	2.56	0.16	4.01
Sumberkatimoho	Shallot	3.37	0.16	2.49
Sebaroh	Paddy	1.07	0.35	1.20
<i>Gading District</i>				
Mojolegi	Paddy	1.24	0.20	0.93
Kaliacar	Chili	3.74	0.21	3.89
Wangkal	Corn	0.82	0.03	0.40
<i>Tiris District</i>				
Ranu Agung	Cassava	3.61	0.25	0.65
Andung Sari	Coffee	2.34	0.16	1.31
Tiris	Cassava	2.09	0.16	1.32
<i>Krucil District</i>				
Betek	Cassava	2.05	0.07	0.65
Krucil	Coffee	2.35	0.11	1.57
Bremi	Cassava	2.31	0.30	2.74

4. CONCLUSION

The low nitrogen and C-organic contents and medium-to-high acidity underline the general deficiency of agricultural lands in Probolinggo Regency. As for nutrients, the low contents of nitrogen, phosphor, and calcium and the high content of manganese depict the imbalance in their soil production and plant consumption. This calls for biomass production to fix the land composition as well as meet the plant nutrient requirements through an organic approach. Involving organic fertilizers is therefore recommended, especially sludge and slurry from anaerobic digestion. Further research to map out land deterioration for biomass production is expected to run in other areas so that proper treatments can be performed in damaged soil and preventive steps can be taken to avoid it from happening. This action requires the role of field agricultural advisors to make people aware of sustainable agriculture.

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6. CONFLICT OF INTEREST

The authors declared no conflict of interest

7. REFERENCES

1. E.C. Brevik, and R.W. Arnold. Is the traditional pedologic definition of soil meaningful in the modern context? *Soil Horizons* 56(3): 1–8 (2015). DOI: 10.2136/sh15-01-0002
2. D.H. Goenadi, R.H. Setyobudi, E. Yandri, K. Siregar, A. Winaya, D. Damat, W. Widodo, A. Wahyudi, P.G.

- Adinurani, M. Mel, I. Zekker, M.Z. Mazwan, D.D. Siskawardani, E.D. Purbajanti and I. Ekawati. Land suitability assessment and soil organic carbon stocks as two keys for achieving sustainability of oil palm (*Elaeis guineensis* Jacq.). *Sarhad Journal of Agriculture* 37(Special issue 1): 184–196(2021). DOI: 10.17582/journal.sja/2022.37.s1.184.196
3. C. Dazzi, and G.L. Papa. A new definition of soil to promote soil awareness, sustainability, security and governance. *International Soil and Water Conservation Research* 10(1):99–108(2022). DOI: 10.1016/j.iswcr.2021.07.001
 4. B.W. Murphy. Impact of soil organic matter on soil properties—a review with emphasis on Australian soils. *Soil Research* 53(6):605– 635(2015). DOI:10.1071/SR14246
 5. J. Bouma. Soil security as a roadmap focusing soil contributions on sustainable development agendas. *Soil Security* 100001:1–5(2020). DOI: 10.1016/j.soisec.2020.100001
 6. E.D. Purbajanti, W. Slamet, and E. Fuskhah. Effects of organic and inorganic fertilizers on growth, activity of nitrate reductase and chlorophyll contents of peanuts (*Arachis hypogaea* L.). *IOP Conference Series: Earth and Environmental Science* 250(012048):1–8(2019). DOI: 10.1088/1755-1315/250/1/012048.
 7. R. Lal. Soil health and carbon management. *Food and Energy Security* 5(4):212–222(2016). DOI: 10.1002/fes3.96
 8. M.M. Tahat, K.M. Alananbeh, Y.A. Othman, and D.I. Leskovar. Soil health and sustainable agriculture. *Sustainability (Switzerland)* 12(12):1–26(2020). DOI: 10.3390/SU12124859
 9. M.A. Shahputra, and Z. Zen. Positive and negative impacts of oil palm expansion in Indonesia and the prospect to achieve sustainable palm oil. *IOP Conference Series: Earth and Environmental Science* 122(012008):1–7(2018). DOI: 10.1088/1755-1315/122/1/012008
 10. Z. Vincevica-Gaile, T. Teppand, M. Kriipsalu, K. Krievans, Y. Jani, M. Klavins, R.H. Setyobudi, I. Grinfelde, V. Rudovica, T. Tamm, M. Shanskiy, E. Saaremaa, I. Zekker, and J. Burlakovs. Towards sustainable soil stabilization in peatlands: Secondary raw materials as an alternative. *Sustainability* 13(126726):1–24(2021). DOI: 10.3390/su13126726
 11. R.M. Lehman, V. Acosta-Martinez, J.S. Buyer, C.A. Cambardella, H.P. Collins, T.F. Ducey, J.J. Halvorson, V.L. Jin, J.M.F. Johnson, R.J. Kremer, J.G. Lundgren, D.K. Manter, J.E. Maul, J.L. Smith, and D.E. Stott. Soil biology for resilient, healthy soil. *Journal of Soil and Water Conservation* 70(1):12A-18A(2015). DOI: 10.2489/jswc.70.1.12A
 12. E.A.C. Costantini, and R. Lorenzetti. Soil degradation processes in the Italian agricultural and forest ecosystems. *Italian Journal of Agronomy* 8(4):233–243(2013). DOI: 10.4081/ija.2013.e28
 13. L. Köhl, F. Oehl, and M.G.A. van der Heijden. Agricultural practices indirectly influence plant productivity and ecosystem services through effects on soil biota. *Ecological Applications* 24(7):1842–1853(2014). DOI: 10.1890/13-1821.1
 14. A. Beretta-Blanco, O. Pérez, and L. Carrasco-Letelier. Soil quality decrease over 13 years of agricultural production. *Nutrient Cycling in Agroecosystems* 114:45–55(2019). DOI: 10.1007/s10705-019-09990-3
 15. A. Gholoubi, H. Emami, and A. Alizadeh. Soil quality change 50 years after forestland conversion to tea farming. *Soil Research* 56(5):509–517(2018). DOI: 10.1071/SR18007
 16. K.T. Osman. Soil resources and soil degradation. In: *Soil Degradation, Conservation and Remediation*. Springer, Dordrecht, (2014). p. 1–43. DOI: 10.1007/978-94-007-7590-9_1
 17. J. Schiefer, G.J. Lair, and W.E.H. Blum. Potential and limits of land and soil for sustainable intensification of European agriculture. *Agriculture, Ecosystems and Environment* 230:283–293(2016). DOI: 10.1016/j.agee.2016.06.021
 18. I.O. Ghadeer and S.I. AlKhalil. A study of the environmental impacts of the Gishori industrial complex on plant diversity in Tulkarm, Palestine. *Jordan Journal of Biological Sciences* 12(4):487–494(2019).
 19. R.M.A. Machado, and R.P. Serralheiro. Soil salinity: Effect on vegetable crop growth. Management practices to prevent and mitigate soil salinization. *Horticulturae* 3(2):1–13(2017). DOI: 10.3390/horticulturae3020030
 20. D. Vanham, L. Alfieri, M. Flörke, S. Grimaldi, V. Lorini, A. de Roo and L. Feyen. The number of people exposed to water stress in relation to how much water is reserved for the environment: A global modelling study. *The Lancet Planetary Health* 5(11):e766–e774(2021). DOI: 10.1016/S2542-5196(21)00234-5
 21. G. Imfeld, and S. Vuilleumier. Measuring the effects of pesticides on bacterial communities in soil: A critical review. *European Journal of Soil Biology* 49:22–30(2012). DOI: 10.1016/j.ejsobi.2011.11.010

22. M. Muhammad, U. Isnatin, P. Soni, and P.G. Adinurani. Effectiveness of mycorrhiza, plant growth promoting rhizobacteria and inorganic fertilizer on chlorophyll content in *Glycine max* (L.) cv. Detam-4 Prida. *E3S Web of Conferences* 226(00031):1–5(2021). DOI: 10.1051/e3sconf/202122600031
23. W. Riah, K. Laval, E. Laroche-Ajzenberg, C. Mougín, X. Latour, and I. Trinsoutrot-Gattin. Effects of pesticides on soil enzymes: A review. *Environmental Chemistry Letters* 12(2):257–273(2014). DOI: 10.1007/s10311-014-0458-2
24. E.D. Purbajanti, F. Kusmiyati, W. Slamet, P.G. Adinurani. Chlorophyll, crop growth rate and forage yield of *Brachiaria* (*Brachiaria brizantha* Stapf.) as the result of goat manure in various nitrogen dosage. *AIP Conference Proceedings* 1755(130013):1–5(2016). DOI: 10.1063/1.4958557
25. S. Thiele-Bruhn, J. Bloem, F.T. de Vries, K. Kalbitz, and C. Wagg. Linking soil biodiversity and agricultural soil management. *Current Opinion in Environmental Sustainability* 4(5):523–528(2012). DOI: 10.1016/j.cosust.2012.06.004
26. R. Budiono, P.G. Adinurani and P. Soni. Effect of new NPK fertilizer on lowland rice (*Oryza sativa* L.) growth. *IOP Conference Series: Earth and Environmental Science* 293(012034):1–10(2019). DOI: 10.1088/1755-1315/293/1/012034
27. C. Chandini, R. Kumar, R. Kumar, and O. Prakash. The impact of chemical fertilizers on our environment and ecosystem. In: *Research Trends in Environmental Science*. Edition: 2nd. Chapter: 5(2019). pp. 69–86). https://www.researchgate.net/publication/331132826_The_Impact_of_Chemical_Fertilizers_on_our_Environment_and_Ecosystem
28. Z. Vincevica-Gaile, K. Stankevica, M. Klavins, R.H. Setyobudi, D. Damat, P.G. Adinurani, L. Zalizar, M.Z. Mazwan, J. Burlakovs, D.H. Goenadi, R. Anggriani, and A. Sohail. On the way to sustainable peat-free soil amendments. *Sarhad Journal of Agriculture* 37(Special issue 1):122–135(2021). DOI: 10.17582/journal.sja/2021.37.s1.122.135
29. A.K. Nafiu, M.O. Abiodun, I.M. Okpara, and V.O. Chude. Soil fertility evaluation: A potential tool for predicting fertilizer requirement for crops in Nigeria. *African Journal of Agricultural Research* 7(47):6204–6214(2012). DOI: 10.5897/ajar12.210
30. L.A. Ilagan, R.P. Tablizo, R.B.B. Jr, and N.A. Marquez. Soil fertility evaluation for rice production in Catanduanes province, Philippines. *International Journal of Scientific and Technology Research* 3(12):81–87(2014). <https://www.ijstr.org/final-print/dec2014/Soil-Fertility-Evaluation-For-Rice-Production-In-Catanduanes-Province-Philippines.pdf>
31. M. Damayani, G. Herdiansyah, D.S. Saribun, and R. Hudaya. Evaluation and mapping of soil fertilizer status in the international geopark Ciletuh area, Sukabumi Regency. *IOP Conference Series: Earth and Environmental Science* 393(012050):1–10(2019). DOI: 10.1088/1755-1315/393/1/012050
32. P.G. Adinurani, S. Rahayu, E.D. Purbajanti, D.D. Siskawardani, K. Stankevica and R.H. Setyobudi. Enhanced of root nodules, uptake NPK, and yield of peanut plant (*Arachis hypogaea* L.) using rhizobium and mycorrhizae applications. *Sarhad Journal of Agriculture* 37(Special issue 1): 6–24(2021). DOI: 10.17582/journal.sja/2021/37.s1.16.24
33. A. Prihandiani, D.R. Bella, N.R. Chairani, Y. Winarto, and J. Fox. The tsunami of pesticide use for rice production on Java and Its consequences. *Asia Pacific Journal of Anthropology* 22(4):276–297(2021). DOI: 10.1080/14442213.2021.1942970
34. R.R. Utami, G.W. Geerling, I.R.S. Salami, S. Notodarmojo, and A.M.J. Ragas. Agricultural pesticide use in the upper Citarum river basin: Basic data for model-based risk management. *Journal of Environmental Science and Sustainable Development* 3(2):235–260(2020). DOI: 10.7454/jessd.v3i2.1076
35. H. Batoa, M.A. Limi, A. Hamzah, E.D. Cahyono, P. Arimbawa, W.O. Yusria, and A. Gafaruddin. External factors affecting lowland rice farmers' use of chemical pesticides in Welala Village, Kolaka Timur Regency, Indonesia. *Journal of Agricultural Extension* 23(2):80–89(2019). DOI: 10.4314/jae.v23i2.9
36. K.S. Wicaksono, S. Suratman, R. Suharyadi, and S.H. Murti. Determination of the spatial variability of soil nitrogen content based on reliefs in an apple orchard, Batu, Indonesia. *Journal of Degraded and Mining Lands Management* 6(3):1713–1718(2019). DOI: 10.15243/jdmlm.2019.063.1713
37. I.S. Banuwa, K.F. Hidayat, I. Zulkarnain, P. Sanjaya, A. Afandi, and A. Rahmat. Soil loss and cassava yield under ridge tillage in humid tropical climate of Sumatera, Indonesia. *International Journal of Geomate* 18(67):1–7(2020). DOI: 10.21660/2020.67.78211
38. K. Noerwijati, S. Sholihin, T.S. Wahyuni, R. Budiono, N.V. Minh, R.H. Setyobudi, Z. Vincēviča-Gaile, and L. Husna. Cluster analysis based

- selection in seedling population of cassava clones. *Sarhad Journal of Agriculture* 37(2):398-405(2021). DOI: 10.17582/journal.sja/2021/37.2.398.405
39. S. Supriyadi, M.M.A.R. Rosariastuti, A.P. Kusumawardani, M. Maris, and R.D.A. Putri. Soil quality index relationship with rice production in Sukoharjo Regency, Central Java, Indonesia. *International Conference on Climate Change* 222–234(2016). DOI: 10.15608/iccc.y2016.567
 40. E.D. Purbajanti, F. Kusmiyati, E. Fuskhah, R. Rosyida, P.G. Adinurani, and Z. Vincēviča-Gaile. Selection for drought-resistant rice (*Oryza sativa* L.) using polyethylene glycol. *IOP Conference Series: Earth and Environmental Science* 293(012014):1–8(2019). DOI: 10.1088/1755-1315/293/1/012014
 41. E. Pratama, Y. Sepriani, B.A. Dalimunthe, and K.D. Sitanggang. Analysis of post-paddy fields into red chili cultivation land in Aek Paing Village Rantau Utara District Labuhanbatu. *International Journal of Science and Environment* 2(2):58–62(2022).
 42. I. Comte, F. Colin, O. Grünberger, S. Follain, J.K. Whalen, and J.P. Caliman. Landscape-scale assessment of soil response to long-term organic and mineral fertilizer application in an industrial oil palm plantation, Indonesia. *Agriculture, Ecosystems and Environment* 169:58–68(2013). DOI: 10.1016/j.agee.2013.02.010
 43. S. Kurniawan, M.D. Corre, S.R. Utami, and E. Veldkamp. Soil biochemical properties and nutrient leaching from smallholder oil palm plantations, Sumatra-Indonesia. *Agrivita* 40(2):257–266(2018). DOI: 10.17503/agrivita.v40i2.1723
 44. P.B.O. Herry, A. Dermawan, Q.P. Ilham, P. Pacheco, F. Nurfatriani, and E. Suhendang. Reconciling oil palm economic development and environmental conservation in Indonesia: A value chain dynamic approach. *Forest Policy and Economics* 111(102089):1–12(2020). DOI: 10.1016/j.forpol.2020.102089
 45. T. Hsiao-Hang, E.M. Slade, K.J. Willis, J.P. Caliman, and J.L. Snaddon. Effects of soil management practices on soil fauna feeding activity in an Indonesian oil palm plantation. *Agriculture, Ecosystems and Environment* 218:133–140(2016). DOI: 10.1016/j.agee.2015.11.012
 46. C. Petrenko, J. Paltseva and S. Searle. *Ecological impacts of palm oil expansion in Indonesia*. International Council on Clean Transportation, Washington, USA (2016). p.28. https://theicct.org/sites/default/files/publications/Indonesia-palm-oil-expansion_ICCT_july2016.pdf
 47. T. Guillaume, A.M. Holtkamp, M. Damris, B. Brümmer, and Y. Kuzyakov. Soil degradation in oil palm and rubber plantations under land resource scarcity. *Agriculture, Ecosystems and Environment* 232:110–118(2016). DOI: 10.1016/j.agee.2016.07.002
 48. A.E. Hartemink. *Soil Fertility Decline in the Tropics: With Case Studies on Plantations*. Cabi Publishing, London, UK (1964).
 49. R.P. Putra, M.R.R. Ranomahera, M.S. Rizaludin, R. Supriyanto, and V.A.K. Dewi. Investigating environmental impacts of longterm monoculture of sugarcane farming in Indonesia through DPSIR framework. *Biodiversitas* 21(10):4945–4958(2020). DOI: 10.13057/biodiv/d211061
 50. A. Kusumawati, E. Hanudin, B.H. Purwanto, and M. Nurudin. Composition of organic C fractions in soils of different texture affected by sugarcane monoculture. *Soil Science and Plant Nutrition* 66(1):206–213(2020). DOI: 10.1080/00380768.2019.1705740
 51. R.L. Lubis, J. Juniarti, S.L. Rajmi, A.N. Armer, F.R. Hidayat, H. Zuhakim, N. Yulanda, I.F. Syukri, and D. Fiantis. Chemical properties of volcanic soil after 10 years of the eruption of Mt. Sinabung (North Sumatera, Indonesia). *IOP Conference Series: Earth and Environmental Science* 757(012043):1–11(2021). DOI: 10.1088/1755-1315/757/1/012043
 52. W. Astiko, N.M.L. Ernawati, and I.P. Silawibawa. Nutrient concentration of nitrogen and phosphorus on intercropping of several varieties maize and soybean in dryland North Lombok, Indonesia. *IOP Conference Series: Earth and Environmental Science* 824(1):1–11(2021). DOI: 10.1088/1755-1315/824/1/012001
 53. S. Sufardi, D. Darusman, Z. Zaitun, S. Zakaria, and T.F. Karmil. Chemical characteristics and status of soil fertility on some dryland areas of Aceh Besar District (Indonesia). *International Conference on Sustainable Agriculture (ICOSA2017)* 1–7(2017). <https://www.researchgate.net/publication/326200445>
 54. P.G. Adinurani, S. Rahayu, L.S. Budi, A. Nindita, P. Soni, and M. Mel. Biomass and sugar content of some varieties of sorghum (*Sorghum bicolor* L. Moench) on dry land forest as feedstock bioethanol. *MATEC Web of Conferences* 164(01035):1–5(2018). DOI: 10.1051/mateconf/201816401035
 55. W. Wiryono, Z. Mukhtar, D. Deselina, S. Nurliana, H. Aningtias, and P.M. Anugrah. Soil organic carbon in forest and other land use types at Bengkulu

- City, Indonesia. *Jurnal Manajemen Hutan Tropika* 27(3):184–192(2021). DOI: 10.7226/jtfm.27.3.184
56. L.M. Rachman. Using soil quality index plus to assess soil conditions and limiting factors for dryland farming. *Sains Tanah* 17(2):100–107(2021). DOI: 10.20961/STJSSA.V17I2.46889
 57. RI. Peraturan Pemerintah (PP) No. 150 Tahun 2000 tentang Pengendalian Kerusakan Tanah Untuk Produksi Biomassa [Government Regulations No 150/2000 on Land Deterioration Management for Biomass Production]. Sekretaris Negara Republik Indonesia (2000)
 58. Subardja, D., S. Ritung, M. Anda, Sukarman, E. Suryani, dan R.E. Subandiono. Petunjuk Teknis Klasifikasi Tanah Nasional [National Soil Classification Technical Guidelines] Balai Besar Penelitian dan Pengembangan Sumberdaya Lahan Pertanian, Badan Penelitian dan Pengembangan Pertanian, Bogor. pp 22 (2014).
 59. D. Qi, X. Wieneke, J. Tao, X. Zhou, and U. Desilva. Soil pH is the primary factor correlating with soil microbiome in Karst Rocky desertification regions in the Wushan County, Chongqing, China. *Frontiers in Microbiology* 9(1027):1–12(2018). DOI: 10.3389/fmicb.2018.01027
 60. K. Zhalnina, R. Dias, P.D. de Quadros, A. Davis-Richardson, F.A.O. Camargo, I.M. Clark, S.P. McGrath, P.R. Hirsch, and E.W. Triplett. Soil pH determines microbial diversity and composition in the park grass experiment. *Microbial Ecology* 69(2):395–406(2015). DOI: 10.1007/s00248-014-0530-2
 61. S.O. Oshunsanya. Introductory chapter: Relevance of soil pH to agriculture. In: *Soil pH for Nutrient Availability and Crop Performance*. IntechOpen (2019). DOI: 10.5772/intechopen.82551
 62. D. Tian and S. Niu. A global analysis of soil acidification caused by nitrogen addition. *Environmental Research Letters* 10(24019):1–10(2015). DOI: 10.1088/1748-9326/10/2/024019
 63. H. Wang, J. Xu, X. Liu, D. Zhang, L. Li, W. Li, and L. Sheng. Effects of long-term application of organic fertilizer on improving organic matter content and retarding acidity in red soil from China. *Soil and Tillage Research* 195(104382)(2019). DOI:10.1016/j.still.2019.104382
 64. K.S. Karthika, I. Rashmi, R. Srinivasan, S. Neenu, and P.S. Philip. Acidification of soils and amelioration. *Soil Health/Fertility Management* 1(1): 22–23(2018). <https://iiss.icar.gov.in/eMagazine/v1i1/13.pdf>
 65. D. Neina. The role of soil pH in plant nutrition and soil remediation. *Applied and Environmental Soil Science* 2019:1–9(2019). DOI: 10.1155/2019/5794869
 66. S. Sukmawati, A. Adnyana, D.N. Suprpta, M. Proborini, P. Soni, and P.G. Adinurani. Multiplication arbuscular mycorrhizal fungi in corn (*Zea mays* L.) with pots culture at greenhouse. *E3S Web of Conferences* 226(00044):1–10(2021). DOI: 10.1051/e3sconf/202122600044
 67. H. Wibowo, and A. Kasno. Soil organic carbon and total nitrogen dynamics in paddy soils on the Java Island, Indonesia. *IOP Conference Series: Earth and Environmental Science* 648(012192):1–10(2021). DOI: 10.1088/1755-1315/648/1/012192
 68. S. Kumar, R.S. Meena, S. Sheoran, C.K. Jangir, M.K. Jhariya, A. Banerjee, and A. Raj. Remote sensing for agriculture and resource management. *Natural Resources Conservation and Advances for Sustainability* 91–135(2022). DOI: 10.1016/B978-0-12-822976-7.00012-0
 69. W.R. Horwath, and Y. Kuzyakov. The potential for soils to mitigate climate change through carbon sequestration. In: *Developments in Soil Science* 35:61–92(2018). DOI: 10.1016/B978-0-444-63865-6.00003-X
 70. H. Sukorini, E.R.T. Putri, E. Ishartati, S. Sufianto, R.H. Setyobudi, and S. Suwannarat. Drought stress, salinity, and potential test of IAA (Indole Acetic Acid) of dryland bacteria isolates. *Jordan Journal of Biological Sciences* (2023). INPRESS.
 71. X. Liu, S.J. Herbert, A.M. Hashemi, X. Zhang, and G. Ding. Effects of agricultural management on soil organic matter and carbon transformation - A review. *Plant, Soil and Environment* 52(12): 531–543(2006). DOI: 10.17221/3544-pse
 72. S. Thiele-Bruhn, J. Bloem, F.T. de Vries, K. Kalbitz, and C. Wagg. Linking soil biodiversity and agricultural soil management. In: *Current Opinion in Environmental Sustainability*. 4(5):523–528(2012). DOI: 10.1016/j.cosust.2012.06.004
 73. C.S. Akrotas, A.G. Tekerlekopoulou, I.A. Vasiliadou, and D.V. Vayenas. Composting of olive mill waste for the production of soil amendments. In: *Olive Mill Waste*. C.M. Galanakis (Ed.), Elsevier, p. 161–182(2017). DOI: 10.1016/B978-0-12-805314-0.00008-X
 74. V. Jílková, P. Straková, and J. Frouz. Foliage C:N ratio, stage of organic matter decomposition and interaction with soil affect microbial respiration and its response to C and N addition more than C:N changes during decomposition. *Applied Soil*

- Ecology* 152(103568):(2020). DOI: 10.1016/j.apsoil.2020.103568
75. J. Yanai, T. Omoto, A. Nakao, K. Koyama, A. Hartono, and S. Anwar. Evaluation of nitrogen status of agricultural soils in Java, Indonesia. In: *Soil Science and Plant Nutrition* 60(2):188–195(2014). DOI: 10.1080/00380768.2014.891925
 76. D. Sinha, and P.K. Tandon. An overview of nitrogen, phosphorus and potassium: Key players of nutrition process in plants. In: *Sustainable Solutions for Elemental Deficiency and Excess in Crop Plants*. K. Mishra, P. K. Tandon, and S. Srivastava (Eds.), Springer Singapore, p. 85–117(2020). DOI: 10.1007/978-981-15-8636-1_5
 77. L. Luo, Y. Zhang, and G. Xu. How does nitrogen shape plant architecture?. *Journal of Experimental Botany* 71(15):4415–4427(2020). DOI: 10.1093/jxb/eraa187
 78. A. Krapp. Plant nitrogen assimilation and its regulation: A complex puzzle with missing pieces. *Current Opinion in Plant Biology* 25:115–122(2015).
 79. D.P. Snustad, and M.J. Simmons. *Principles of genetics*. Wiley(2012).
 80. R. Kathpalia, and S.C. Bhatla. Plant mineral nutrition. In: *Plant Physiology, Development and Metabolism*. Springer Singapore p. 37–81(2018). DOI: 10.1007/978-981-13-2023-1_2
 81. V.O. Sadras, and R.A. Richards. Improvement of crop yield in dry environments: benchmarks, levels of organisation and the role of nitrogen. *Journal of Experimental Botany* 65(8):1981–1995(2014). DOI: 10.1093/jxb/eru061
 82. B. Onwuka. Effects of soil temperature on some soil properties and plant growth. *Advances in Plants and Agriculture Research* 6(3):89–93(2018). DOI: 10.15406/apar.2018.08.00288
 83. I. Ekawati, I. Isdiantoni, and Z. Purwanto. Application of immature rice straw compost, azolla, and urea for increasing rice fields production based on local wisdom. *Journal of Basic and Applied Scientific Research* 4(12):130-134(2014).
 84. N.A. Pulungan, S.N.H. Utami, B.H. Purwanto, and J. Sartohadi. Analysis of SOM and soil nutrients for sustainable agriculture in hilly areas: Central Part of Bogowonto Catchment, Java, Indonesia. In: *Proceeding of the 1st International Conference on Tropical Agriculture*. Springer International Publishing, p. 197–208(2017). DOI: 10.1007/978-3-319-60363-6_19
 85. M. Butusov, and A. Jernelöv. Phosphorus in the organic life: Cells, tissues, organisms. In: *Phosphorus. Springer Briefs in Environmental Science*. Springer, p. 13–17(2013). DOI: 10.1007/978-1-4614-6803-5_2
 86. H. Malhotra, V. Vandana, S. Sharma, and R. Pandey. Phosphorus nutrition: Plant growth in response to deficiency and excess. In: *Plant Nutrients and Abiotic Stress Tolerance*. M. Hasanuzzaman, M. Fujita, K. Oku, H., Nahar, and B. Hawrylak-Nowak (Eds.), Springer Singapore, p. 171–190(2018). DOI: 10.1007/978-981-10-9044-8_7
 87. P.J. White, and E.J. Veneklaas. Nature and nurture: The importance of seed phosphorus content. *Plant and Soil* 357(1–2):1–8(2012). DOI: 10.1007/s11104-012-1128-4
 88. K. Goulding, T.S. Murrell, R.L. Mikkelsen, C. Rosolem, J. Johnston, H. Wang and M.A. Alfaro. Outputs: Potassium losses from agricultural systems. In: *Improving Potassium Recommendations for Agricultural Crops*. Murrell, T.S. et al. (Eds.) Cham: Springer International Publishing, pp. 75–97(2021). DOI: 10.1007/978-3-030-59197-7_3
 89. G.P. Acharya, M.A. McDonald, B.P. Tripathi, R.M. Gardner, and K.J. Mawdesley. Nutrient losses from rain- fed bench terraced cultivation systems in high rainfall areas of the mid-hills of Nepal. *Land and Degradation Development* 18(5):486–499(2007). DOI: 10.1002/ldr.792
 90. X. Wang, L. Lang, T. Hua, H. Li, C. Zhang and W. Ma. Effects of aeolian processes on soil nutrient loss in the Gonghe Basin, Qinghai–Tibet Plateau: An experimental study. *Journal of Soils and Sediments* 18(1):229–238(2018). DOI: 10.1007/s11368-017-1734-0
 91. W.D.C. Mendes, J.A. Júnior, P.C.R. da Cunha, A.R. da Silva, A.W.P. Evangelista, D. Casaroli. Potassium leaching in different soils as a function of irrigation depths. *Revista Brasileira de Engenharia Agrícola e Ambiental* 20(11):972–977(2016). DOI: 10.1590/1807-1929/agriambi.v20n11p972-977
 92. J. Divya, and S. Belagali. Effect of chemical fertilizers on physico-chemical characteristics of agricultural soil samples of Nanjangud taluk, Mysore District, Karnataka, India. *Journal of Environmental Sciences* 6:181–187(2012). <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.1088.8610&rep=rep1&type=pdf>
 93. K. Prajapati, and H.A. Modi. The importance of potassium in plant growth – A review. *Indian Journal of Plant Sciences* 1:177–186(2012).
 94. D.M. Oosterhuis, D.A. Loka, E.M. Kawakami, and

- W.T. Pettigrew. The physiology of potassium in crop production. In: *Advances in Agronomy*. D.L. Sparks (Ed.). Academic Press, p. 203–233(2014). DOI: 10.1016/B978-0-12-800132-5.00003-1
95. M. Wang, Q. Zheng, Q. Shen, and S. Guo. The critical role of potassium in plant stress response. *International Journal of Molecular Sciences* 14(4):7370–7390(2013). DOI: 10.3390/ijms14047370
 96. O. Husson. Redox potential (Eh) and pH as drivers of oil/plant/microorganism systems: A transdisciplinary overview pointing to integrative opportunities for agronomy. *Plant and Soil* 362(1–2):389–417(2013). DOI: 10.1007/s11104-012-1429-7
 97. S.K. Das. Role of micronutrient in rice cultivation and management strategy in organic agriculture—A reappraisal. *Agricultural Sciences* 05(09):765–769(2014). DOI: 10.4236/as.2014.59080
 98. K.K. Mathan, and A. Amberger. Influence of iron on the uptake of phosphorus by maize. *Plant and Soil* 46(2):413–422(1977). DOI: 10.1007/BF00010097
 99. M. Broadley, P. Brown, I. Cakmak, Z. Rengel, and F. Zhao. Function of nutrients. In: *Marschner's Mineral Nutrition of Higher Plants*. P. Marschner (Ed.), Elsevier, p. 191–248(2012). DOI: 10.1016/B978-0-12-384905-2.00007-8
 100. J.M. Connorton, J. Balk, and J. Rodríguez-Celma. Iron homeostasis in plants – A brief overview. *Metallomics* 9(7):813–823(2017). DOI: 10.1039/C7MT00136C
 101. Y. Yao, G. Xu, D. Mou, J. Wang, and J. Ma. Subcellular Mn compartation, anatomic and biochemical changes of two grape varieties in response to excess manganese. *Chemosphere* 89(2):150–157(2012). DOI: 10.1016/j.chemosphere.2012.05.030
 102. E. Gimeno-García, V. Andreu, and R. Boludab. Heavy metals incidence in the application of inorganic fertilizers and pesticides to rice farming soils. *Environmental Pollution* 92(1):19–25(1996). DOI: 10.1016/0269-7491(95)00090-9
 103. M.M. Onakpa, A.A. Njan, and O.C. Kalu. A review of heavy metal contamination of food crops in Nigeria. *Annals of Global Health* 84(3):488–494(2018). DOI: 10.29024/aogh.2314
 104. I. Ekawati and Z. Purwanto. Potensi abu limbah pertanian sebagai sumber alternatif unsur hara kalium, kalsium, dan magnesium untuk menunjang kelestarian produksi tanaman. [Potential of agricultural waste ash as an alternative source of potassium, calcium and magnesium nutrients to support sustainable crop production]. *Prosiding Seminar Nasional Kedaulatan Pangan dan Energi Universitas Trunojoyo* 27:135-139(2012).
 105. Y. Ngongo, T. Basuki, B. de Rosari, E.Y. Hosang, J. Nulik, H. daSilva, D.K. Hau, A. Sitorus, N.R.E. Kotta, G.N. Njurumana, E. Pujiono, L. Ishaq, A.V. Simamora, and Y.S. Mau, Local wisdom of West Timorese farmers in land management. *Sustainability* 14(10-6023): 1–21(2022). DOI: 10.3390/su14106023
 106. H. Riastyadiningrum and I. Ekawati. Manajemen tanaman sehat budidaya padi untuk meningkatkan produksi dan pendapatan usahatani padi. [Management of healthy rice cultivation plants to increase production and income of rice farming]. *Cemara* 17(2): 25–34(2020).
 107. I. Ekawati and Z. Purwanto. Alih teknologi pestisida nabati berbasis sumberdaya lokal pada petani padi. [Transfer of local resource-based vegetable pesticide technology to rice farmers]. *Cemara* 10(1):36–40(2013).
 108. R.H. Setyobudi, E. Yandri, M.F.M. Atoum, S.M. Nur, I. Zekker, R. Idroes, T.E. Talle, P.G. Adinurani, Z. Vincēviča-Gaile, W. Widodo, L. Zalizar, N. Van-Minh, H. Susanto, R.K. Mahaswa, Y.A. Nugroho, S.K. Wahono, and Z. Zahriah. Healthy-smart concept as standard design of kitchen waste biogas digester for urban households. *Jordan Journal of Biological Sciences* 14(3):613–620(2021). DOI: 10.54319/jjbs/140331
 109. H. Susanto, E. Yandri, R.H. Setyobudi, D. Sugiyanto, S.M. Nur, P.G. Adinurani, H. Herianto, Y. Jani, S.K. Wahono, Y. Nurdiansyah and A. Yaro. Development of the biogas-energized livestock feed making machine for breeders. *E3S Web of Conferences* 188(00010):1–13(2020). DOI: 10.1051/e3sconf/202018800010
 110. H. Susanto, A.S. Uyun, R.H. Setyobudi, S.M. Nur, E. Yandri, J. Burlakovs, A. Yaro, K. Abdullah, S.K. Wahono, and Y.A. Nugroho. Development of moving equipment for fishermen's catches using the portable conveyor system. *E3S Web of Conferences* 190(00014):1-10(2020) DOI: 10.1051/e3sconf/202019000014
 111. J. Burlakovs, Z. Vincevica-Gaile, V. Bisters, W. Hogland, M. Kriipsalu, I. Zekker, R.H. Setyobudi, Y. Jani, and O. Anne. Application of anaerobic digestion for biogas and methane production from fresh beach-cast biomass. *Proceedings EAGE GET 2022 – 3rd Eage Global Energy Transition, The Hague, Netherlands 2022:1–5(2022)*. DOI:

- 10.3997/2214-4609.202221028
112. K. Abdullah, A.S. Uyun, R. Soegeng, E. Suherman, H. Susanto, R.H. Setyobudi, J. Burlakovs, and Z. Vincēviča-Gaile. Renewable energy technologies for economic development. *E3S Web of Conference* 188(00016):1–8 (2020). DOI: 10.1051/e3sconf/202018800016
113. P.G. Adinurani, R.H. Setyobudi, S.K. Wahono, M. Mel, A. Nindita, E. Purbajanti, S.S. Harsono, A.R. Malala, L.O. Nelwan, and A. Sasmito. Ballast weight review of capsule husk *Jatropha curcas* Linn. on acid fermentation first stage in two-phase anaerobic digestion. *Proceedings of the Pakistan Academy of Sciences: Part B* 54(1):47–57(2017).
114. R. Hendroko, T. Liwang, P.G. Adinurani, L.O. Nelwan, Y. Sakri, and S.K. Wahono. 2013. The modification for increasing productivity at hydrolysis reactor with *Jatropha curcas* Linn. capsule husk as bio-methane feedstocks at two-stage digestion. *Energy Procedia* 32:47–54. DOI: 10.1016/j.egypro.2013.05.007
115. P.G. Adinurani, R. Hendroko, S.K. Wahono, A. Sasmito, L.O. Nelwan, A. Nindita and T. Liwang. Optimization of concentration and EM4 augmentation for improving bio-gas productivity from *Jatropha curcas* Linn. capsule husk. *International Journal of Renewable Energy Development* 3(1):73–78(2014). DOI: 10.14710/ijred.3.1.73-78
116. F.J. Arriaga, J. Guzman, and B. Lowery. 2017. Conventional agricultural production systems and soil functions. In: *Soil Health and Intensification of Agroecosystems*. pp. 109–125(2017). Academic Press. DOI: 10.1016/B978-0-12-805317-1.00005-1
117. N. Miles, J.H. Meyer, and R. van Antwerpen. Soil organic matter data: What do they mean. *Proceedings South African Sugar Technologists Association* 81:324–332(2008).
118. M. Krauss, M. Wiesmeier, A. Don, F. Cuperus, A. Gattinger, S. Gruber, and M. Steffens. Reduced tillage in organic farming affects soil organic carbon stocks in temperate Europe. *Soil and Tillage Research* 216(105262)(2022). DOI: 10.1016/j.still.2021.105262