Proceedings of the Pakistan Academy of Sciences: B Life and Environmental Sciences 60(3): 367-376 (2023) Copyright © Pakistan Academy of Sciences ISSN (Print): 2518-4261; ISSN (Online): 2518-427X http://doi.org/10.53560/PPASB(60-3)905



Review Article

# Role of *Dof* Transcription Factors under Abiotic Stresses

Amjid Khan<sup>1,2</sup>\*, Fouzia Bibi<sup>1</sup>, Sohail Ahmad Jan<sup>3</sup>, and Zabta Khan Shinwari<sup>1</sup>

<sup>1</sup>Department of Plant Sciences, Quaid-i-Azam University, Islamabad, Pakistan <sup>2</sup>Department of Botany, University of Mianwali, Mianwali, Pakistan <sup>3</sup>Department of Bioinformatics and Biosciences, Capital University of Science and Technology, Islamabad, Pakistan

Abstract: For the survival of the rapidly growing global population, plant species must exhibit tolerance towards climate change. Plants possess mechanisms to respond to stress by changing their biological processes and stimulating stress-responsive genes. The Dof (TFs) family, which binds to DNA with a single finger, reflects a plant-specific group of TFs that play an important part in regulating plants that are facing different types of abiotic stresses which may influence their growth and development. Discovery of this family has made a significant impact on the field of plant sciences. However, the characterization of *Dof* transcription factors in crop plants is currently limited reported. Several Dof transcription factors (TFs) of plants have been shown in nature. The transcription factors TaDofs, StDof, MnDofs, JrDof3TF, Va/VvDofs, GhDof1, OsDof1, SmeDof, ZmDof, CsDof, DcDof, CaDofs, ThDof, BraDof, and AcDof are important for abiotic stressors such heat, cold, salt, drought, and heavy metals. In addition, Dof transcription factors play a role in the regulation of factors related to yield and quality. Nevertheless, some outstanding issues remain. The review article provides a summary of the role of various stress-responsive Dof transcription factors in response to abiotic stresses. Additionally, this study investigates the limitations and possible opportunities associated with Dof transcription factors in the development of crops that are capable of withstanding climate change. Therefore, it is recommended to conduct comprehensive research on Dof transcription factors (TFs) across many different transcription factors fields to find their potential novel functionality, which will be beneficial to our retention of the delicate biological processes in plants.

Keywords: Abiotic stress, Dof, Transcription factor, Salinity stress, Crop improvement

## **1. INTRODUCTION**

Transcription factors (TFs) are essential to plant regulation and signaling networks [1, 2]. TFs control gene expression by binding to promoter DNA sequences [3-5]. In adaptation to abiotic stress, such as osmotic stress, cold, heat, and drought [1, 6-10] plants need more TFs than animals [5]. Drought, salinity, cold, etc. significantly reduce plant productivity. Stresses can reduce major crop yields by 50 %. Genes or other transcription factors (TFs) linked to abiotic stress increase proline content, close stomata to decrease transpiration, enhance stress-protective enzyme synthesis, and enhance abiotic stress tolerance [11-13]. The genetic factors TATA, CAAT Box, ARR1, GATA, AGAAA, CAAT, and DNA-binding are major contributors to genetics. A Zinc Finger (*Dof*) factor that's found in the promoter region of *OsRGLP2* is responsible for regulating several plant processes such as defense, light responses, development, and growth. [14]. Transcription factors (TFs) known as DNA-binding with one finger (*DoF*) are linked to the processes of plant growth and development. The *Dof* transcription factors exhibit a DNA-binding domain (C2/C2) that consists of 52 amino acids and a zinc finger that's capable of binding to the 5-(T/A) AAAG-3 target DNA sequence along with a resilient target DNA sequence [15]. The first Dof gene in maize was successfully identified [16]

Received: May 2023; Revised: July 2023; Accepted: August 2023

<sup>\*</sup>Corresponding Author: Amjid Khan <amjidkhan@bs.qau.edu.pk>

and the interaction of multiple regulatory proteins responsible for the regulation of *Dof* transcription factors via their C-terminal region [17, 18]. Low/ high temperatures, high salt treatment, and drought significantly influence plant development and productivity. Abiotic stresses reduce vegetable quality and production [19]. Pepper showed the expression of CaDofs as its response to abiotic stress [20], and researchers in the field of tea plant cultivation have carried out investigations into many different transcription factors (TFs). However, the regulatory mechanism of *Dof* transcription factors in tea plants remains difficult to identify [17, 21]. The Dof transcription factors (TFs) have significance in various biological processes throughout every stage of the life cycle of plants, and have importance for growth, seed storage protein synthesis, germination of seeds and development, regulatory metabolism, the photosynthesis process, flowering, and responses to stress [22-24]. Dof transcription factors regulate secondary metabolic processes like glucosinolate biosynthesis, cell cycle regulation, and flavonoids [25, 26]. They also regulate the cell cycle, phytochrome and cryptochrome signaling, plant hormonal signaling, abaxial-abaxial polarity, nitrogen use, and abiotic and biotic stress tolerance [27, 28]. The DcDof transcription factor, as compared to other plant Dof factors, has been given little focus in academic research. Furthermore, certain Dof genes can regulate both plant growth and stress responses. The enhanced expression of Arabidopsis CDF3 has been observed to enhance the resistance to drought, cold, and osmotic stress, while also slowing down the start of flowering. This suggests that the gene in concern exerts a regulatory effect over both flowering time and tolerance to abiotic stress [29]. Overexpression of the Dof transcription factor TDDF1 in tomatoes increased flowering-time control gene expression, causing early flowering and increased tolerance to drought, salt, and Phytophthora infestans late blight [30]. Under salt stress, the growth of the primary root in rice is decreased. This can be attributed to the suppression of OsDOF15 expression in the roots, resulting in to decrease in the production of ethylene [31] and studies have shown the importance of *Dof* transcription factors in various biological processes, promoting plant growth and development [32]. Current investigation showed that *Dof* transcription factors play an important role in multiple signaling pathways regulation, in

response to different abiotic stresses, plant growth and development, and other biological processes. The results indicate that *Dof* transcription factors have the potential for regulating both lipid metabolism and stress responses. Nevertheless, the members of the *Dof* TF family have currently to be reported. Current research provides an extensive overview of the resistance to abiotic stress, as well as the evolutionary links and the *Dof* transcription factor family.

# 2. STRUCTURE, FUNCTION, AND MECHANISM OF ACTION OF VARIOUS GROUPS OF *DOF* TRANSCRIPTION FACTORS

Suspendisse Sequence motifs that match DNA binding domains classify TFs. As shown by comparing the principal classes, Dof transcription factors have distinct functions in the hierarchy of response, although many interact as part of their response. Plants use several mechanisms to tolerate ecological problems such as drought, salinity, oxidative stress, cold, heat, and other infections that affect plant growth and development. Due to their sessile nature, environmental challenges can cause suboptimal growth conditions, requiring metabolic pathways to be changeable to allow plants to resist, tolerate, or recover from stress. The complex structure of the abiotic stress response in plants results from the polygenic nature and association between events involving signal transduction and stress response factor synthesis during a time of stress. As plants are sessile (immobile), environmental cues including drought, water logging, salt, mineral toxicity, and temperature change (frost, cold, heat) negatively affect their metabolism, growth, and development [33, 34].

In the same way, the upregulation of *ZmDOF36* resulted in a reduction in the levels of reduced sugars and soluble sugars within the endosperm of maize seeds. On the other hand, an increase in soluble sugar levels was found to promote the synthesis of starch. *ZmDOF36* was found to exert a positive regulatory effect on the expression of various genes associated with starch synthesis, including *ZmAGPS1a*, *ZmAGPL1*, *ZmISA1*, *ZmISA3*, *ZmGBSSI*, and *ZmSSIIa*. This regulation was achieved through the binding of *ZmDOF36* to specific motifs located in the promoters downstream

of these genes [35]. The previous study showed that ThDOF14 enhances the ability of plants to tolerate salt and osmotic stress. This is achieved through an increase in proline levels and an improvement in the plant's ability to scavenge reactive oxygen species (ROS) [36]. Similarly, it has been observed that ThDOF14 exhibits a specific affinity for the DOF motif present in the downstream promoter region of TheIF1A. This interaction suggests that *ThDOF14* may play a role in the plant's response to salt stress and osmotic stress by regulating the expression or engaging in molecular interactions with TheIF1A [37]. The study conducted by Cai et al. (2016) showed that the tomato SlDOF22 gene had an impact on the accumulation of ascorbic acid (AsA) and also enhanced salt tolerance in plants [38]. Figure 1 shows how plants respond to various stresses via activating pathways, gene interaction, and molecular "crosstalk". Plants can defend themselves against different types of stress, which makes possible the identification of the most adaptable and resistant varieties for the benefit of producing a plant with desirable characteristics. Abiotic resistance genes usually stimulate a signal transduction pathway to identify pathogens and acquire resistance. The molecular understanding of

abiotic stress response primarily depends on genetic engineering techniques, such as over-expression or mutation studies, to provide insight into the sensor, signal transduction factors, and antimicrobial factor genes.

# 3. ROLE OF *DOF* TRANSCRIPTION FACTORS UNDER ABIOTIC STRESSES

Abiotic stressors like heat, cold, drought, flooding, heavy metals, and salt are affecting crop life due to global warming. Abiotic factors limit maize, cotton, rice, and wheat yields by 50 % [39]. Table 1 shows how *Dof* TFs/genes/proteins affect plant abiotic stress tolerance.

#### 3.1. Drought Stress

Drought stress, which is caused by excessive groundwater, decreased precipitation, and high temperatures, depicts a primary contributor to agricultural problems, which exert an adverse effect on the worldwide economy and the food security of a significant number of people by limiting crop growth and production [39, 58].

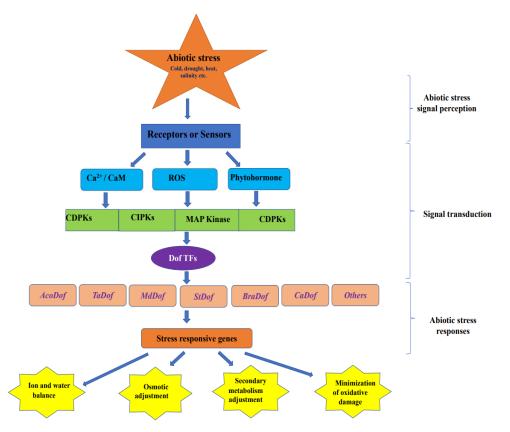


Fig. 1. Model for *Dof* TFs regulation abiotic stress signaling pathways.

| Stress  | Crop                        | Dof TFs / Genes/ Proteins   | References |
|---|-----------------------------|---|------------|
| Salinity, cold, heat, and drought stress                                | Ananas comosus              | AcoDof1, AcoDof12, AcoDof26,<br>AcoDof22, AcoDof9, AcoDof19,<br>AcoDof23, AcoDof11, AcoDof17,<br>AcoDof20, and AcoDof1. | [40]       |
| Heat, drought, and heavy metals stress                                  | Triticum aestivum           | TaDof, TaDof14, TaDof14   | [41, 42]   |
| Salinity, drought, cold stress  | Malus domestica             | MdDof   | [5]        |
| Salinity, osmotic, heat and<br>low temperature, and drought<br>stresses | Solanum lycopersicum        | SICDFs, SICDF1-5  | [27]       |
| Drought, salinity, low<br>temperature, and high<br>temperature          | Glycine max                 | GmDof4.2  |            |
| Cold, heat, salinity, and drought stress                                | Brassica pekinensis Rupr    | BraDof  | [18]       |
| Salt and osmotic stress   | Tamarix hispida             | ThDof, ThelF1A  | [37]       |
| Temperature, salinity, heat, and irritation                             | Daucus carota               | DcDof   | [44]       |
| Heat stress   | Oryza sativa                | OsDOF27   | [45]       |
| Salinity and drought stress   | Camellia sinensis           | CsDof   | [17]       |
| Cold stress   | Brassica napus              | BnCDF1  | [46]       |
| Salt stress   | Nelumbo nucifera            | NnDofs  | [47]       |
| Heat and salinity stress  | Capsicum annuum             | CaDofs  | [20]       |
| Salinity and heat stress  | Chrysanthemum<br>morifolium | CmDOFs  | [48]       |
| Salinity stress   | Camelina sativa             | CsDof   | [49]       |
| Drought Salt and stress   | Tamarix hispida             | WRKY (ThWRKY4)  | [50]       |
| Salinity stress   | Zea mays                    | ZmDof, ZmDof16, ZmDof22,<br>ZmDof36   | [51]       |
| Osmotic, heat, and drought stress                                       | Populus trichocarpa         | PtrDofs   | [52]       |
| Cold stress   | Vitis vinifera grapevine    | VaDof17d  | [8]        |
| Salinity and salt stress  | Gossypium hirsutism         | GhDofl  | [53]       |
| Heat, cold, heat, and salt stress                                       | Saccharum spontaneum        | SsDofs  | [15]       |
| Drought and salinity stress   | Solanum melongena           | SmeDof  | [54]       |
| Salt and drought stress   | Vaccinium corymbosum        | VcDof   | [55]       |
| Drought and salinity stress   | Solanum tuberosum           | StDof   | [24]       |
| Drought and salt stress   | Rosa chinensis              | RchDofs   | [56]       |
| Salt stress   | Medicago truncatula         | MtDof32   | [57]       |

Table 1. Role of different Dof transcriptional factor gene families in abiotic stress tolerance in plants

The transcription factor *VyDOF8*, which comes from *Vitis yeshanensis*, a species of Chinese wild grapevine, exhibited a significant rise in expression levels under conditions of drought, cold, and saltinduced stress. *Dof* transcription factors play an important role in improving the responses of plants to drought stress. The overexpression of *VyDOF8*  in tobacco plants has the potential to enhance their ability to withstand drought conditions. This has led to the reported increase in abscisic acid (ABA) concentration, root growth, proline, and chlorophyll accumulation, stress response gene expression, and antioxidant activity [59]. *SICDF1* and *SICDF3* overexpression in *Arabidopsis* increased drought tolerance [17, 60]. *TaDofs* are nucleusbased transcription factors that affect growth, development, and abiotic responses. Customized annotation revealed that drought stress produced *TaDofs*, which were associated with defense, phytohormone response, growth, development, and metabolism [61]. The increased expression of *Solanum tuberosum Dof* (*StDof*) genes which can improve growth, development, and abiotic stress tolerance, has been found in response to both drought stress and abscisic acid (ABA) treatments. [24].

## 3.2. Salinity Stress

The process of global climate change has resulted in several types of environmental stressors, including soil salinity. This abiotic factor is the second largest cause of global productivity in agriculture loss. The adverse impacts of salinity on the growth and productivity of plants are mediated through various physiological, biochemical, and molecular mechanisms. These responses include ion homeostasis regulation, biosynthesis of phytohormones, and antioxidant defense systems activation. Plants respond to salinity by inducing the expression of stress-related genes, proteins, and metabolites that help to alleviate some of the adverse effects of salinity. Increasing salt tolerance becomes essential for maintaining global agronomic productivity. Transcription factors play an important part in the mechanism that causes plants' salt tolerance [62, 63]. MnDofs improve lotus salt tolerance [47]. The salinity responses of transgenic tobacco plants have been improved through the application of BZIP transcription factors obtained from salt-tolerant lotus root tips [43, 64]. Overexpression of SICDF1 or SICDF3 in Arabidopsis plants improved salt tolerance, stressresponsive gene expression (COR15, RD29A, RD10), flowering time, and specific target genes and metabolites [65]. GmWRKY54 overexpression reduced soybean salinity stress [66]. The overexpression of GhWRKY34 in Arabidopsis plants results in a specific absorption of Na<sup>+</sup> or K<sup>+</sup> ions in both roots and leaves, which helps promote the development of salt tolerance [67].

## 3.3. Heat Stress

Heat stress is a major environmental issue

because of its negative impact on plant growth and development. The negative effects of high temperatures on cellular integrity and viability are well-studied. However, it is significant that plants have developed a mechanism of heat shock response that mitigates the negative impacts of abrupt temperature changes. Thermotolerant plants can endure heat stress, which can alter vegetable crop development, yield, and quality [2, 68-70]. Heat stress affects plant growth, health, and physiological, phenotypic, and genetic expressions [71, 72]. The JrGRAS2 gene was recently identified as a regulator of heat shock protein (HSP) expression, thereby enhancing the capability of plants to tolerate heat stress. Therefore, JrGRAS2 is considered an important gene for plant genetic engineering that focuses on improving heat response. The HsfA1 protein is considered to be an important activator of the key response to heat stress [73-75]. Transcription factors have been found to have an adverse effect on gene expression in response to abiotic stress and the quality of grains. Additionally, the number of Dof family members has been shown to regulate glycogen and starch production in grains in response to high temperatures (HS) [76]. The findings suggest that Arabidopsis plants with an insertional mutant of Mterf18 exhibit greater resistance to heat stress and elevated levels of HSP transcripts compared to their wild-type plants [77, 78]. Previous study shows that shot1 mutants and ATAD3-disrupted plants exhibit mitochondrial absorption and signaling problems, ultimately resulting in enhanced heat tolerance in plants. The moso bamboo that survived heat stress exhibits a stimulation of LTR retrotransposons, especially PHRE1 and PHRE2 [79].

#### 3.4. Heavy Metal Stress

The effect of heavy metal stress (HM) on plants can be seen through different pathways such as growth inhibition, physiological process delays, and decreased crop productivity. These effects occur due to the change in cell membrane integrity, cellular ionic balance, metabolic balance, protein, and enzyme activity [69, 70]. Heavy metals can cause reactive oxygen species (ROS) and affect physiological functions such as photosynthesis, respiration, and vascular and enzymatic activity. Furthermore, a high level of heavy metals can cause difficulties in ion homeostasis [80]. A previous study showed that OsHMP09, OsHMP018, and OsHMP22 exhibited higher expression levels throughout all tissues. Alternatively, AtHMP20, AtHMP23, AtHMP25, AtHMP31, AtHMP35, and AtHMP46 showed higher expression levels in the roots and leaves under various heavy metal stresses [81]. The NtSOD gene family enhances heavy metal toxicity tolerance in Nicotiana tabacum [82]. A previous study showed that JrDof3TF improves the heat stress response of JrGRAS2 and is a heat response candidate gene in plant molecular breeding [75].

#### 3.5. Cold Stress

The distribution, growth, and yield of crops are all significantly affected by cold stress. Sessile plants have many kinds of physiological and biochemical responses to cold stress, resulting in a 40 % decrease in temperate yields for agriculture. Cold stress adaptation is regulated by TFs and proteins [83, 84]. Cold stress causes 51-82 % of global crop output losses [85]. The study found that overexpression of Va/VvDofs resulted in improvements in root development, germination rate, and seed development, which led to improved cold resistance. However, the Dofl7d-Ed mutant exhibited reduced cold tolerance, as evidenced by a decrease in rafnose family oligosaccharides [8] and moreover, in Gossypium hirsutum GhDof1 overexpression can improved cold tolerance [53]. Previous study showed that OsDofl can enhance cold tolerance in rice act as a potential target for rice genetic breeding [86].

## 4. CONCLUSION AND FUTURE PERSPECTIVE

This review shows how *Dof* TFs can improve plant responses to various abiotic stresses. These studies show how DNA-binding with one finger (*Dof*) transcription factors can combat environmental challenges and increase yield and productivity under stress. Climate change threatens growth, development, and crop yield due to abiotic factors such salt, heat, cold, waterlogging, drought, and heavy metals. By focusing on various genes and or their regulators, it is urgently necessary to create crops that are tolerant to abiotic stress. The considerable number of *Dof* functional studies presented in this review highlights the existence of many unanswered questions regarding Dof transcription factors. The functional diversity of Dof transcription factors has garnered significant interest in recent years. However, the investigation into the molecular mechanisms through which these transcription factors regulate specific biological processes, such as plant growth, dormancy, and germination, has been restricted to a few model plants. Therefore, additional research must be conducted to further current knowledge in this area. In the future, it is of great interest to understand how these factors contribute to the stimulation of plant defense mechanisms against environmental stresses, ultimately resulting in an increase in crop yield and some members of *Dof* may become potential factors to have quite wide application prospects for the development of the food processing and biofuel industries. Therefore, it is important to conduct comprehensive research on Dof transcription factors (TFs) across many different fields in order to find their potential novel functionality, which will be beneficial to our retention of the delicate biological processes in plants.

## 5. CONFLICT OF INTEREST

The authors declared no contlict of interest.

#### 6. **REFERENCES**

- S. Lindemose, C. O'Shea, M.K. Jensen and K. Skriver. Structure, function and networks of transcription factors involved in abiotic stress responses. *International journal of molecular sciences* 14(3): 5842-5878 (2013).
- B. Yu, F. Ming, Y. Liang, Y. Wang, Y. Gan, Z. Qiu, S. Yan and B. Cao. Heat Stress Resistance Mechanisms of Two Cucumber Varieties from Different Regions. *International Journal of Molecular Sciences* 23(3): 1817 (2022).
- M. Raices and M.A. D'Angelo. Nuclear pore complexes and regulation of gene expression. *Current opinion in cell biology* 46: 26-32 (2017).
- K. Rybak, P.T. See, H.T. Phan, R.A. Syme, C.S. Moffat, R.P. Oliver and K.C. Tan. A functionally conserved Zn2Cys6 binuclear cluster transcription factor class regulates necrotrophic effector gene expression and host-specific virulence of two major Pleosporales fungal pathogens of wheat. *Molecular plant pathology* 18(3): 420-434 (2017).
- 5. Z. Zhang, L. Yuan, X. Liu, X. Chen and X. Wang. Evolution analysis of Dof transcription factor family and their expression in response to multiple abiotic

stresses in Malus domestica. Gene 639: 137-148 (2018).

- X. Sun, J.T. Matus, D.C.J. Wong, Z. Wang, F. Chai, L. Zhang, T. Fang, L. Zhao, Y. Wang and Y. Han. The GARP/MYB-related grape transcription factor AQUILO improves cold tolerance and promotes the accumulation of raffinose family oligosaccharides. *Journal of experimental botany* 69(7): 1749-1764 (2018).
- M.K. Udvardi, K. Kakar, M. Wandrey, O. Montanari, J. Murray, A. Andriankaja, J.-Y. Zhang, V. Benedito, J.M. Hofer and F. Chueng. Legume transcription factors: global regulators of plant development and response to the environment. *Plant Physiology* 144(2): 538-549 (2007).
- Z. Wang, Y. Wang, Q. Tong, G. Xu, M. Xu, H. Li, P. Fan, S. Li and Z. Liang. Transcriptomic analysis of grapevine Dof transcription factor gene family in response to cold stress and functional analyses of the VaDof17d gene. *Planta* 253(2): 1-14 (2021).
- H. Xin, W. Zhu, L. Wang, Y. Xiang, L. Fang, J. Li, X. Sun, N. Wang, J.P. Londo and S. Li. Genome wide transcriptional profile analysis of Vitis amurensis and Vitis vinifera in response to cold stress. *PloS one* 8(3): e58740 (2013).
- J.-K. Zhu. Abiotic stress signaling and responses in plants. *Cell* 167(2): 313-324 (2016).
- H. Qamar, M. Ilyas, S.A. Jan, H.S.B. Mustafa, A. Arshad, M.S. Yar, M. Ahmed, S. Hussain, R. Wells, and H. Khurshid. Recent trends in molecular breeding and biotechnology for the genetic improvement of Brassica species against drought stress. *Fresenius Environ. Bull* 29(1): 19-25 (2020).
- Z.K. Shinwari, S.A. Jan, K. Nakashima and K. Yamaguchi-Shinozaki. Genetic engineering approaches to understanding drought tolerance in plants. *Plant Biotechnology Reports* 14: 151-162 (2020).
- Z.K. Shinwari, F. Tanveer and I. Iqrar. Role of microbes in plant health, disease management, and abiotic stress management. *Microbiome in Plant Health and Disease: Challenges and Opportunities* 231-250 (2019).
- T. Mahmood, T. Tahir, F. Munir, and Z.K. Shinwari. Characterization of regulatory elements in OsRGLP2 gene promoter from different rice accessions through sequencing and in silico evaluation. *Computational Biology and Chemistry* 73: 206-212 (2018).
- M. Cai, J. Lin, Z. Li, Z. Lin, Y. Ma, Y. Wang and R. Ming. Allele specific expression of Dof genes responding to hormones and abiotic stresses in sugarcane. *PloS one* 15(1): e0227716 (2020).
- A.L. Arce, J.V. Cabello and R.L. Chan. Patents on plant transcription factors. *Recent Patents on Biotechnology* 2(3): 209-217 (2008).
- 17. H. Li, W. Huang, Z.-W. Liu, Y.-X. Wang and J.

Zhuang. Transcriptome-based analysis of Dof family transcription factors and their responses to abiotic stress in tea plant (Camellia sinensis). *International Journal of Genomics* 2016: (2016).

- J. Ma, M.-Y. Li, F. Wang, J. Tang and A.-S. Xiong. Genome-wide analysis of Dof family transcription factors and their responses to abiotic stresses in Chinese cabbage. *BMC Genomics* 16(1): 1-15 (2015).
- D. Golldack, I. Lüking and O. Yang. Plant tolerance to drought and salinity: stress regulating transcription factors and their functional significance in the cellular transcriptional network. *Plant cell reports* 30(8): 1383-1391 (2011).
- Z. Wu, J. Cheng, J. Cui, X. Xu, G. Liang, X. Luo, X. Chen, X. Tang, K. Hu and C. Qin. Genomewide identification and expression profile of Dof transcription factor gene family in pepper (Capsicum annuum L.). *Frontiers in Plant Science* 7: 574 (2016).
- Z.-J. Wu, X.-H. Li, Z.-W. Liu, H. Li, Y.-X. Wang and J. Zhuang. Transcriptome-based discovery of AP2/ ERF transcription factors related to temperature stress in tea plant (Camellia sinensis). *Functional & integrative genomics* 15(6): 741-752 (2015).
- Y. Huang, M.-Y. Li, F. Wang, Z.-S. Xu, W. Huang, G.-L. Wang, J. Ma and A.-S. Xiong. Heat shock factors in carrot: genome-wide identification, classification, and expression profiles response to abiotic stress. *Molecular biology reports* 42(5): 893-905 (2015).
- M. Noguero, R.M. Atif, S. Ochatt and R.D. Thompson. The role of the DNA-binding One Zinc Finger (DOF) transcription factor family in plants. *Plant Science* 209: 32-45 (2013).
- J. Venkatesh and S.W. Park. Genome-wide analysis and expression profiling of DNA-binding with one zinc finger (Dof) transcription factor family in potato. *Plant Physiology and Biochemistry* 94: 73-85 (2015).
- A. Skirycz, S. Jozefczuk, M. Stobiecki, D. Muth, M.I. Zanor, I. Witt and B. Mueller-Roeber. Transcription factor AtDOF4; 2 affects phenylpropanoid metabolism in Arabidopsis thaliana. *New Phytologist* 175(3): 425-438 (2007).
- C.-I. Wen, Q. Cheng, L. Zhao, A. Mao, J. Yang, S. Yu, Y. Weng and Y. Xu. Identification and characterisation of Dof transcription factors in the cucumber genome. *Scientific Reports* 6(1): 1-11 (2016).
- X. Cai, Y. Zhang, C. Zhang, T. Zhang, T. Hu, J. Ye, J. Zhang, T. Wang, H. Li, and Z. Ye. Genome-wide analysis of plant-specific Dof transcription factor family in tomato. *Journal of Integrative Plant Biology* 55(6): 552-566 (2013).
- 28. A. Skirycz, A. Radziejwoski, W. Busch, M.A.

Hannah, J. Czeszejko, M. Kwaśniewski, M.I. Zanor, J.U. Lohmann, L. De Veylder and I. Witt. The DOF transcription factor OBP1 is involved in cell cycle regulation in Arabidopsis thaliana. *The Plant Journal* 56(5): 779-792 (2008).

- A.R. Corrales, L. Carrillo, P. Lasierra, S.G. Nebauer, J. Dominguez-Figueroa, B. Renau-Morata, S. Pollmann, A. Granell, R.V. Molina and J. Vicente-Carbajosa. Multifaceted role of cycling DOF factor 3 (CDF3) in the regulation of flowering time and abiotic stress responses in Arabidopsis. *Plant, cell* & environment 40(5): 748-764 (2017).
- M. Ewas, E. Khames, K. Ziaf, R. Shahzad, E. Nishawy, F. Ali, H. Subthain, M.H. Amar, M. Ayaad and O. Ghaly. The tomato DOF daily fluctuations 1, TDDF1 acts as flowering accelerator and protector against various stresses. *Scientific Reports* 7(1): 1-16 (2017).
- 31. H. Qin, J. Wang, X. Chen, F. Wang, P. Peng, Y. Zhou, Y. Miao, Y. Zhang, Y. Gao, and Y. Qi. Rice Os DOF 15 contributes to ethylene-inhibited primary root elongation under salt stress. *New Phytologist* 223(2): 798-813 (2019).
- 32. Y. Zhou, Y. Cheng, C. Wan, J. Li, Y. Yang and J. Chen. Genome-wide characterization and expression analysis of the Dof gene family related to abiotic stress in watermelon. *PeerJ* 8: e8358 (2020).
- N. Suzuki, R.M. Rivero, V. Shulaev, E. Blumwald and R. Mittler. Abiotic and biotic stress combinations. *New Phytologist* 203(1): 32-43 (2014).
- L.N. Tolosa and Z. Zhang. The role of major transcription factors in solanaceous food crops under different stress conditions: current and future perspectives. *Plants* 9(1): 56 (2020).
- 35. J. Wu, L. Chen, M. Chen, W. Zhou, Q. Dong, H. Jiang and B. Cheng. The DOF-domain transcription factor ZmDOF36 positively regulates starch synthesis in transgenic maize. *Frontiers in Plant Science* 10: 465 (2019).
- L. Zhang, B. Liu, G. Zheng, A. Zhang, and R. Li. Genome-wide characterization of the SiDof gene family in foxtail millet (Setaria italica). *Biosystems* 151: 27-33 (2017).
- 37. G. Yang, L. Yu, Y. Wang, C. Wang and C. Gao. The translation initiation factor 1A (TheIF1A) from Tamarix hispida is regulated by a Dof transcription factor and increased abiotic stress tolerance. *Frontiers in plant science* 8: 513 (2017).
- 38. X. Cai, C. Zhang, W. Shu, Z. Ye, H. Li, and Y Zhang. The transcription factor SIDof22 involved in ascorbate accumulation and salinity stress in tomato. *Biochemical and Biophysical Research Communications* 474(4): 736-741 (2016).
- R. Shahzad, S. Jamil, S. Ahmad, A. Nisar, Z. Amina, S. Saleem, M.Z. Iqbal, R.M. Atif and X. Wang.

Harnessing the potential of plant transcription factors in developing climate resilient crops to improve global food security: Current and future perspectives. *Saudi Journal of Biological Sciences* 28(4): 2323-2341 (2021).

- 40. S.M. Azam, Y. Liu, Z.U. Rahman, H. Ali, C. Yan, L. Wang, S. Priyadarshani, B. Hu, X. Huang and J. Xiong. Identification, characterization and expression profiles of Dof transcription factors in pineapple (Ananas comosus L). *Tropical plant biology* 11(1): 49-64 (2018).
- 41. L.M. Shaw, C.L. McIntyre, P.M. Gresshoff, and G.P. Xue. Members of the Dof transcription factor family in Triticum aestivum are associated with lightmediated gene regulation. *Functional & integrative genomics* 9(4): 485-498 (2009).
- 42. Y. Liu, N. Liu, X. Deng, D. Liu, M. Li, D. Cui, Y. Hu and Y. Yan. Genome-wide analysis of wheat DNAbinding with one finger (Dof) transcription factor genes: evolutionary characteristics and diverse abiotic stress responses. *BMC genomics* 21(1): 1-18 (2020).
- 43. Y. Zhai, J. Chen, J. He, J. Zhang, W. Sha, H. Yu, Y. Zhao, T. Ma, T. Sun, and M. Zhang. Isolation, characterization and functional validation of a soybean transcription factor, GmDof4. 2 improves drought tolerance in transgenic tobacco. *Plant Cell*, *Tissue and Organ Culture (PCTOC)* 1-11 (2022).
- 44. W. Huang, Y. Huang, M.-y. Li, F. Wang, Z.-s. Xu and A.-s. Xiong. Dof transcription factors in carrot: genome-wide analysis and their response to abiotic stress. *Biotechnology Letters* 38(1): 145-155 (2016).
- 45. N. Gandass and P. Salvi. Intrinsically disordered protein, DNA binding with one finger transcription factor (OsDOF27) implicates thermotolerance in yeast and rice. *Frontiers in Plant Science* 13: (2022).
- 46. J. Xu and H. Dai. Brassica napus Cycling Dof Factor1 (BnCDF1) is involved in flowering time and freezing tolerance. *Plant growth regulation* 80(3): 315-322 (2016).
- 47. X. Cao, W. Wan, H. Mao, D. Yin, X. Deng, H. Yan and L. Ren. Genome-Wide Identification and Expression Analysis of Dof Transcription Factors in Lotus (Nelumbo nucifera Gaertn.). *Plants* 11(15): 2057 (2022).
- 48. A. Song, T. Gao, P. Li, S. Chen, Z. Guan, D. Wu, J. Xin, Q. Fan, K. Zhao, and F. Chen. Transcriptome-wide identification and expression profiling of the DOF transcription factor gene family in Chrysanthemum morifolium. *Frontiers in plant science* 7: 199 (2016).
- 49. T. Luo, Y. Song, H. Gao, M. Wang, H. Cui, C. Ji, J. Wang, L. Yuan, and R. Li. Genome-wide identification and functional analysis of Dof transcription factor family in Camelina sativa. BMC

genomics 23(1): 1-17 (2022).

- L. Zheng, G. Liu, X. Meng, Y. Liu, X. Ji, Y. Li, X. Nie and Y. Wang. A WRKY gene from Tamarix hispida, ThWRKY4, mediates abiotic stress responses by modulating reactive oxygen species and expression of stress-responsive genes. *Plant molecular biology* 82(4): 303-320 (2013).
- Y. Chen and J. Cao. Comparative analysis of Dof transcription factor family in maize. *Plant Molecular Biology Reporter* 33(5): 1245-1258 (2015).
- 52. H. Wang, S. Zhao, Y. Gao, and J. Yang. Characterization of Dof transcription factors and their responses to osmotic stress in poplar (Populus trichocarpa). *PLoS One* 12(1): e0170210 (2017).
- 53. Y. Su, W. Liang, Z. Liu, Y. Wang, Y. Zhao, B. Ijaz and J. Hua. Overexpression of GhDof1 improved salt and cold tolerance and seed oil content in Gossypium hirsutum. *Journal of Plant Physiology* 218: 222-234 (2017).
- 54. Q. Wei, W. Wang, T. Hu, H. Hu, W. Mao, Q. Zhu and C. Bao. Genome-wide identification and characterization of Dof transcription factors in eggplant (Solanum melongena L.). *PeerJ* 6: e4481 (2018).
- 55. T. Li, X. Wang, D. Elango, W. Zhang, M. Li, F. Zhang, Q. Pan, and Y. Wu. Genome-wide identification, phylogenetic and expression pattern analysis of Dof transcription factors in blueberry (Vaccinium corymbosum L.). *PeerJ* 10: e14087 (2022).
- 56. H. Nan, R.A. Ludlow, M. Lu, and H. An. Genomewide analysis of Dof genes and their response to abiotic stress in rose (Rosa chinensis). *Frontiers in Genetics* 12: 538733 (2021).
- 57. T. Guo, S. Wang, T. Zhang, L. Xu, Y. Li, Y. Chao, and L. Han. Expression of the Medicago truncatula MtDof32 transcription factor regulates plant growth and enhances abiotic stress tolerances in transgenic Arabidopsis. *Environmental and Experimental Botany* 183: 104339 (2021).
- A. AghaKouchak, A. Farahmand, F. Melton, J. Teixeira, M. Anderson, B.D. Wardlow and C. Hain. Remote sensing of drought: Progress, challenges and opportunities. *Reviews of Geophysics* 53(2): 452-480 (2015).
- 59. G. Li, W. Xu, P. Jing, X. Hou, and X. Fan. Overexpression of VyDOF8, a Chinese wild grapevine transcription factor gene, enhances drought tolerance in transgenic tobacco. *Environmental and Experimental Botany* 190: 104592 (2021).
- 60. A.R. Corrales Ducuara, S.G. Nebauer, L. Carrillo Gil, P. Fernandez Nohales, J. Marqués, B. Renau Morata, A. Granel, S. Pollmann, J. Vicente Carbajosa and R.V. Molina. Characterization of tomato Cycling Dof Factors reveals conserved and new functions in the control of flowering time and

abiotic stress responses. *Journal of Experimental Botany* 65(4): 995-1012 (2013).

- Z. Fang, W. Jiang, Y. He, D. Ma, Y. Liu, S. Wang, Y. Zhang, and J. Yin. Genome-wide identification, structure characterization, and expression profiling of Dof transcription factor gene family in wheat (Triticum aestivum L.). *Agronomy* 10(2): 294 (2020).
- 62. A. Raza, J. Tabassum, A.Z. Fakhar, R. Sharif, H. Chen, C. Zhang, L. Ju, V. Fotopoulos, K.H. Siddique, and R.K. Singh. Smart reprograming of plants against salinity stress using modern biotechnological tools. *Critical Reviews in Biotechnology* 1-28 (2022).
- T. Zhang and Y. Zhou. Plant transcription factors and salt stress, Plant Transcription Factors. *Elsevier*, pp. 369-381 (2023).
- 64. L. Cheng, S. Li, J. Hussain, X. Xu, J. Yin, Y. Zhang, X. Chen, and L. Li. Isolation and functional characterization of a salt responsive transcriptional factor, LrbZIP from lotus root (Nelumbo nucifera Gaertn). *Molecular biology reports* 40(6): 4033-4045 (2013).
- 65. A.-R. Corrales, S.G. Nebauer, L. Carrillo, P. Fernández-Nohales, J. Marqués, B. Renau-Morata, A. Granell, S. Pollmann, J. Vicente-Carbajosa and R.-V. Molina. Characterization of tomato Cycling Dof Factors reveals conserved and new functions in the control of flowering time and abiotic stress responses. *Journal of experimental botany* 65(4): 995-1012 (2014).
- 66. Q.Y. Zhou, A.G. Tian, H.F. Zou, Z.M. Xie, G. Lei, J. Huang, C.M. Wang, H.W. Wang, J.S. Zhang, and S.Y. Chen. Soybean WRKY-type transcription factor genes, GmWRKY13, GmWRKY21, and GmWRKY54, confer differential tolerance to abiotic stresses in transgenic Arabidopsis plants. *Plant biotechnology journal* 6(5): 486-503 (2008).
- T. Finatto, V.E. Viana, L.G. Woyann, C. Busanello, L.C.d. Maia and A.C.d. Oliveira. Can WRKY transcription factors help plants to overcome environmental challenges?, *Genetics and molecular biology* 41: 533-544 (2018).
- P. Albertos, G. Dündar, P. Schenk, S. Carrera, P. Cavelius, T. Sieberer and B. Poppenberger. Transcription factor BES1 interacts with HSFA1 to promote heat stress resistance of plants. *The EMBO journal* 41(3): e108664 (2022).
- N.-H. Ghori, T. Ghori, M. Hayat, S. Imadi, A. Gul, V. Altay and M. Ozturk. Heavy metal stress and responses in plants. *International journal of environmental science and technology* 16(3): 1807-1828 (2019).
- 70. S. Kumar, S.H. Shah, Y. Vimala, H.S. Jatav, P. Ahmad, Y. Chen and K.H. Siddique. Abscisic acid: Metabolism, transport, crosstalk with other plant growth regulators, and its role in heavy metal stress

mitigation. *Frontiers in Plant Science* 13: 972856 (2022).

- J. Lü, M. Yang, Q. Meng, K. Zhuang and N. Ma. Chloroplast metalloproteinase SIL2 reduces the thermotolerance of tomato by decreasing the content of SICDJ1. *Protoplasma* 1-13 (2023).
- V. Martinez, M. Nieves-Cordones, M. Lopez-Delacalle, R. Rodenas, T.C. Mestre, F. Garcia-Sanchez, F. Rubio, P.A. Nortes, R. Mittler and R.M. Rivero. Tolerance to stress combination in tomato plants: New insights in the protective role of melatonin. *Molecules* 23(3): 535 (2018).
- 73. Y. Huang, J. An, S. Sircar, C. Bergis, C.D. Lopes, X. He, B. Da Costa, F.-Q. Tan, J. Bazin and J. Antunez-Sanchez. HSFA1a modulates plant heat stress responses and alters the 3D chromatin organization of enhancer-promoter interactions. *Nature Communications* 14(1): 469 (2023).
- 74. N. Lohani, A.A. Golicz, M.B. Singh, and P.L. Bhalla. Genome-wide analysis of the Hsf gene family in Brassica oleracea and a comparative analysis of the Hsf gene family in B. oleracea, B. rapa and B. napus. *Functional & integrative genomics* 19(3): 515-531 (2019).
- 75. G. Yang, X. Gao, K. Ma, D. Li, C. Jia, M. Zhai and Z. Xu. The walnut transcription factor JrGRAS2 contributes to high temperature stress tolerance involving in Dof transcriptional regulation and HSP protein expression. *BMC plant biology* 18(1): 1-14 (2018).
- A. Arenas-M, F.M. Castillo, D. Godoy, J. Canales and D.F. Calderini. Transcriptomic and Physiological Response of Durum Wheat Grain to Short-Term Heat Stress during Early Grain Filling. *Plants* 11(1): 59 (2021).
- 77. M. Kim, V. Schulz, L. Brings, T. Schoeller, K. Kühn and E. Vierling. mTERF18 and ATAD3 are required for mitochondrial nucleoid structure and their disruption confers heat tolerance in Arabidopsis thaliana. *New Phytologist* 232(5): 2026-2042 (2021).
- L. Wobbe. The molecular function of plant mTERFs as key regulators of organellar gene expression. *Plant and Cell Physiology* 61(12): 2004-2017

(2020).

- 79. P.K. Papolu, M. Ramakrishnan, Q. Wei, K.K. Vinod, L.-H. Zou, K. Yrjala, R. Kalendar and M. Zhou. Long terminal repeats (LTR) and transcription factors regulate PHRE1 and PHRE2 activity in Moso bamboo under heat stress. *BMC Plant Biology* 21(1): 1-19 (2021).
- G. He, W. Tian, L. Qin, L. Meng, D. Wu, Y. Huang, D. Li, D. Zhao and T. He. Identification of novel heavy metal detoxification proteins in Solanum tuberosum: Insights to improve food security protection from metal ion stress. *Science of The Total Environment* 779: 146197 (2021).
- 81. J. Li, M. Zhang, J. Sun, X. Mao, J. Wang, H. Liu, H. Zheng, X. Li, H. Zhao and D. Zou. Heavy metal stress-associated proteins in rice and Arabidopsis: genome-wide identification, phylogenetics, duplication, and expression profiles analysis. *Frontiers in Genetics* 11: 477 (2020).
- 82. C. Huo, L. He, T. Yu, X. Ji, R. Li, S. Zhu, F. Zhang, H. Xie and W. Liu. The Superoxide Dismutase Gene Family in Nicotiana tabacum: Genome-Wide Identification, Characterization, Expression Profiling and Functional Analysis in Response to Heavy Metal Stress. *Frontiers in Plant Science* 13 (2022).
- D. Hwarari, Y. Guan, B. Ahmad, A. Movahedi, T. Min, Z. Hao, Y. Lu, J. Chen and L. Yang. ICE-CBF-COR signaling cascade and its regulation in plants responding to cold stress. *International Journal of Molecular Sciences* 23(3): 1549 (2022).
- 84. F.N. Ritonga and S. Chen. Physiological and molecular mechanism involved in cold stress tolerance in plants. *Plants* 9(5): (2020) 560.
- 85. M. Manna, T. Thakur, O. Chirom, R. Mandlik, R. Deshmukh and P. Salvi. Transcription factors as key molecular target to strengthen the drought stress tolerance in plants. *Physiologia Plantarum* 172(2): 847-868 (2021).
- 86. G.K. Rai, G. Jamwal, G. Rai and M. Singh. Understanding Transcription Factors in Plant Response to Drought Stress. *Indian Journal of Agricultural Biochemistry* 34(2): 116-125 (2021).